



Clinical Genomics

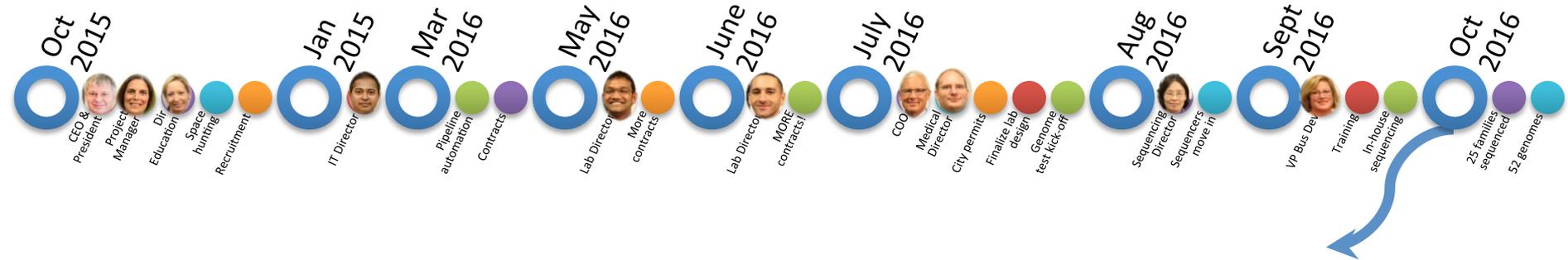
Narayanan "Ray" Veeraraghavan, PhD

Director of Informatics

Rady Children's Institute for Genomic Medicine, San Diego, CA
USA

Rady Children's Institute

Genomic Medicine



Outline

- WHY genomics in clinic?
- WHAT is Clinical Genomics
 - Steps involved in Clinical Genomics
 - Ever increasing bioinformatics tool set
- HOW to be a good clinical bioinformaticist
 - Skills to develop
 - Compliance
 - Resources

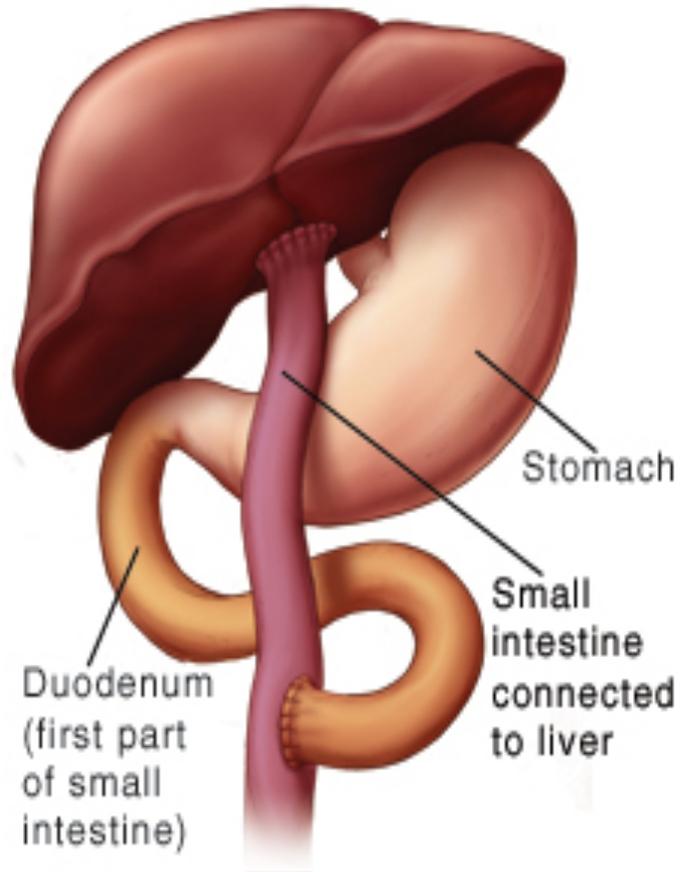
Let us begin with a story

A newborn baby is lying in a hospital bed in a Neonatal Intensive Care Unit (NICU). The baby is wearing a white blanket and has several medical sensors attached to their chest and limbs. The room is filled with medical equipment, including multiple infusion pumps (some labeled 'RADI CHILDRENS') and monitors. The text 'Baby admitted to regional Level 4 NICU' is overlaid on the image in a white box with a blue border.

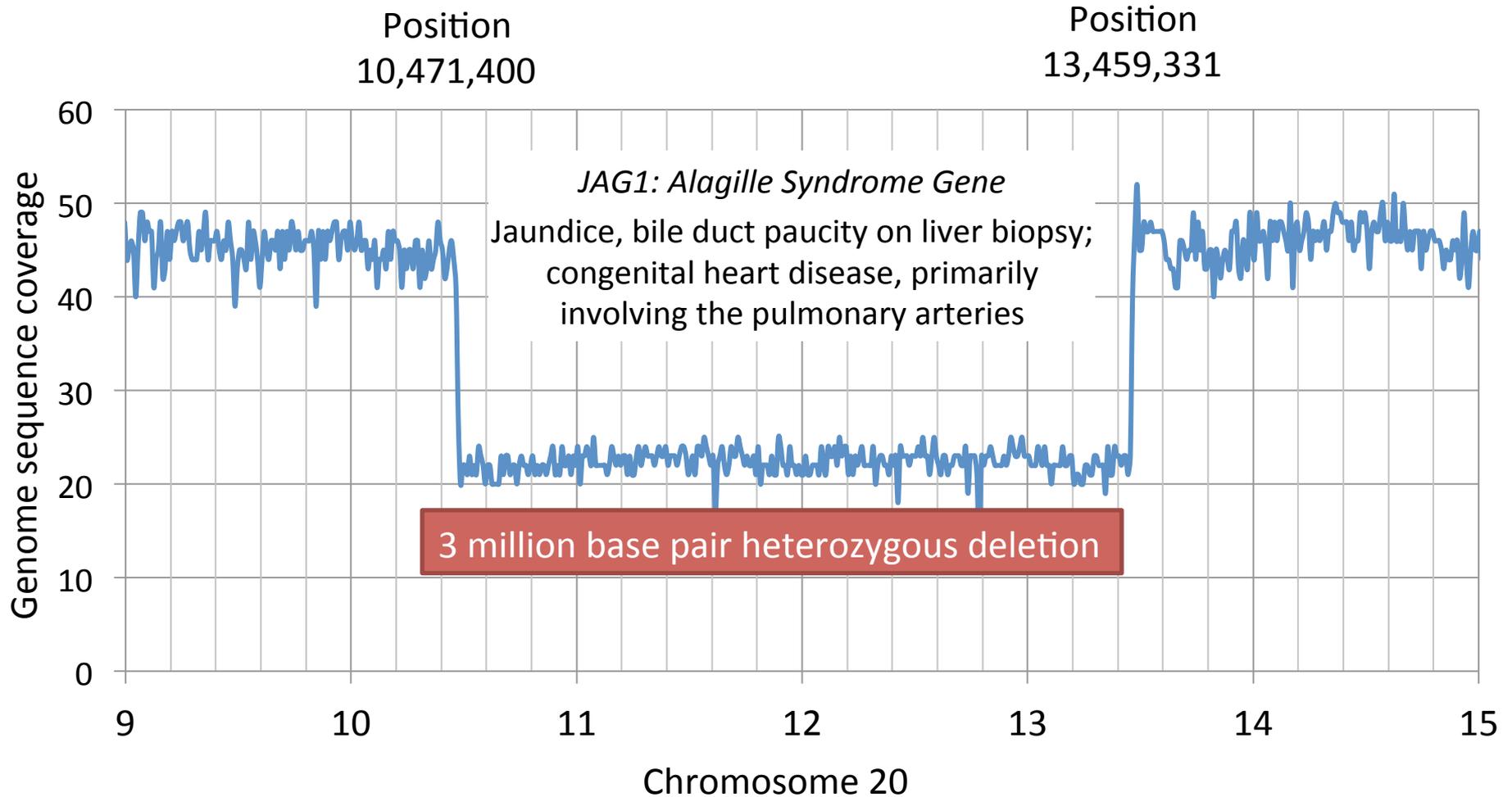
**Baby admitted
to regional
Level 4 NICU**

Rady Children's Hospital Baby 6026

- 2 month old admitted to PICU with severe jaundice & poor weight gain for 1 month
- Weight 0.05%^{ile}
- Acidotic, tachypneic
- Echo: Congenital heart disease, underdeveloped pulmonary arteries
- Clinical diagnosis: biliary atresia
 - Incidence 1 in 10,000
- Empiric treatment: Kasai procedure
 - Prognosis worsens with time to surgery
- Liver biopsy not diagnostic: giant cell hepatitis, sample too small to count bile ducts



43 Hours Later: Provisional Diagnosis



Clinical Impact & Outcome

- Kasai procedure and intraoperative cholangiogram scheduled at 11:00 am
- Genetic diagnosis communicated to clinical team just before operation – procedure cancelled
- Studies have shown kids with Alagille Syndrome w/o Kasai have 50% decrease in mortality

“Infants with Alagille syndrome are occasionally misdiagnosed as biliary atresia and subsequently undergo Kasai operation during infancy.

*Among 15 children with Alagille syndrome, mortality was 60% among the Kasai group, and 10% among the non-Kasai group. Liver transplantation was performed in 100% of the Kasai group, and **20% of the non-Kasai group.**”*

On an Average Day in the US...



10,926

Births



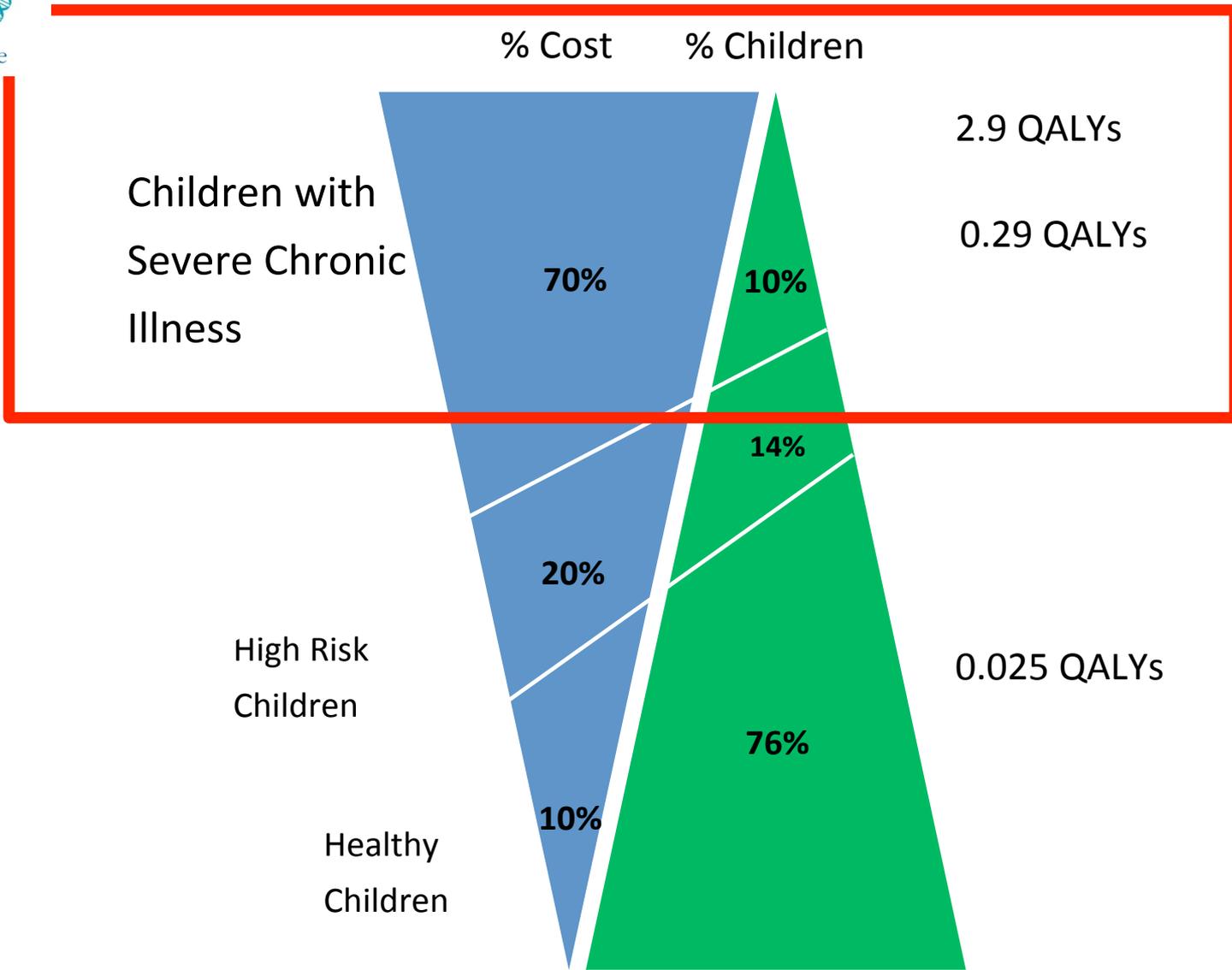
437

Babies born with 1 of
8,000 genetic diseases



64

Babies die before
1st birthday



On an Average Day...

Baby born in
community hospital
Acutely ill
Transferred to
regional NICU

4%

Interim empirical
treatment

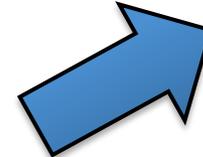
Discharged home
Palliative care
Death

Search for
etiologic
diagnosis

60 days

Improvement or
worsening

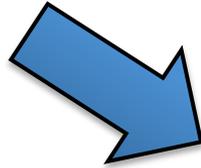
Modify
treatment



...Rady Children's Hospital – San Diego

Baby born in
community hospital
Acutely ill
Transferred to
regional NICU

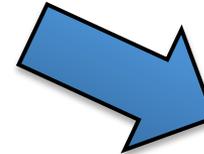
4%



Rapid Whole
Genome
Sequencing



Rapid
Precision
Medicine



2 days



24% improved outcome
75% shorter hospital stay
85% decrease in cost

Based off first 42 babies seen

40% rapid
diagnoses



76%
precision
medicine

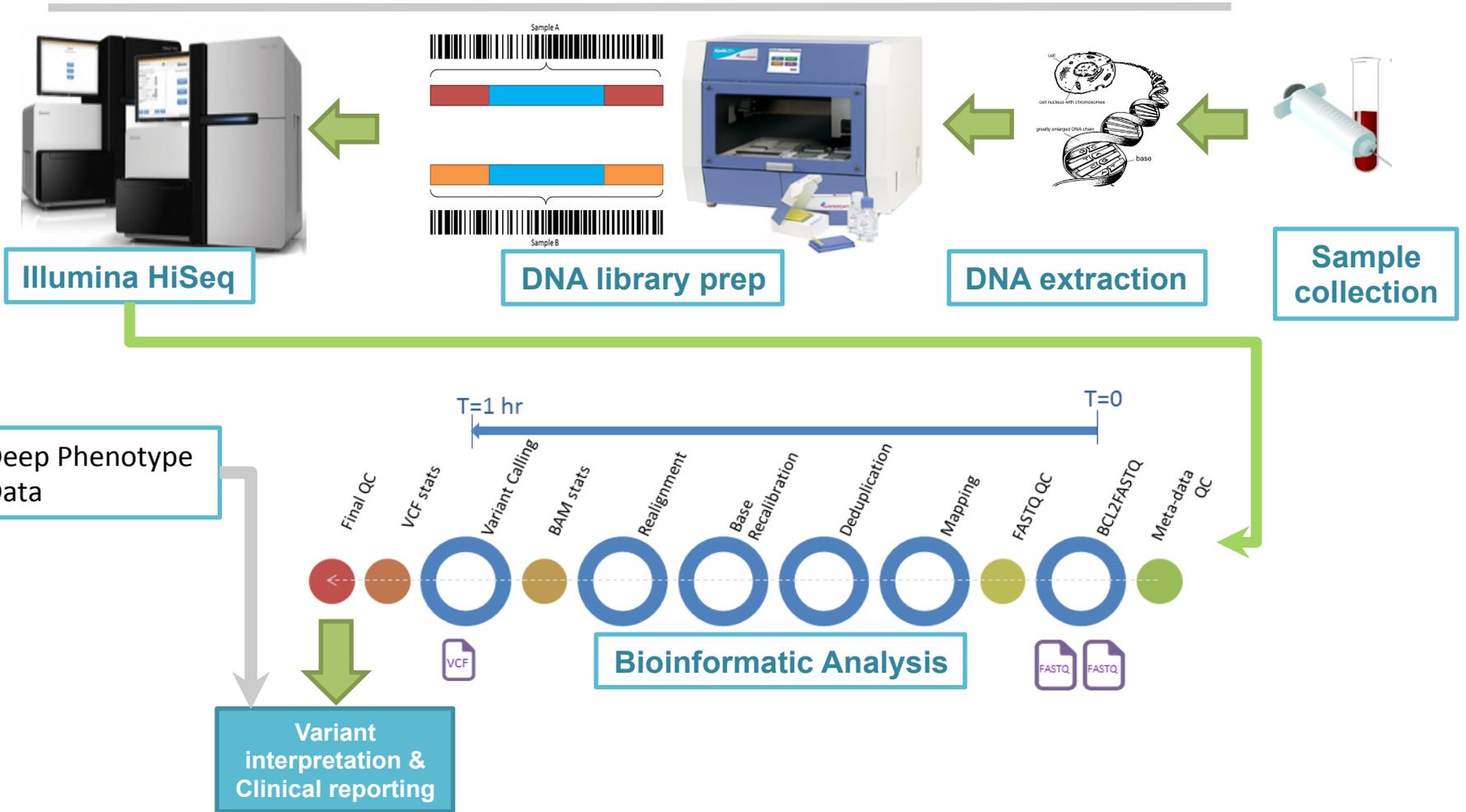


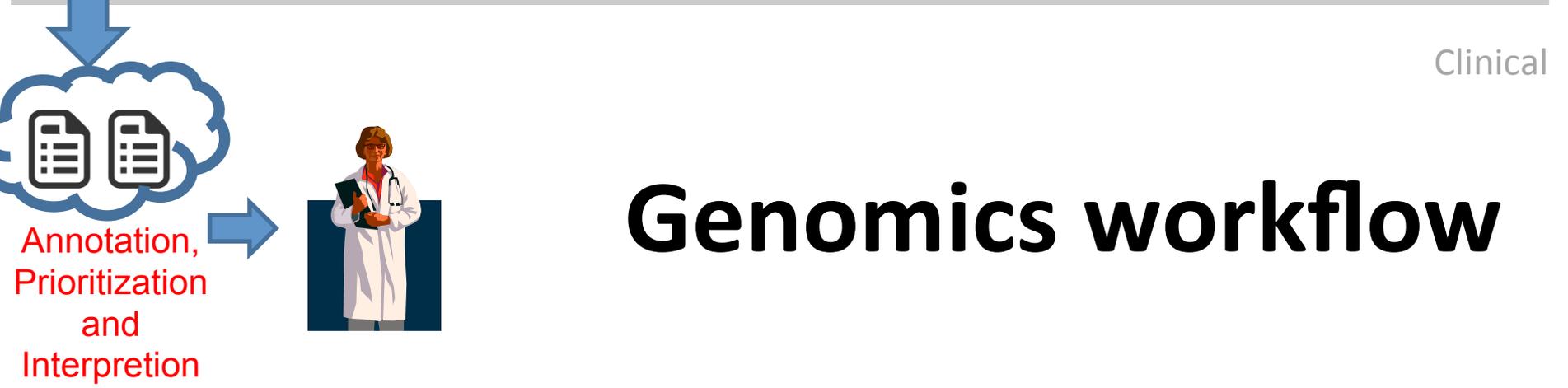
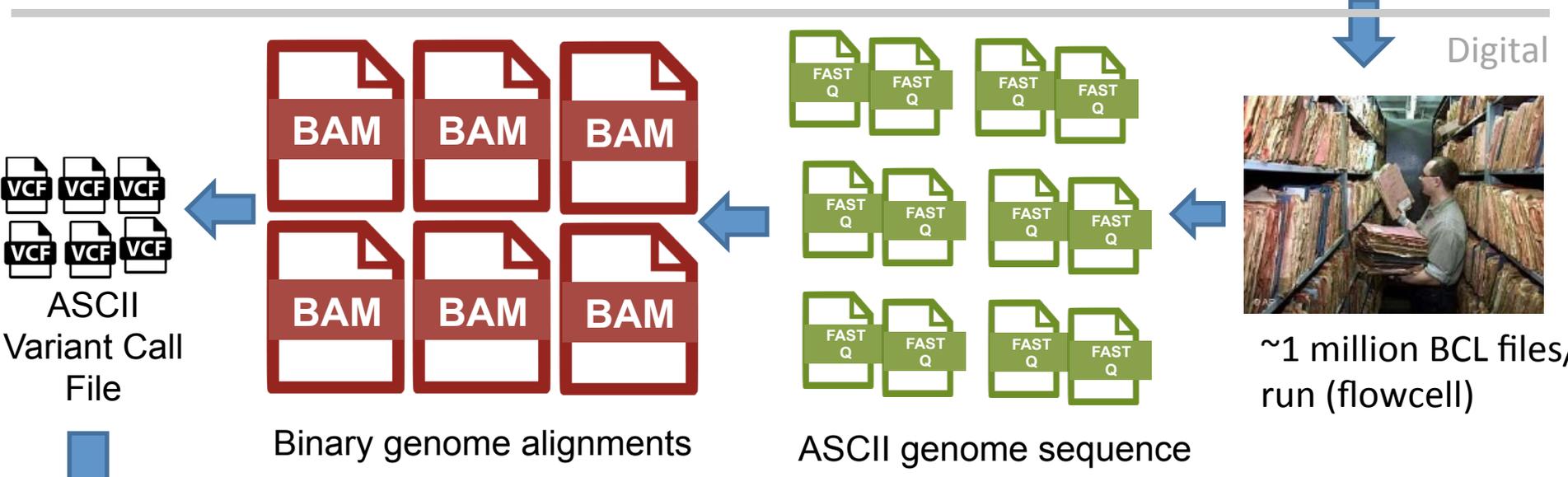
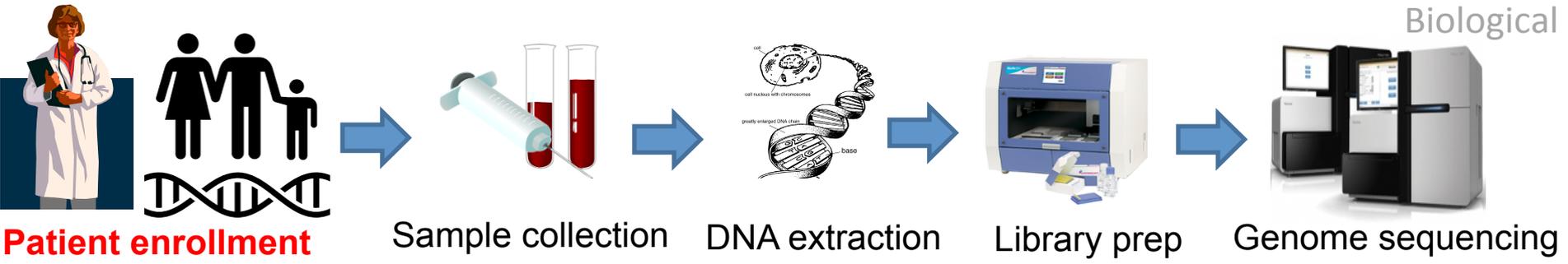
77%
improved
outcome



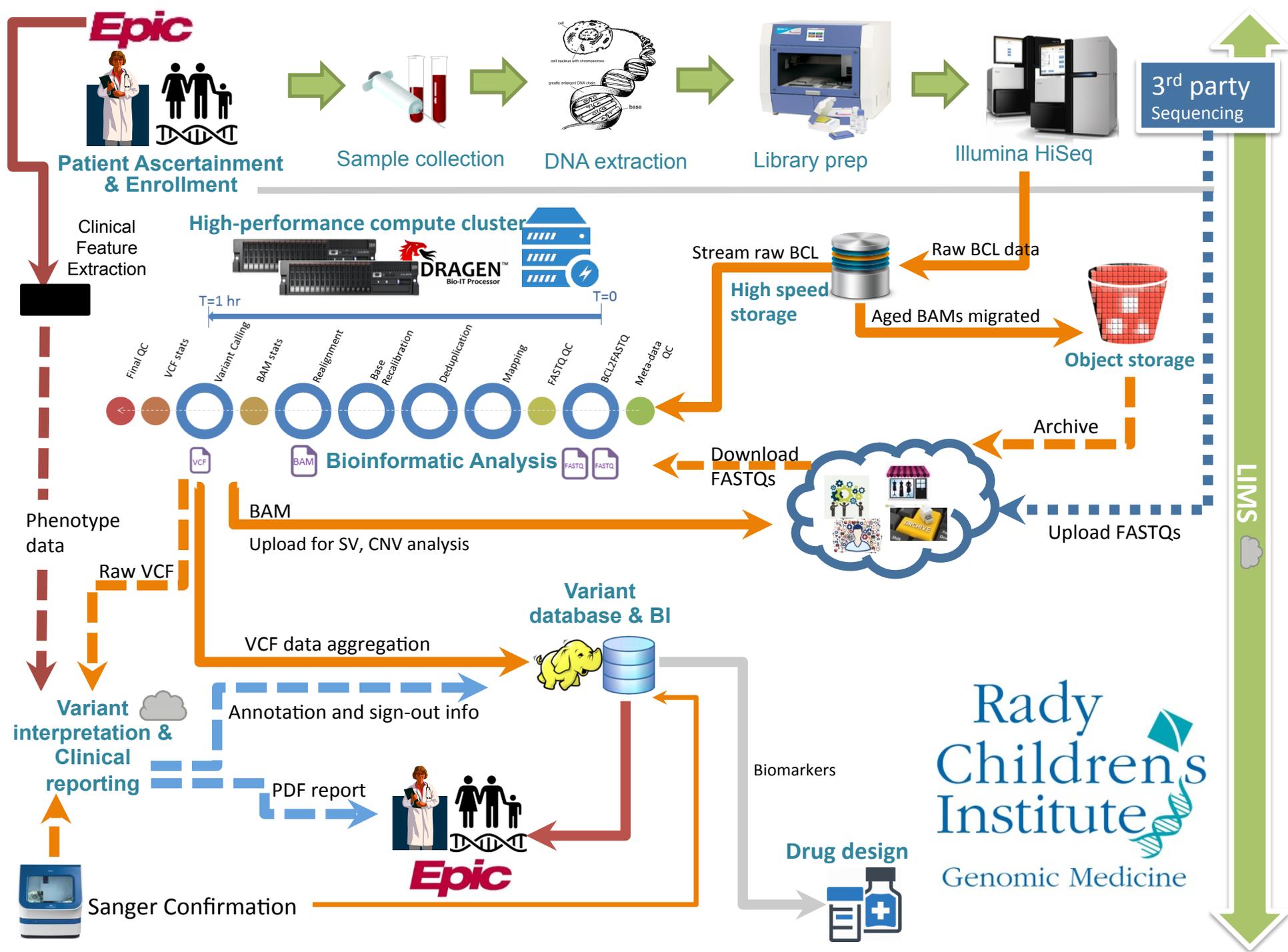
75% reduction in healthcare utilization
85% reduction in cost

"Core" genomics workflow





Genomics workflow



p. 26

A blood test that can detect cancer could change the way we treat the disease

p. 30

Researchers are rethinking clinical trials, and that's a good thing

p. 48

Wearable tech: Are you ready for this new era of medicine?

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YOUR HEALTH IS PERSONAL SUMMER & FALL 2014

Genome



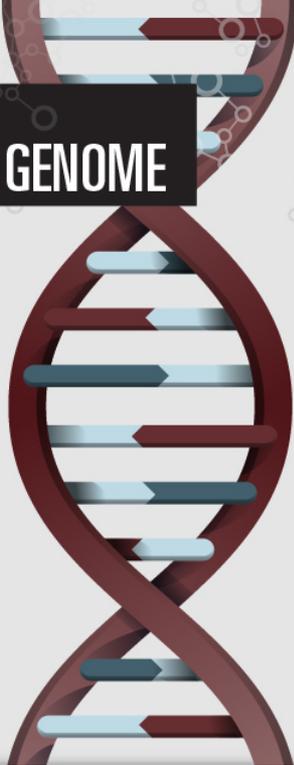
THE FAMILY CONDITION

HOW THE GENES YOU INHERIT AFFECT YOUR RISK FOR DISEASE

A GENOMIC REVOLUTION:
THE MAPPING OF THE HUMAN GENOME

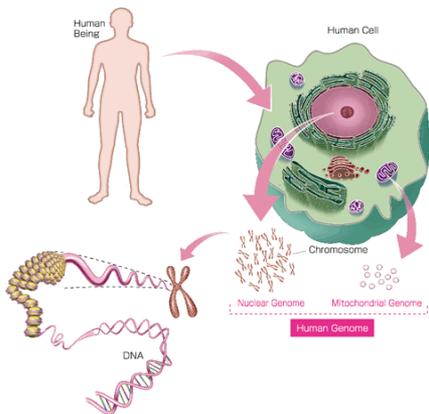
THE SEQUENCING OF THE HUMAN GENOME WAS
THE LARGEST
SINGLE UNDERTAKING
 IN THE HISTORY OF BIOLOGICAL SCIENCE.

1990 IT TOOK **13 YEARS** 2003
TO SEQUENCE 3 BILLION
 DNA BASE PAIRS AND IDENTIFY
 EACH HUMAN GENE.



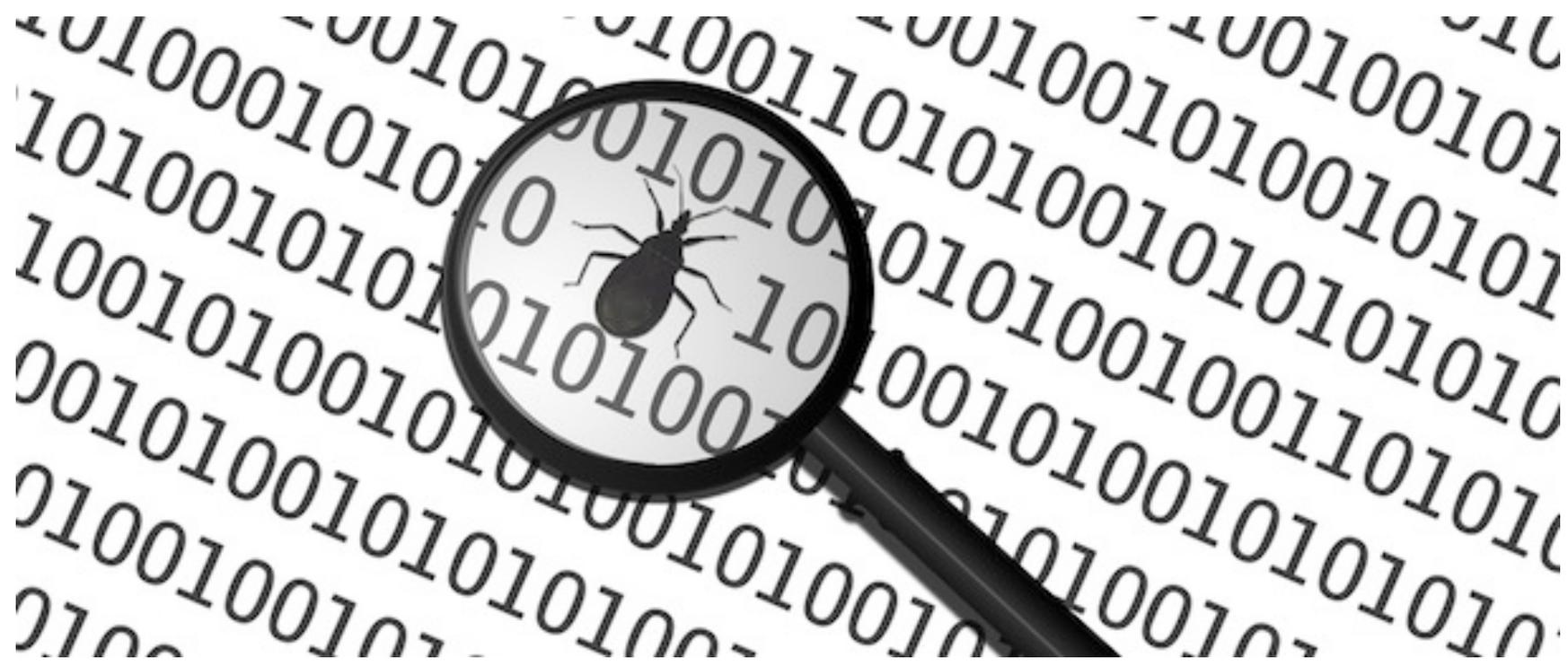
Only 10 years after the human genome was first mapped,
THE IMPACT IS ALREADY SIGNIFICANT AND WIDESPREAD.

2013

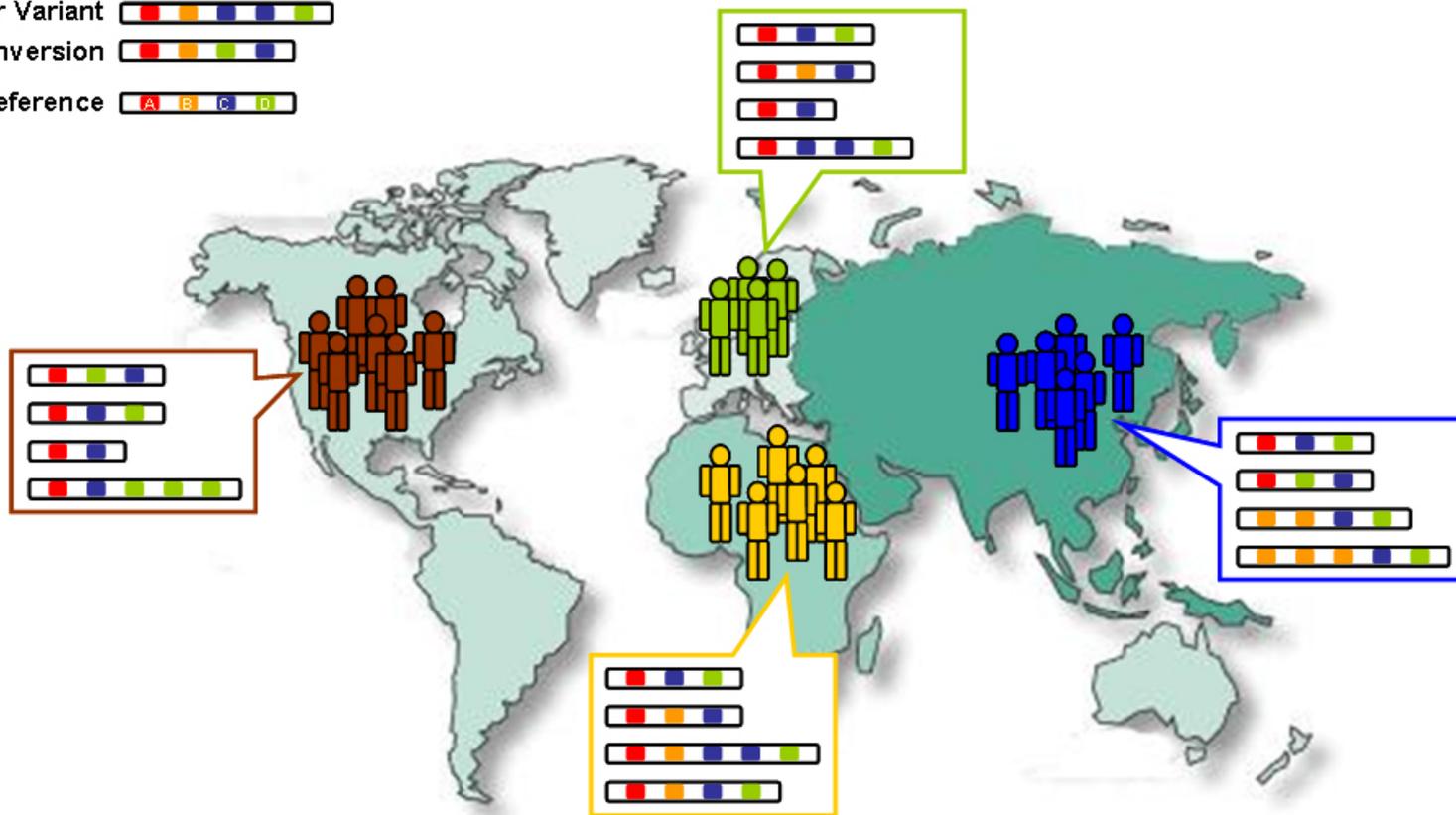


© ZOOM





- Insertion
- Deletion
- Copy Number Variant
- Inversion
- Reference



Kinds of data analytics

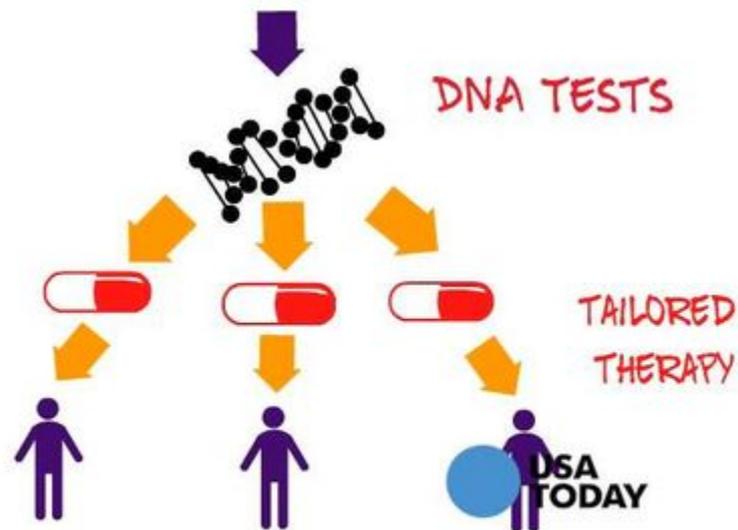
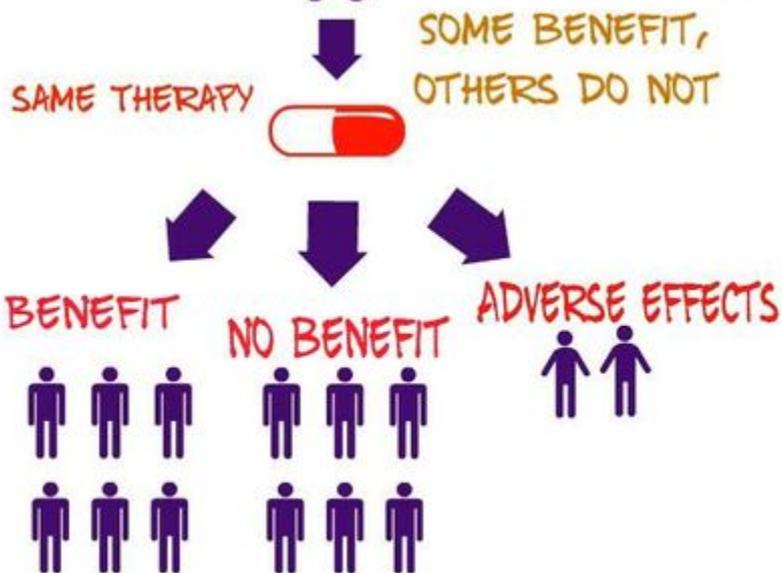
- Single patient level:
 - what genetic variation could be causative of XYZ clinical presentations
- Population level:
 - How frequent is variation XYZ in Hispanic population
 - What kinds of genetic variation do we typically observe for patients who have XYZ types of clinical presentations
 - Give me all patients with ABC phenotype and PQR genotype (genetic variation)
 - What genetic variation co-occur in LMN disease

WITHOUT
PRECISION MEDICINE

WITH
PRECISION MEDICINE

PATIENT

EACH PATIENT BENEFITS



Alexis and Noah, born 1996

Colic ???

Twins wrongly diagnosed with cerebral palsy at age 2

2 yrs later, after multiple interventions

Low dopamine levels
dopa-responsive dystonia



Twins sleep through the night for the first time in their lives!

9 yrs

2009,
Cough +
breathing
problems



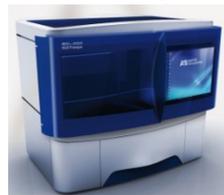
Beery twins, Age 14, 2010 @ Baylor College of Medicine, Houston, TX

<https://www.youtube.com/watch?v=yUQFHecs8EQ>



Dr. Jennifer Friedman

2011, Sequencing



Mutations in a gene called *SPR* that encodes the enzyme sepiapterin reductase, which was previously linked to some cases of dopa-responsive dystonia

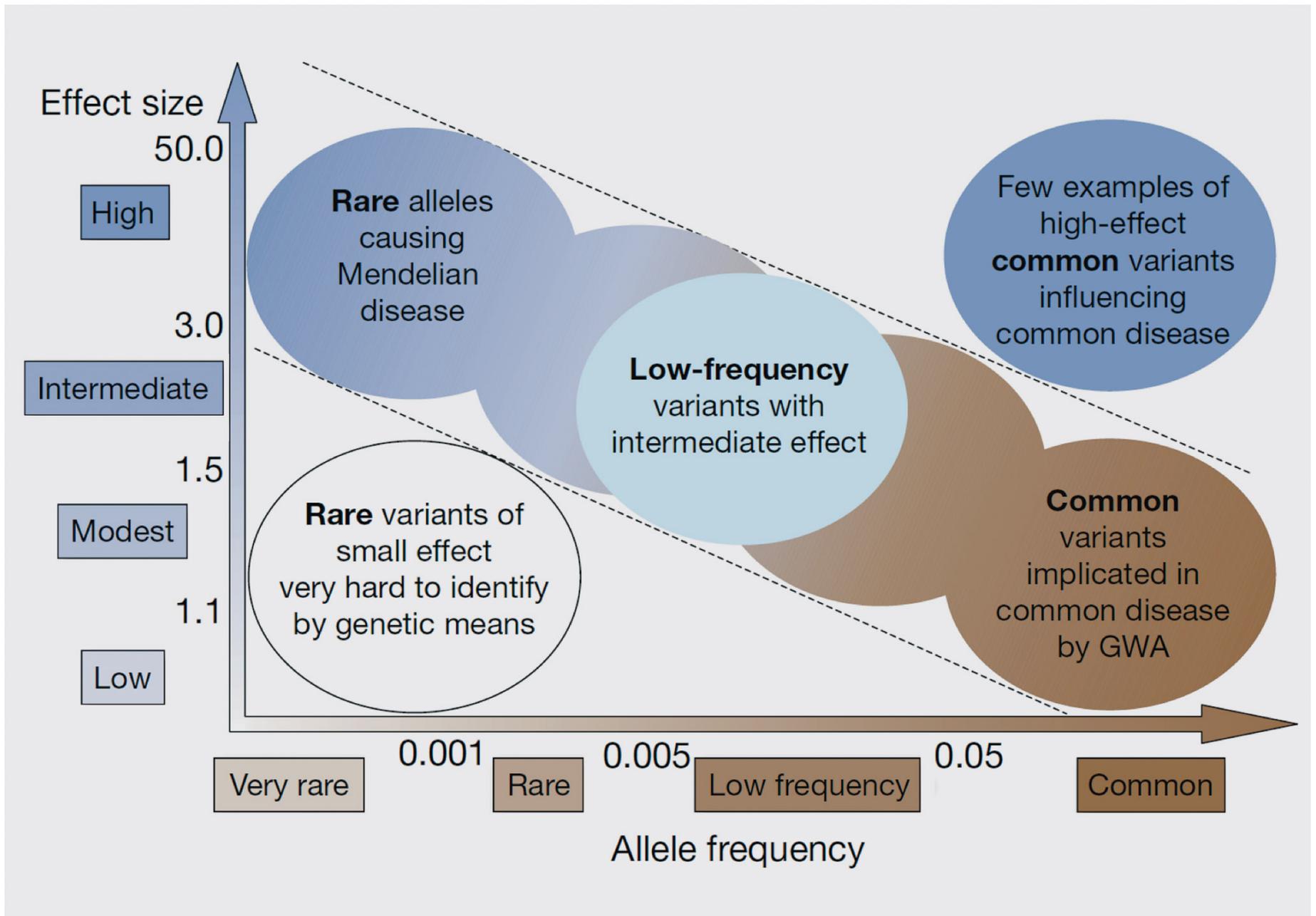
affect not only dopamine but also *serotonin* production

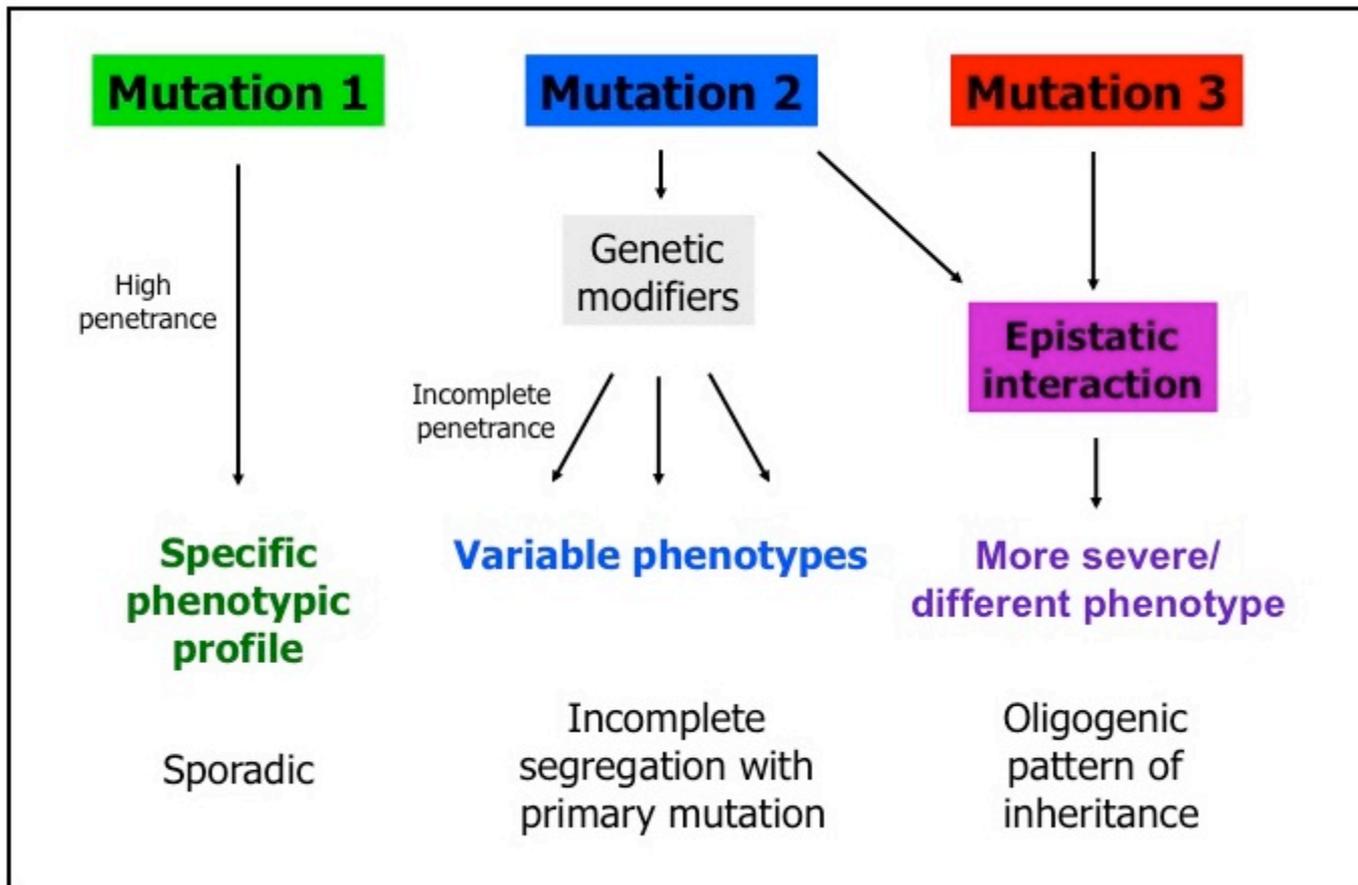


Usual suspects



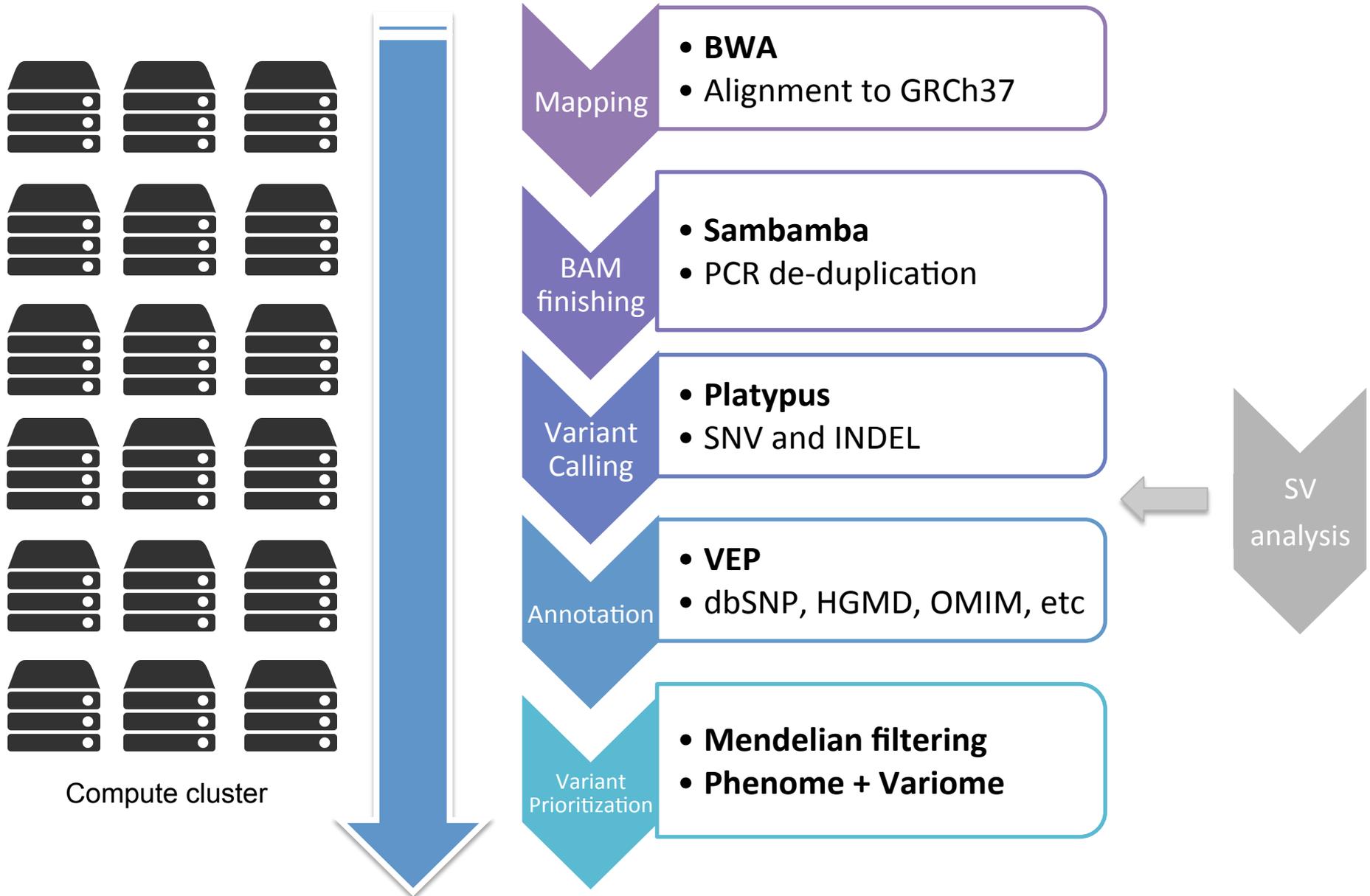
Rare Variants





Common Bioinformatics Steps

Bioinformatics Pipeline



Common Bioinformatics Steps

- Note
 - We will keep things simple
 - Many nuances and side paths, but lets not get distracted

 - We will not get into algorithms or underlying math
 - We will not be afraid of the command line

Mapping



Many mapping tools out there....

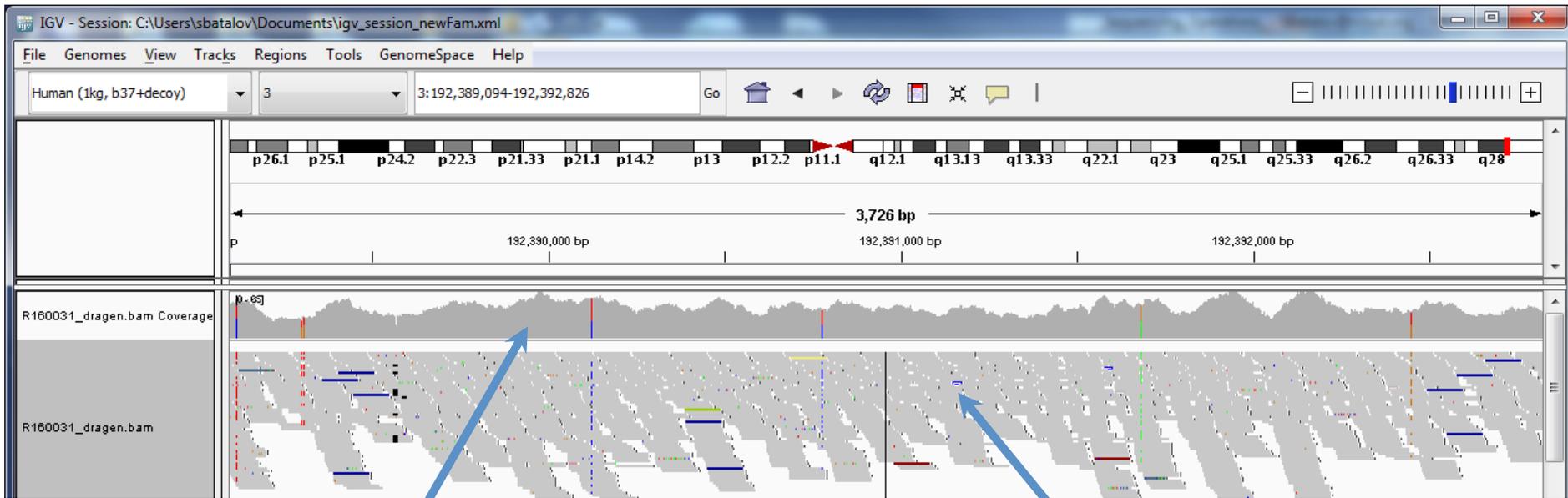
- Bfast
- BioScope
- Bowtie
- BWA
- CLC bio
- CloudBurst
- Eland/Eland2
- GenomeMapper
- GnuMap
- Karma
- MAQ
- MOM
- Mosaik
- MrFAST/MrsFAST
- NovoAlign
- PASS
- PerM
- Phaster
- RazerS
- RMAP
- SSAHA2
- Segemehl
- SeqMap
- SHRiMP
- Slider/SliderII
- SOAP/SOAP2
- Srprism
- Stampy
- vmatch
- ZOOM
-

Here's what a BAM really looks like

```
@HD      VN:1.0  SO:coordinate
@SQ      SN:chr20      LN:64444167
@PG      ID:TopHat     VN:2.0.14      CL:/srv/dna_tools/tophat/tophat -N 3 --read-edit-dist 5 --read-rea
lign-edit-dist 2 -i 50 -I 5000 --max-coverage-intron 5000 -M -o out /data/user446/mapping_tophat/index/chr
20 /data/user446/mapping_tophat/L6_18_GTGAAA_L007_R1_001.fastq
HWI-ST1145:74:C101DACXX:7:1102:4284:73714      16      chr20      190930      3      100M      *      0      0
      CCGTGTTTAAAGGTGGATGCGGTCACCTTCCCAGCTAGGCTTAGGGATTCTTAGTTGGCCTAGGAAATCCAGCTAGTCTGTCTCTCAGTCCCCCTCT
C      BBDCCDDCCDDDDCCDDDDDDCCDCBC?DDDDDDDDDDDDDDCCDCDDDDDDDDDDCCCEDDDC?DDDDDDDDDDDDDDDDDDDDDDBDHFFFFDC@@
      AS:i:-15      XM:i:3      XO:i:0      XG:i:0      MD:Z:55C20C13A9      NM:i:3      NH:i:2      CC:Z:=      CP:i:55352714      HI:i:0
HWI-ST1145:74:C101DACXX:7:1114:2759:41961      16      chr20      193953      50      100M      *      0      0
      TGCTGGATCATCTGGTTAGTGGCTTCTGACTCAGAGGACCTTCGTCCCCTGGGGCAGTGGACCTTCCAGTGATTCCCCTGACATAAGGGGCATGGACGA
G      DCDDDEDDDDDDDDCCDDDDDDCCDDDDCCDDDDDEEC>DFFFEJJJJJJIGJJJJJIHGBHHGJIIJJJJJJGJJIIJJJJIIHJJJJJJHHHHHFFFFFCCC
      AS:i:-16      XM:i:3      XO:i:0      XG:i:0      MD:Z:60G16T18T3      NM:i:3      NH:i:1
HWI-ST1145:74:C101DACXX:7:1204:14760:4030      16      chr20      270877      50      100M      *      0      0
      GGCTTTATTGGTAAAAAAGGAATAGCAGATTTAATCAGAAATTCCCACCTGGCCCAGCAGCACCAACCAGAAAGAAGGAAGAAGACAGGAAAAACCA
C      DDDDDDDDDCCDDDDDDDDDEEEEEEEFFFEFFEGHHHFGDJJIHJJIIJJIIIIIGGFJJIIHIIIIJJJJJJIGHHFAHGFHJHFGGHFFFD@BB
      AS:i:-11      XM:i:2      XO:i:0      XG:i:0      MD:Z:0A85G13      NM:i:2      NH:i:1
HWI-ST1145:74:C101DACXX:7:1210:11167:8699      0      chr20      271218      50      50M4700N50M      *      0
      0      GTGGCTCTCCACAGGAATGTTGAGGATGACATCCATGTCTGGGGTGCACCTGGGTCTCCGAAGCAGAACATCCTCAAATATGACCTCTCG
```

accepted_hits.sam

Here's how a BAM can be better visualized



Coverage track

Reads pile up

A survey of sequence alignment algorithms for next-generation sequencing

Heng Li and Nils Homer

Submitted: 3rd March 2010; Received (in revised form): 14th April 2010

Abstract

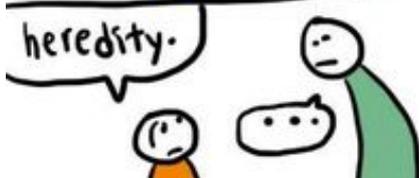
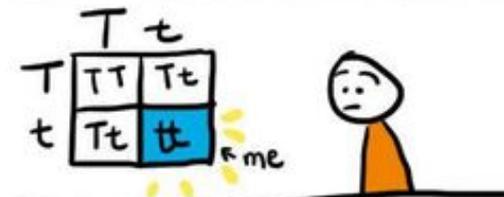
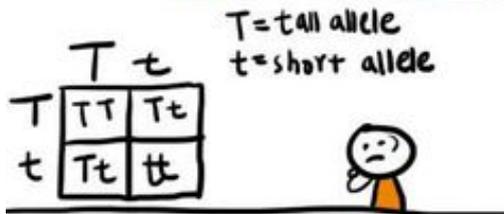
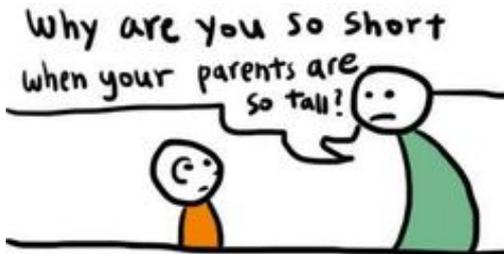
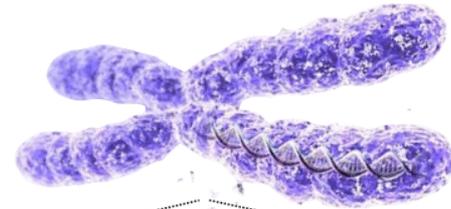
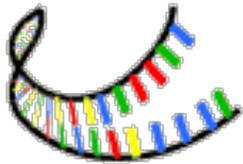
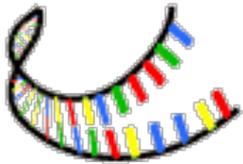
Rapidly evolving sequencing technologies produce data on an unparalleled scale. A central challenge to the analysis of this data is sequence alignment, whereby sequence reads must be compared to a reference. A wide variety of alignment algorithms and software have been subsequently developed over the past two years. In this article, we will systematically review the current development of these algorithms and introduce their practical applications on different types of experimental data. We come to the conclusion that short-read alignment is no longer the bottleneck of data analyses. We also consider future development of alignment algorithms with respect to emerging long sequence reads and the prospect of cloud computing.

**Variant
Calling**



Many Variant Calling tools out there....

- GATK
- Platypus
- Varscan2
- Atlas2
- Senteion
- FreeBayes
- SNVer
- SAMtools
- VarDict
- LoFreq
-



Reference genome

A B C D E

Deletion

A * C D E

Insertion

A B C Q D E

Duplication

A B C D D' E

Inversion

A D C B E

Translocation

A B C X E F
U V W D Y Z

Some of the genomic artifacts that influence variant calling

- Coverage at that particular base
- Coverage around that particular base
- Base quality at that particular base
- Base quality around that particular base
- Strandedness – is there enough support in the “forward” and “reverse” strands
- Variant ratio = # of variant reads/# of reference reads

Here's what a VCF file looks like

```
1 ##fileformat=VCFv4.0
2 ##fileDate=2017-09-03
3 ##source=Platypus_Version_0.8.1
4 ##platypusOptions={'assemblyRegionSize': 1500, 'trimReadFlank': 0, 'assembleBadReads': 1, 'bamFiles': ['NA24385_bw
5 ##INFO=<ID=FR,Number=.,Type=Float,Description="Estimated population frequency of variant">
6 ##INFO=<ID=MMLQ,Number=1,Type=Float,Description="Median minimum base quality for bases around variant">
7 ##INFO=<ID=TCR,Number=1,Type=Integer,Description="Total reverse strand coverage at this locus">
8 ##INFO=<ID=HP,Number=1,Type=Integer,Description="Homopolymer run length around variant locus">
9 ##INFO=<ID=WE,Number=1,Type=Integer,Description="End position of calling window">
10 ##INFO=<ID=Source,Number=.,Type=String,Description="Was this variant suggested by Platypus, Assembler, or from a
11 ##INFO=<ID=FS,Number=.,Type=Float,Description="Fisher's exact test for strand bias (Phred scale)">
12 ##INFO=<ID=WS,Number=1,Type=Integer,Description="Starting position of calling window">
13 ##INFO=<ID=PP,Number=.,Type=Float,Description="Posterior probability (phred scaled) that this variant segregates">
14 ##INFO=<ID=TR,Number=.,Type=Integer,Description="Total number of reads containing this variant">
15 ##INFO=<ID=NF,Number=.,Type=Integer,Description="Total number of forward reads containing this variant">
16 ##INFO=<ID=TCF,Number=1,Type=Integer,Description="Total forward strand coverage at this locus">
17 ##INFO=<ID=NR,Number=.,Type=Integer,Description="Total number of reverse reads containing this variant">
18 ##INFO=<ID=TC,Number=1,Type=Integer,Description="Total coverage at this locus">
19 ##INFO=<ID=END,Number=.,Type=Integer,Description="End position of reference call block">
20 ##INFO=<ID=MGOF,Number=.,Type=Integer,Description="Worst goodness-of-fit value reported across all samples">
21 ##INFO=<ID=SbPval,Number=.,Type=Float,Description="Binomial P-value for strand bias test">
22 ##INFO=<ID=START,Number=.,Type=Integer,Description="Start position of reference call block">
23 ##INFO=<ID=ReadPosRankSum,Number=.,Type=Float,Description="Mann-Whitney Rank sum test for difference between in pc
24 ##INFO=<ID=MQ,Number=.,Type=Float,Description="Root mean square of mapping qualities of reads at the variant posit
25 ##INFO=<ID=QD,Number=1,Type=Float,Description="Variant-quality/read-depth for this variant">
26 ##INFO=<ID=SC,Number=1,Type=String,Description="Genomic sequence 10 bases either side of variant position">
27 ##INFO=<ID=BRF,Number=1,Type=Float,Description="Fraction of reads around this variant that failed filters">
28 ##INFO=<ID=HapScore,Number=.,Type=Integer,Description="Haplotype score measuring the number of haplotypes the vari
29 ##INFO=<ID=Size,Number=.,Type=Integer,Description="Size of reference call block">
30 ##FILTER=<ID=GOF,Description="Variant fails goodness-of-fit test.">
31 ##FILTER=<ID=badReads,Description="Variant supported only by reads with low quality bases close to variant positio
32 ##FILTER=<ID=alleleBias,Description="Variant frequency is lower than expected for het">
33 ##FILTER=<ID=hp10,Description="Flanking sequence contains homopolymer of length 10 or greater">
34 ##FILTER=<ID=Q20,Description="Variant quality is below 20.">
35 ##FILTER=<ID=HapScore,Description="Too many haplotypes are supported by the data in this region.">
36 ##FILTER=<ID=MQ,Description="Root-mean-square mapping quality across calling region is low.">
37 ##FILTER=<ID=strandBias,Description="Variant fails strand-bias filter">
```

```

36 ##FILTER=<ID=MQ,Description="Root-mean-square mapping quality across calling region is low.">
37 ##FILTER=<ID=strandBias,Description="Variant fails strand-bias filter">
38 ##FILTER=<ID=SC,Description="Variants fail sequence-context filter. Surrounding sequence is low-complexity">
39 ##FILTER=<ID=QualDepth,Description="Variant quality/Read depth ratio is low.">
40 ##FILTER=<ID=REFCALL,Description="This line represents a homozygous reference call">
41 ##FILTER=<ID=QD,Description="Variants fail quality/depth filter.">
42 ##FORMAT=<ID=GT,Number=1,Type=String,Description="Unphased genotypes">
43 ##FORMAT=<ID=GQ,Number=.,Type=Integer,Description="Genotype quality as phred score">
44 ##FORMAT=<ID=GOF,Number=.,Type=Float,Description="Goodness of fit value">
45 ##FORMAT=<ID=NR,Number=.,Type=Integer,Description="Number of reads covering variant location in this sample">
46 ##FORMAT=<ID=GL,Number=.,Type=Float,Description="Genotype log10-likelihoods for AA,AB and BB genotypes, where A =
47 ##FORMAT=<ID=NV,Number=.,Type=Integer,Description="Number of reads containing variant in this sample">
48 #CHROM POS ID REF ALT QUAL FILTER INFO FORMAT NA24385
49 1 10439 . AC A 68 PASS BRF=0.0;FR=0.9998;HP=4;HapScore=2;MGOF=39;MMLQ=38;MQ=46.0;NF=1;NR=1;PP=68;QD=3
50 1 846338 . A G 44 PASS BRF=0.0;FR=0.9998;HP=2;HapScore=2;MGOF=0;MMLQ=39;MQ=60.0;NF=1;NR=1;PP=44;QD=39
51 1 866319 . G A 45 PASS BRF=0.0;FR=0.9998;HP=3;HapScore=2;MGOF=0;MMLQ=39;MQ=60.0;NF=2;NR=0;PP=45;QD=39
52 1 871334 . G T 314 PASS BRF=0.0;FR=1.0000;HP=1;HapScore=2;MGOF=0;MMLQ=37;MQ=60.0;NF=2;NR=7;PP=314;QD=3
53 1 876499 . A G 168 PASS BRF=0.0;FR=1.0000;HP=6;HapScore=2;MGOF=0;MMLQ=37;MQ=60.0;NF=5;NR=1;PP=168;QD=3
54 1 877831 . T C 346 PASS BRF=0.17;FR=1.0000;HP=2;HapScore=2;MGOF=3;MMLQ=35;MQ=60.0;NF=9;NR=1;PP=346;QD=
55 1 878314 . G C 342 PASS BRF=0.0;FR=0.5000;HP=2;HapScore=2;MGOF=2;MMLQ=37;MQ=60.0;NF=7;NR=6;PP=342;QD=2
56 1 879676 . G A 517 PASS BRF=0.15;FR=1.0000;HP=3;HapScore=1;MGOF=9;MMLQ=38;MQ=60.0;NF=7;NR=7;PP=517;QD=
57 1 879687 . T C 540 PASS BRF=0.15;FR=1.0000;HP=3;HapScore=1;MGOF=9;MMLQ=38;MQ=60.0;NF=7;NR=8;PP=540;QD=
58 1 880238 . A G 351 PASS BRF=0.0;FR=1.0000;HP=2;HapScore=2;MGOF=0;MMLQ=39;MQ=60.0;NF=3;NR=7;PP=351;QD=3
59 1 881627 . G A 463 PASS BRF=0.13;FR=1.0000;HP=3;HapScore=2;MGOF=3;MMLQ=38;MQ=60.0;NF=6;NR=7;PP=463;QD=
60 1 883625 . A G 45 PASS BRF=0.5;FR=0.9998;HP=3;HapScore=2;MGOF=0;MMLQ=39;MQ=60.0;NF=1;NR=1;PP=45;QD=39
61 1 887560 . A C 379 PASS BRF=0.0;FR=1.0000;HP=1;HapScore=2;MGOF=1;MMLQ=39;MQ=60.0;NF=2;NR=9;PP=379;QD=3
62 1 887801 . A G 697 PASS BRF=0.05;FR=1.0000;HP=2;HapScore=1;MGOF=0;MMLQ=39;MQ=58.63;NF=13;NR=5;PP=697;Q
63 1 888639 . T C 273 PASS BRF=0.31;FR=1.0000;HP=1;HapScore=2;MGOF=17;MMLQ=38;MQ=50.09;NF=3;NR=5;PP=273;Q
64 1 888659 . T C 120 PASS BRF=0.2;FR=1.0000;HP=1;HapScore=2;MGOF=23;MMLQ=34;MQ=47.33;NF=1;NR=3;PP=120;QD
65 1 889158 . GA CC 571 PASS BRF=0.25;FR=1.0000;HP=2;HapScore=1;MGOF=8;MMLQ=39;MQ=60.0;NF=5;NR=3;PP=571;QD=
66 1 892745 . G A 159 PASS BRF=0.0;FR=1.0000;HP=2;HapScore=2;MGOF=0;MMLQ=38;MQ=60.0;NF=0;NR=5;PP=159;QD=3
67 1 894573 . G A 433 PASS BRF=0.0;FR=1.0000;HP=1;HapScore=2;MGOF=0;MMLQ=36;MQ=60.0;NF=8;NR=5;PP=433;QD=3
68 1 896271 . C T 337 PASS BRF=0.13;FR=1.0000;HP=2;HapScore=2;MGOF=0;MMLQ=36;MQ=60.0;NF=3;NR=7;PP=337;QD=
69 1 897325 . G C 691 PASS BRF=0.09;FR=1.0000;HP=2;HapScore=1;MGOF=1;MMLQ=38;MQ=60.0;NF=9;NR=10;PP=691;QD
70 1 897564 . T C 122 PASS BRF=0.0;FR=1.0000;HP=2;HapScore=2;MGOF=0;MMLQ=39;MQ=60.0;NF=4;NR=0;PP=122;QD=3
71 1 898323 . T C 506 PASS BRF=0.24;FR=1.0000;HP=1;HapScore=1;MGOF=0;MMLQ=36;MQ=60.0;NF=3;NR=11;PP=506;QD
72 1 899928 . GGGGAGGGGGGCGCG CGGGAGGGGTCCGCA 164 PASS BRF=0.0;FR=0.9998;HP=3;HapScore=3;MGOF=0;MMLQ=39;MQ=60

```

Format

Genotype

```
TCF=1;TCR=1;TR=2;WE=10447;WS=10429 GT:GL:GOF:GQ:NR:NV 1/1:-7.8,-0.3,0.0:39:5:2:2
s;TC=2;TCF=1;TCR=1;TR=2;WE=846346;WS=846328 GT:GL:GOF:GQ:NR:NV 1/1:-7.9,-0.3,0.0:0:5:2:2
;TC=2;TCF=2;TCR=0;TR=2;WE=866327;WS=866309 GT:GL:GOF:GQ:NR:NV 1/1:-8.0,-0.3,0.0:0:5:2:2
us;TC=9;TCF=2;TCR=7;TR=9;WE=871342;WS=871324 GT:GL:GOF:GQ:NR:NV 1/1:-34.9,-2.41,0.0:0:24:9:9
us;TC=6;TCF=5;TCR=1;TR=6;WE=876507;WS=876489 GT:GL:GOF:GQ:NR:NV 1/1:-20.3,-1.51,0.0:0:15:6:6
pus;TC=10;TCF=9;TCR=1;TR=10;WE=877839;WS=877821 GT:GL:GOF:GQ:NR:NV 1/1:-38.1,-2.71,0.0:3:27:10:10
us;TC=38;TCF=19;TCR=19;TR=13;WE=878322;WS=878304 GT:GL:GOF:GQ:NR:NV 1/0:-38.26,0.0,-81.46:2:99:38:13
pus;TC=14;TCF=7;TCR=7;TR=14;WE=879695;WS=879666 GT:GL:GOF:GQ:NR:NV 1/1:-55.0,-3.74,0.0:9:37:14:14
pus;TC=15;TCF=7;TCR=8;TR=15;WE=879695;WS=879666 GT:GL:GOF:GQ:NR:NV 1/1:-56.9,-3.91,0.0:9:39:15:15
us;TC=10;TCF=3;TCR=7;TR=10;WE=880246;WS=880228 GT:GL:GOF:GQ:NR:NV 1/1:-38.6,-2.71,0.0:0:27:10:10
us;TC=13;TCF=6;TCR=7;TR=13;WE=881635;WS=881617 GT:GL:GOF:GQ:NR:NV 1/1:-49.8,-3.61,0.0:3:36:13:13
s;TC=2;TCF=1;TCR=1;TR=2;WE=883633;WS=883615 GT:GL:GOF:GQ:NR:NV 1/1:-8.0,-0.3,0.0:0:5:2:2
s;TC=11;TCF=2;TCR=9;TR=11;WE=887568;WS=887550 GT:GL:GOF:GQ:NR:NV 1/1:-41.4,-3.01,0.0:1:30:11:11
typus;TC=18;TCF=13;TCR=5;TR=18;WE=887809;WS=887791 GT:GL:GOF:GQ:NR:NV 1/1:-73.2,-5.42,0.0:0:54:18:18
typus;TC=8;TCF=3;TCR=5;TR=8;WE=888647;WS=888629 GT:GL:GOF:GQ:NR:NV 1/1:-30.8,-2.11,0.0:17:21:8:8
typus;TC=4;TCF=1;TCR=3;TR=4;WE=888667;WS=888649 GT:GL:GOF:GQ:NR:NV 1/1:-15.5,-0.9,0.0:23:10:4:4
pus;TC=9;TCF=6;TCR=3;TR=8;WE=889167;WS=889148 GT:GL:GOF:GQ:NR:NV 1/1:-62.4,-2.11,0.0:8:21:9:8
s;TC=5;TCF=0;TCR=5;TR=5;WE=892753;WS=892735 GT:GL:GOF:GQ:NR:NV 1/1:-19.4,-1.2,0.0:0:12:5:5
us;TC=13;TCF=8;TCR=5;TR=13;WE=894581;WS=894563 GT:GL:GOF:GQ:NR:NV 1/1:-46.8,-3.61,0.0:0:36:13:13
pus;TC=10;TCF=3;TCR=7;TR=10;WE=896279;WS=896261 GT:GL:GOF:GQ:NR:NV 1/1:-37.2,-2.71,0.0:0:27:10:10
typus;TC=19;TCF=9;TCR=10;TR=19;WE=897333;WS=897315 GT:GL:GOF:GQ:NR:NV 1/1:-72.6,-5.42,0.0:1:54:19:19
s;TC=4;TCF=4;TCR=0;TR=4;WE=897572;WS=897554 GT:GL:GOF:GQ:NR:NV 1/1:-15.7,-0.9,0.0:0:10:4:4
typus;TC=14;TCF=3;TCR=11;TR=14;WE=898331;WS=898313 GT:GL:GOF:GQ:NR:NV 1/1:-54.1,-3.91,0.0:0:39:14:14
bPval=1.0;Source=Platypus;TC=5;TCF=0;TCR=5;TR=2;WE=899950;WS=899918 GT:GL:GOF:GQ:NR:NV 1/1:-42.5,-1.2,0.0:0:12:5:2
```

Now we have the capability
to evaluate the quality of
sequencing and
bioinformatic analyses

Utah trio (NA12878) Bioinformatics

CORIELL TRIO 1 (UTAH TRIO)	NA12878-proband	NA12891-dad	NA12892-mom	UNITS	REFERENCE RANGE
Sex	F verified	M verified	F verified	trio order verified	
Yield: raw/bulk	201.5	141.1	161.4	Gbp	> 180
% mapped	99.24%	99.11%	99.15%	pct	98-100
% duplicates	8.82%	7.83%	8.17%	pct	< 15%
Yield	181.9	128.8	146.8	Gbp	> 130
Insert size: Mean +/- std.dev	388.2 +/- 84.1	400.0 +/- 87.8	393.3 +/- 85.3	bp	300-480
Average and median coverage across genome	55.0	39.0	43.0	x	> 40
Average coverage over OMIM genes	59.0	42.0	47.0	x	> 40
# of OMIM genes with coverage at <10X (and list)	249	359	260	ENST	< 2% (282)
# of OMIM genes with 100% coverage at >=10X	98.2%	97.5%	98.2%	pct	> 98%
# of OMIM genes with 100% coverage at >=20X	97.7%	91.9%	96.9%	pct	> 94%
# of OMIM genes with 100% coverage at >=30X	95.8%	56.1%	83.1%	pct	> 80%
# of genes with 100% coverage at >=40X	81.5%	7.3%	23.8%	pct	
Variation (VCF) metrics					
# of calls Total	4850124	4783261	4841927		2.5-6.0M
# of PASS calls	4782957	4692663	4758545		2.5-6.0M
# of calls Total coding	27117	26583	26730		25000-30000
Total # of SNVs	3989749 [82.26%]	3953443 [82.65%]	3996596 [82.54%]		
Total # of Indels	860375 [17.74%]	829818 [17.35%]	845331 [17.46%]		
Hom/Het ratio (in coding regions)	0.58 (0.60)	0.58 (0.59)	0.58 (0.59)	ratio	0.5-0.61
Ti/Tv ratio (in coding regions)	2.00 (2.86)	2.00 (2.89)	2.01 (2.88)	ratio	2-2.2 (2.8-3)
# of het calls (# of hom call)	3087082 (1763042)	2980726 (1802535)	3085568 (1756359)	units	
In-silico sample swap check	PASS	PASS	PASS		Mendelian test
Automated upload of VCF to Omicia	PASS	PASS	PASS		
Precision metrics compared to GIAB: total (on SNPs)	0.9895 (0.9915)	n/a	n/a		GIAB only available for NA12878
Sensitivity metrics : total (on SNPs)	0.9947 (0.9975)	n/a	n/a		
Update LIMS	TBD	TBD	TBD		
Download annotated VCF to RCI	N/A	N/A	N/A		

Just a slight detour

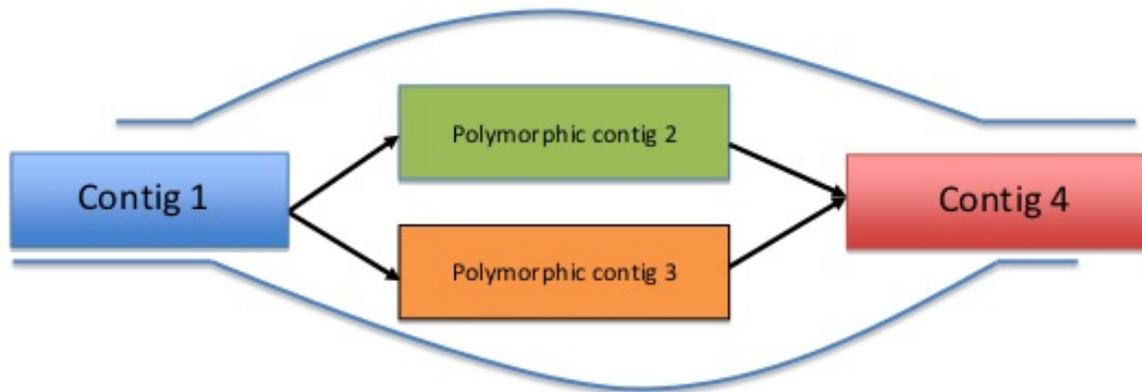
Bigger pixels = better picture

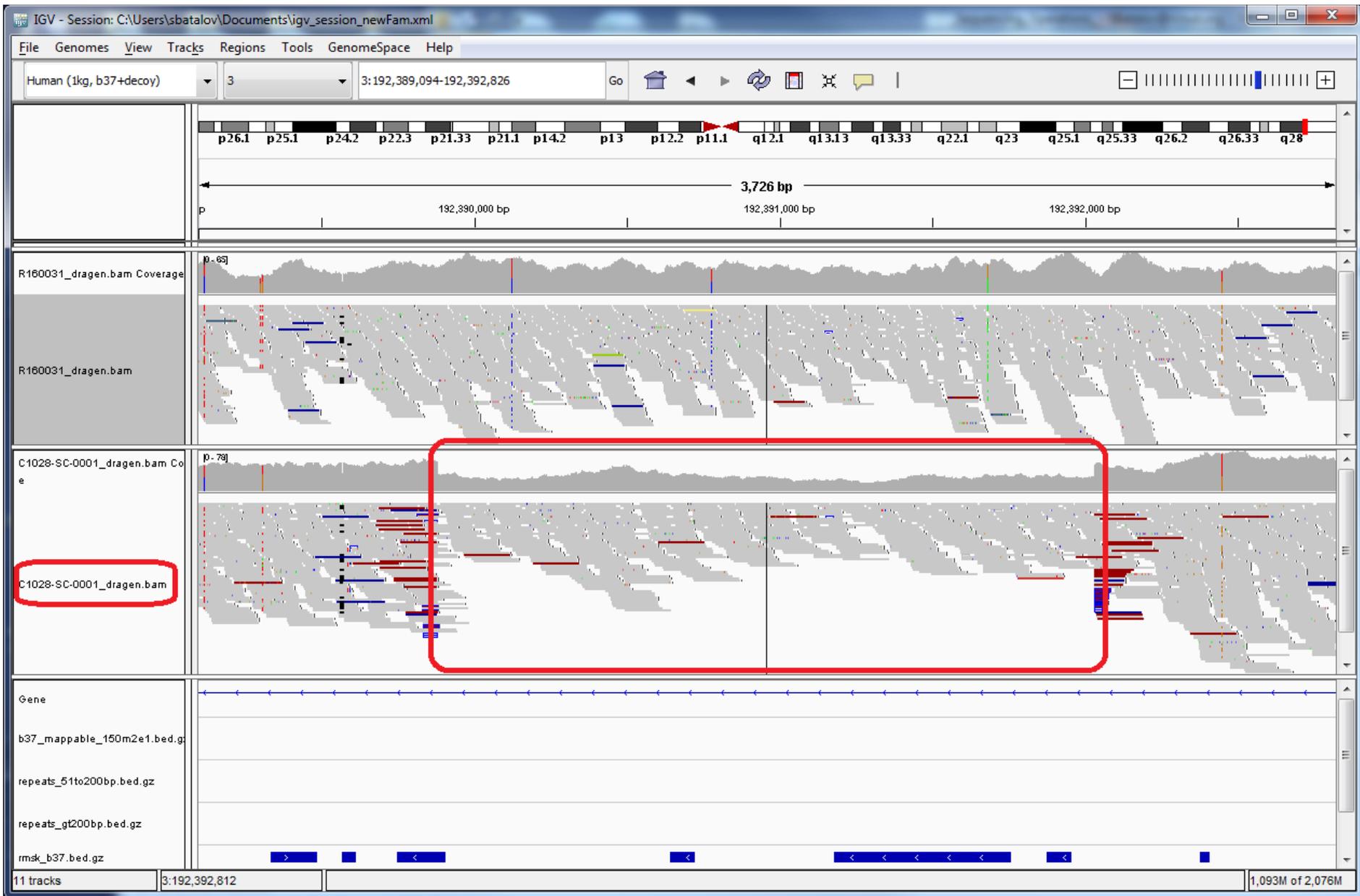


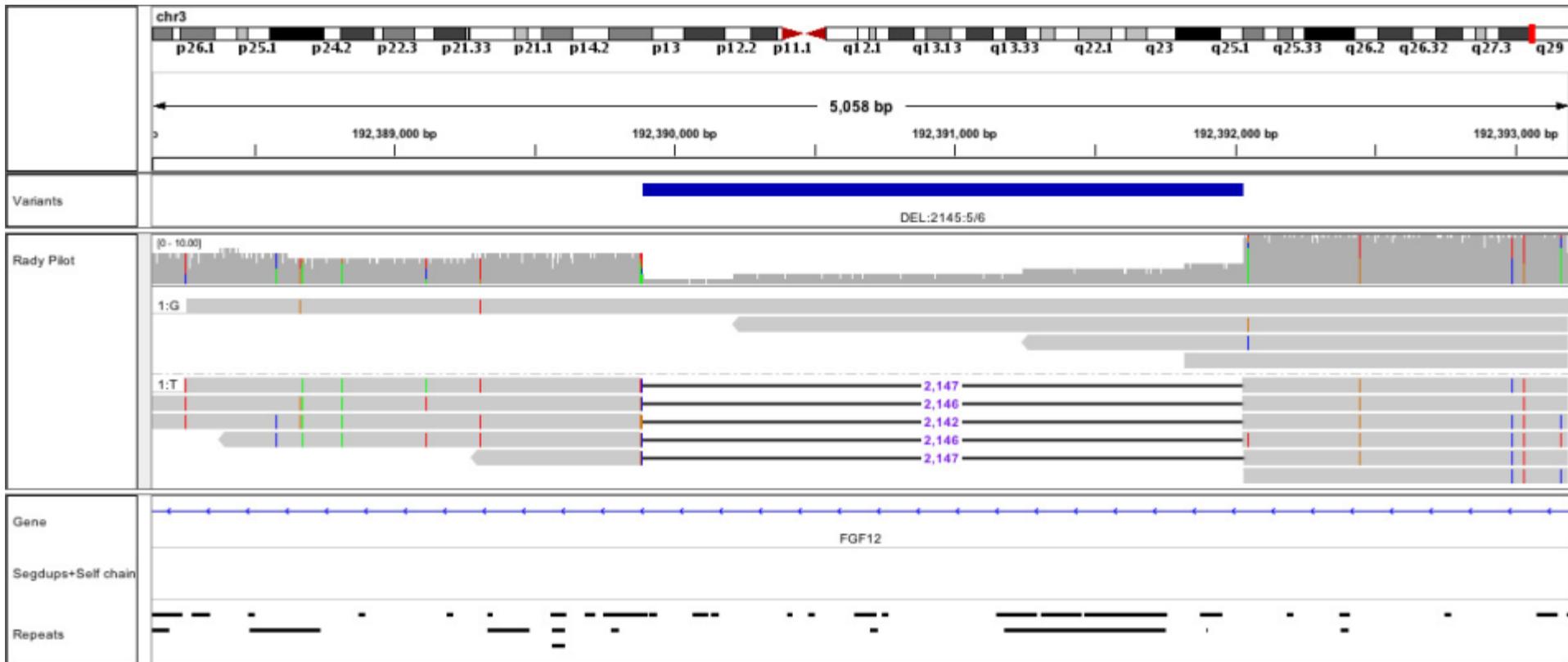
Longer reads?



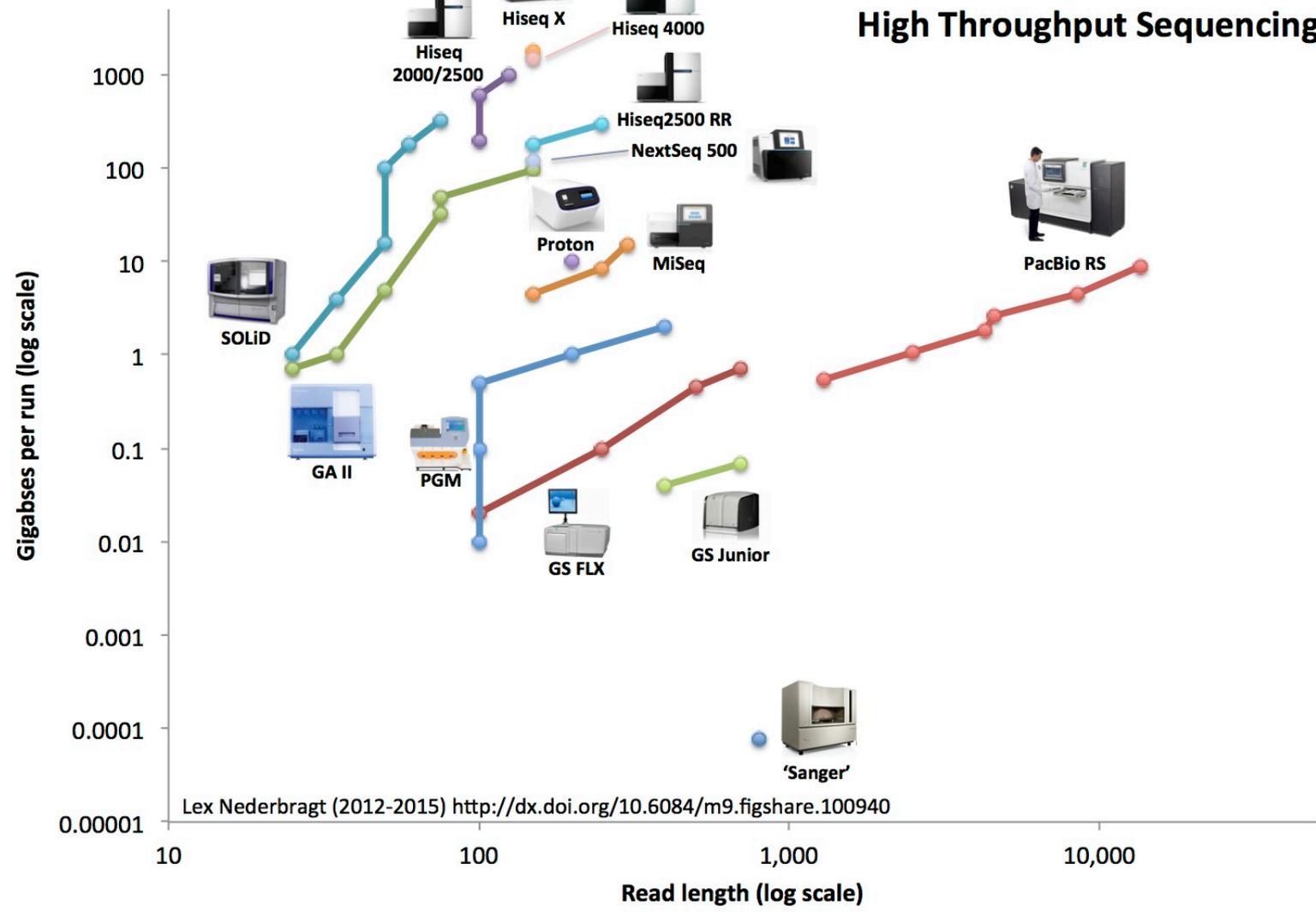
Long reads can span repeats and heterozygous regions







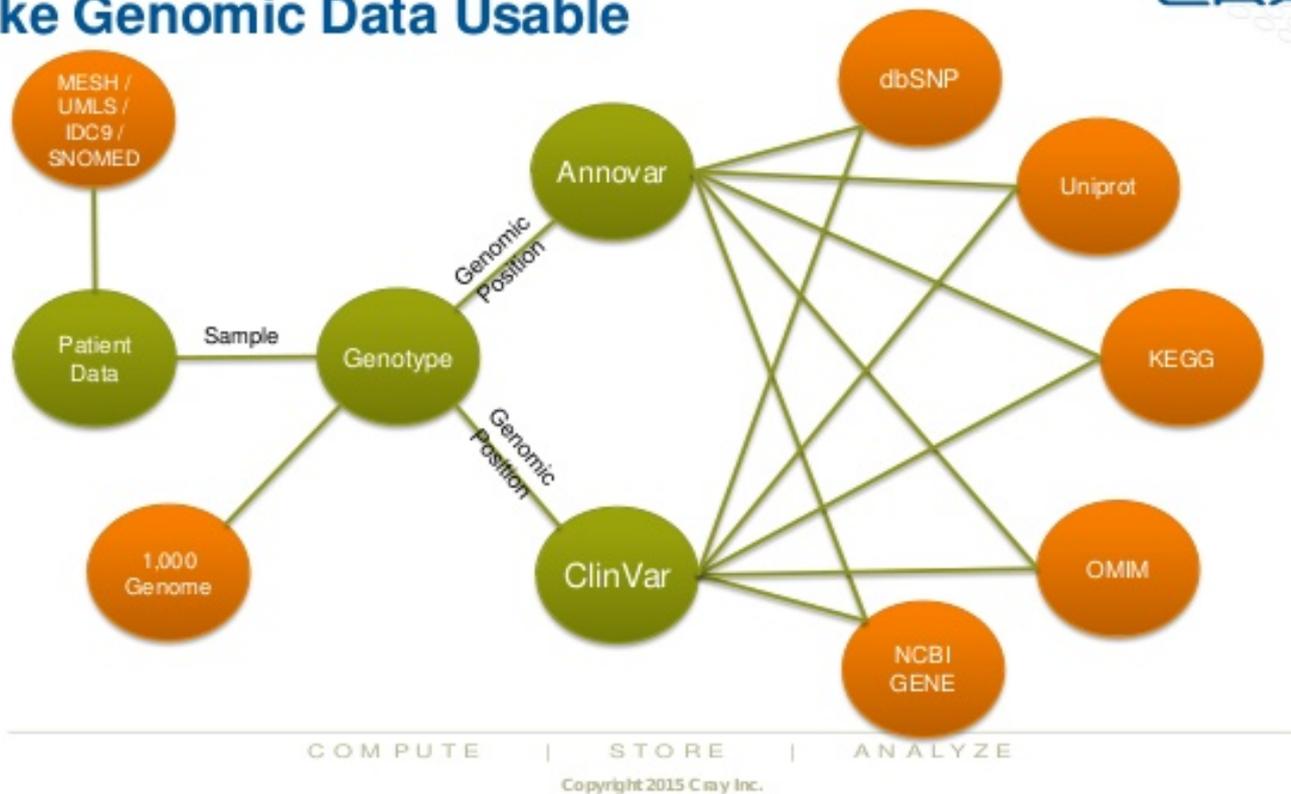
Developments in High Throughput Sequencing



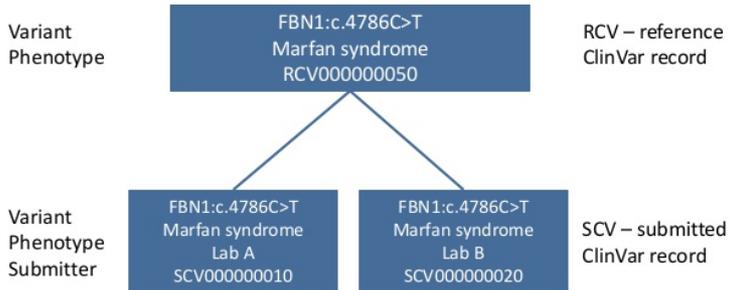
Back to our bioinformatics pipeline....

Annotation

Make Genomic Data Usable

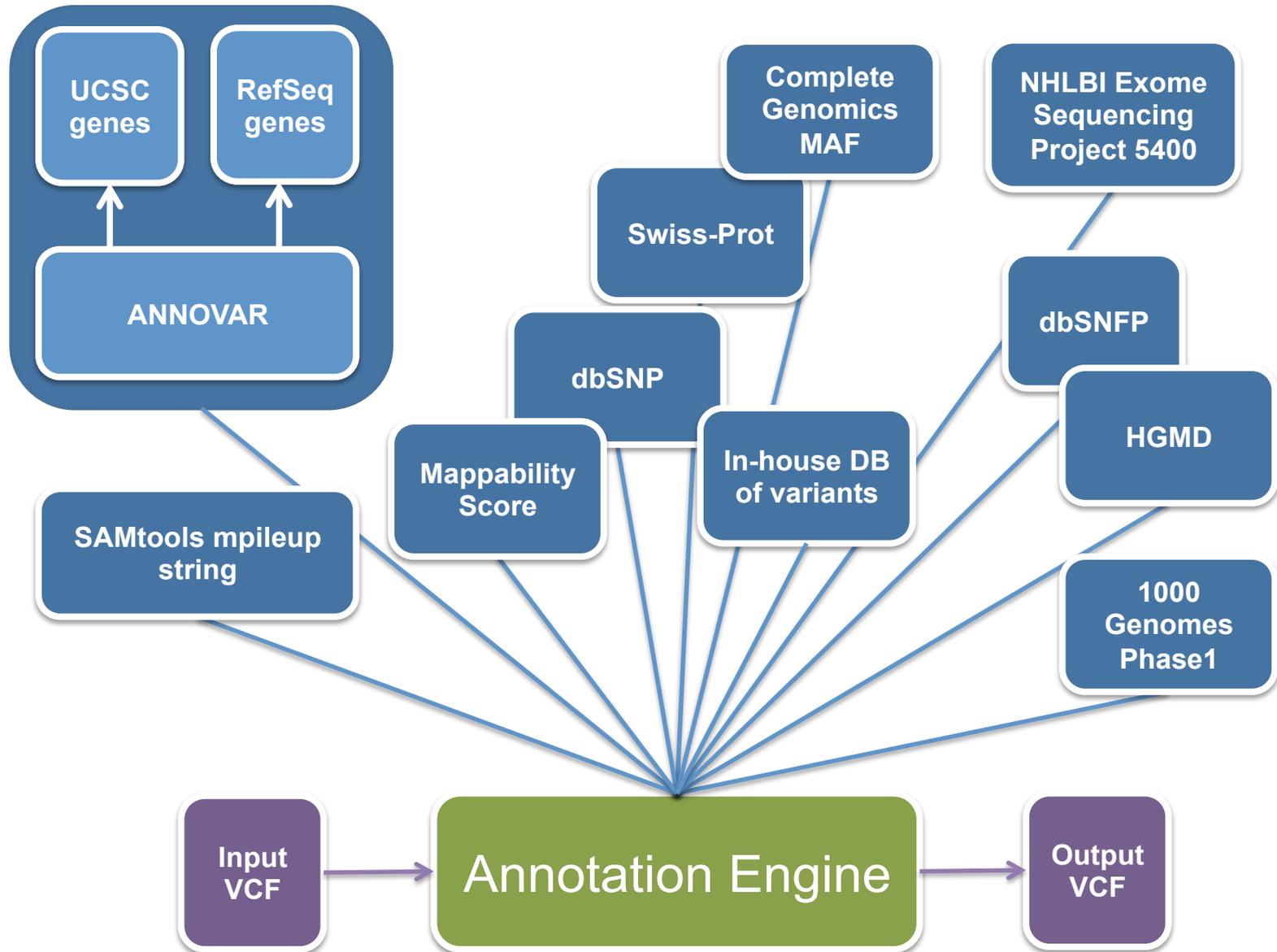


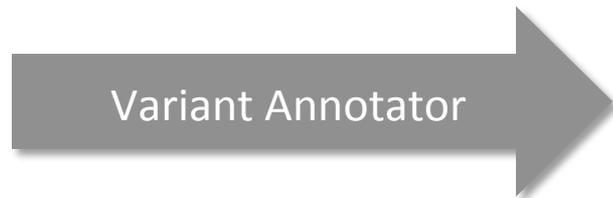
ClinVar aggregates by variant and phenotype



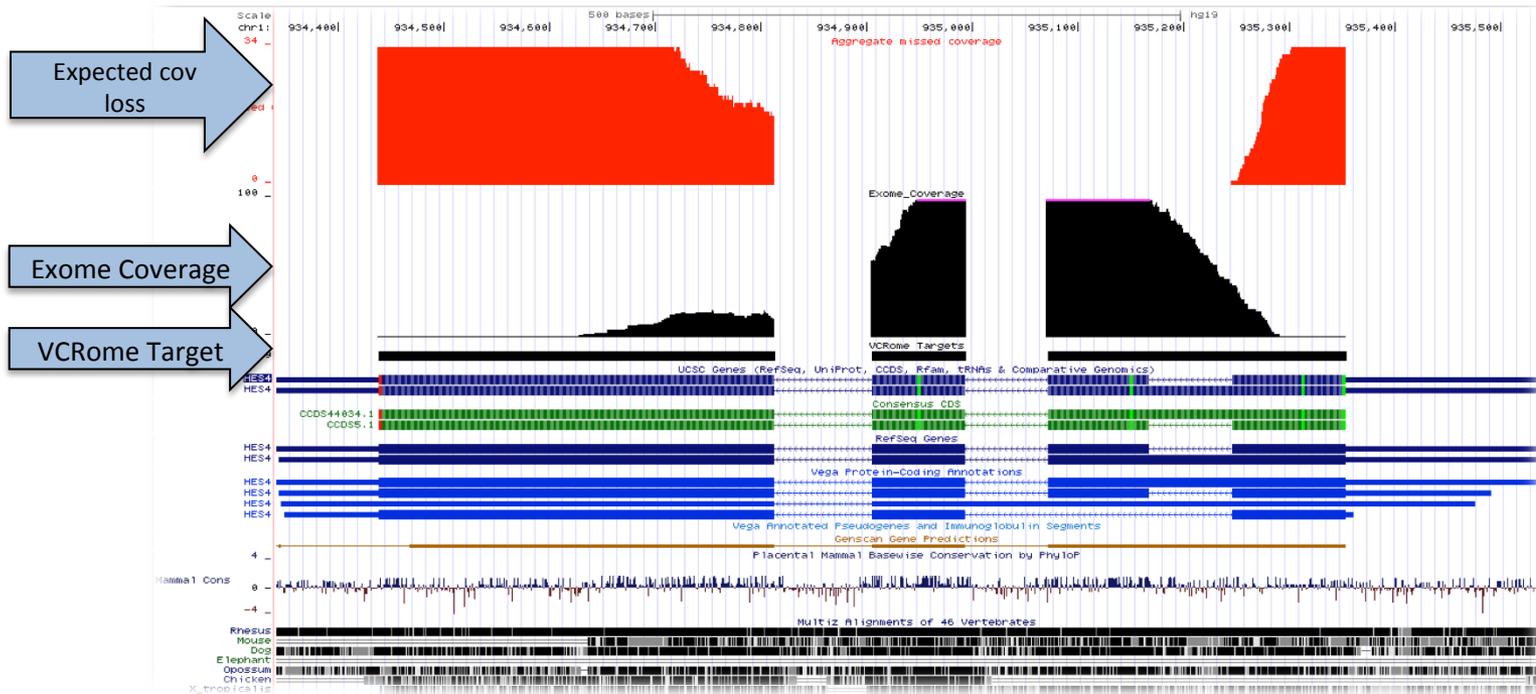
Integrate info from multiple public and private data bases

Variant Annotation Sources



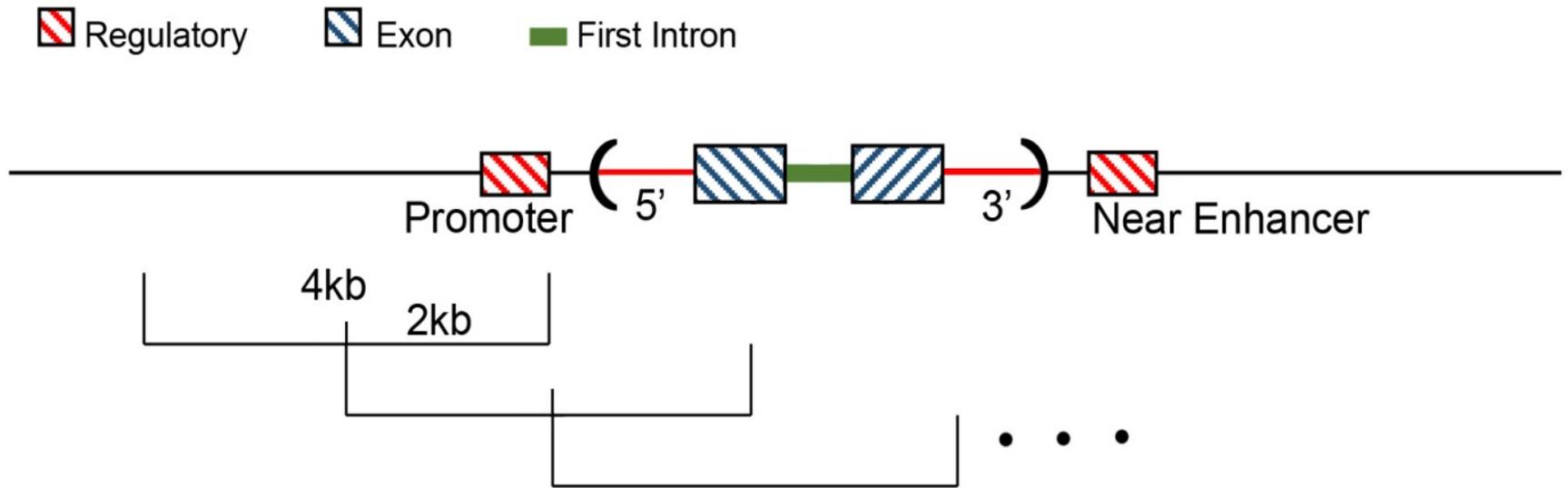


Exome Coverage and Identification (ExCID) Report



ExCID has a visualization component which allows the user to see coverage across a region of interest and correlate that with portions of the exon target information and regions of expected coverage loss.

Annotated functional motifs



Regulatory Domain

- Promoter
- UTR3
- UTR5
- Enhancer

Regulatory Domain Annotation

All SNVs – annotated using the WGSA pipeline, version 05 (Liu et al. 2016).

- Promoter – based on the overlap between the permissive set of Cap Analysis of Gene Expression (CAGE) peaks in FANTOM5 data (FANTOM Consortium and the RIKEN PMI and CLST (DGT) 2014) and the 5 kb upstream region (ANNOVAR(Wang et al. 2010) annotation, refSeq gene model - upstream).
- UTR3' – if variant overlaps a 3' untranslated region (ANNOVAR annotation, RefSeq gene model – UTR3).
- UTR5' – if variant overlaps a 5' untranslated region (ANNOVAR annotation, RefSeq gene model – UTR5).

Enhancer

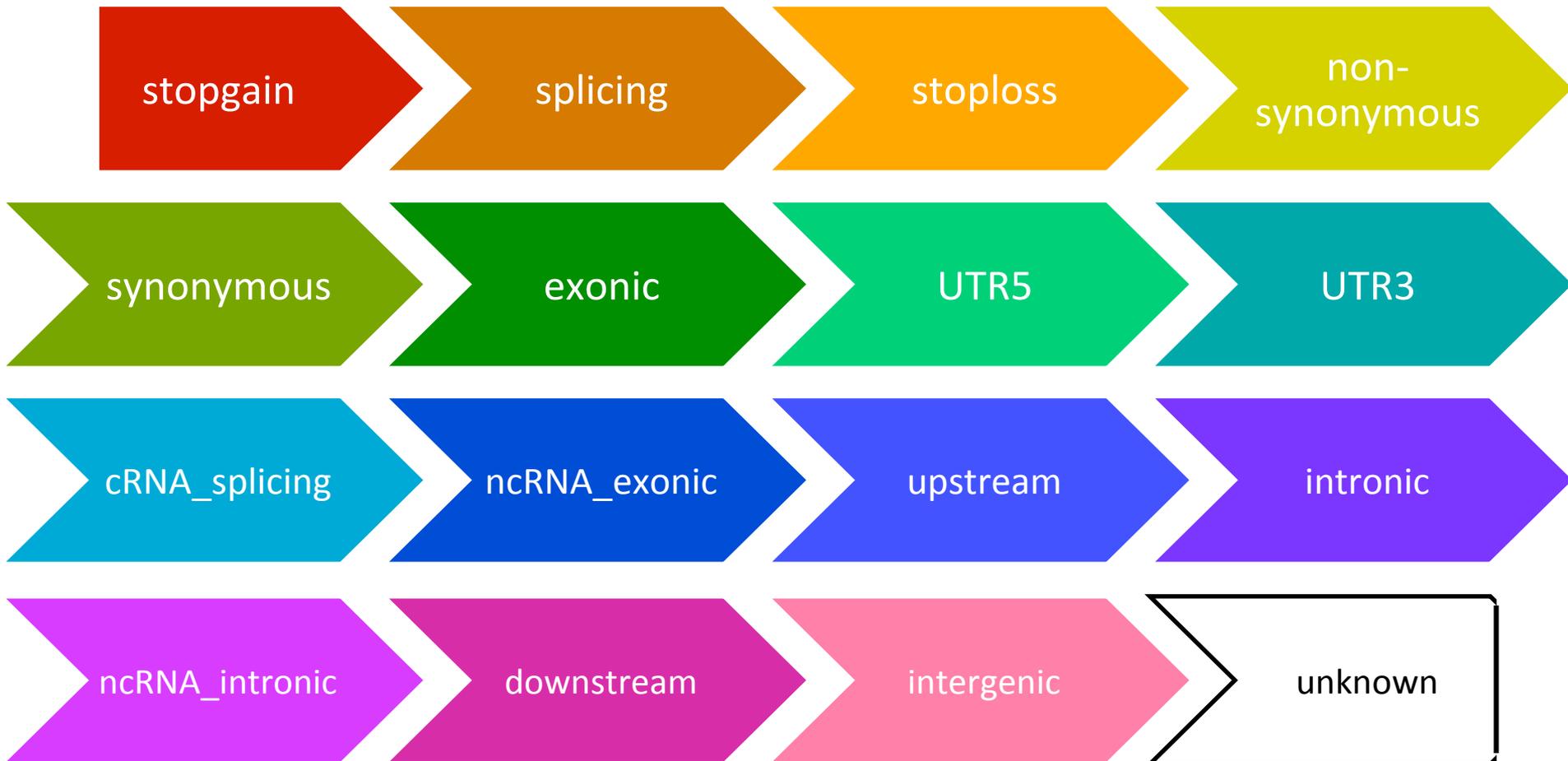
The enhancers and the target genes of the enhancers were defined based on the permissive set of enhancers and enhancer-promoter pairs reported by the FANTOM5 project (Andersson et al. 2014).

Multiple assignment of SNVs

Among 770,137 SNVs ($MAF \leq 0.05$) annotated to **regulatory domain** ($MAC \geq 3$) in AA (ARIC), **11.39%** SNVs (87,718 variants) were assigned to more than 1 gene.

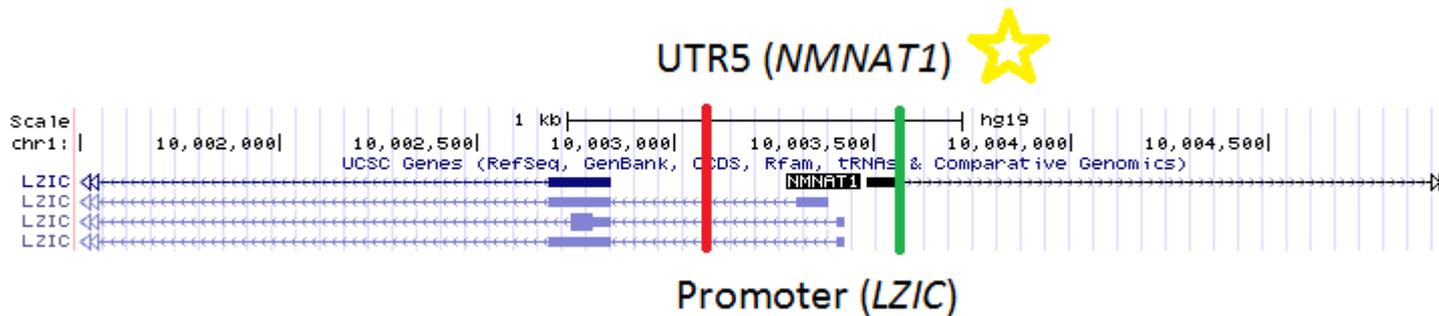
*Unique SNVs assigned to different genes /
Unique SNVs = $87,718 / 770,137 = 0.1139$*

If SNV has multiple gene-based annotations due to overlapping genes, the most “deleterious” consequence (reported by ANNOVAR using the RefSeq gene model) is used, based on the following order:

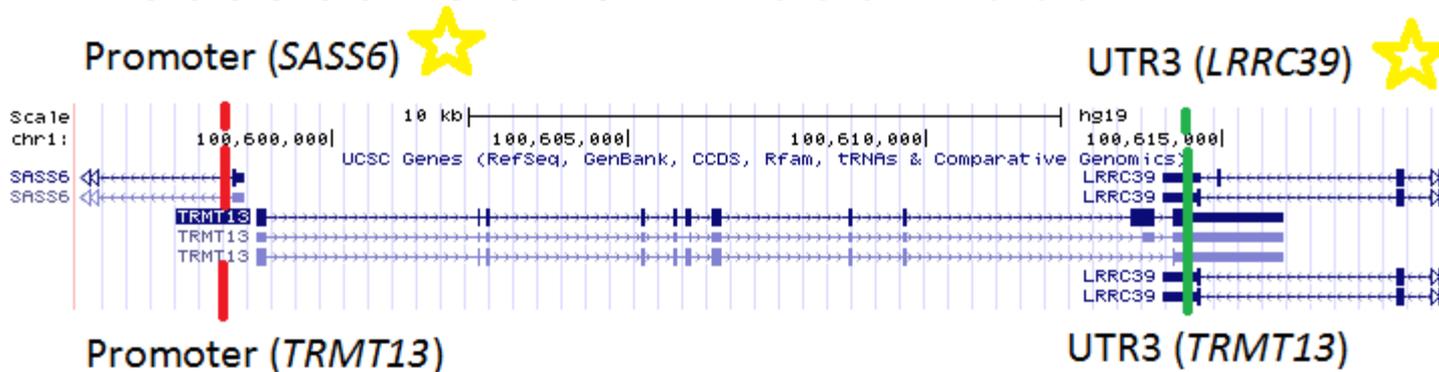


Multiple SNV assignment: examples

1:10003077 and 1:10003539



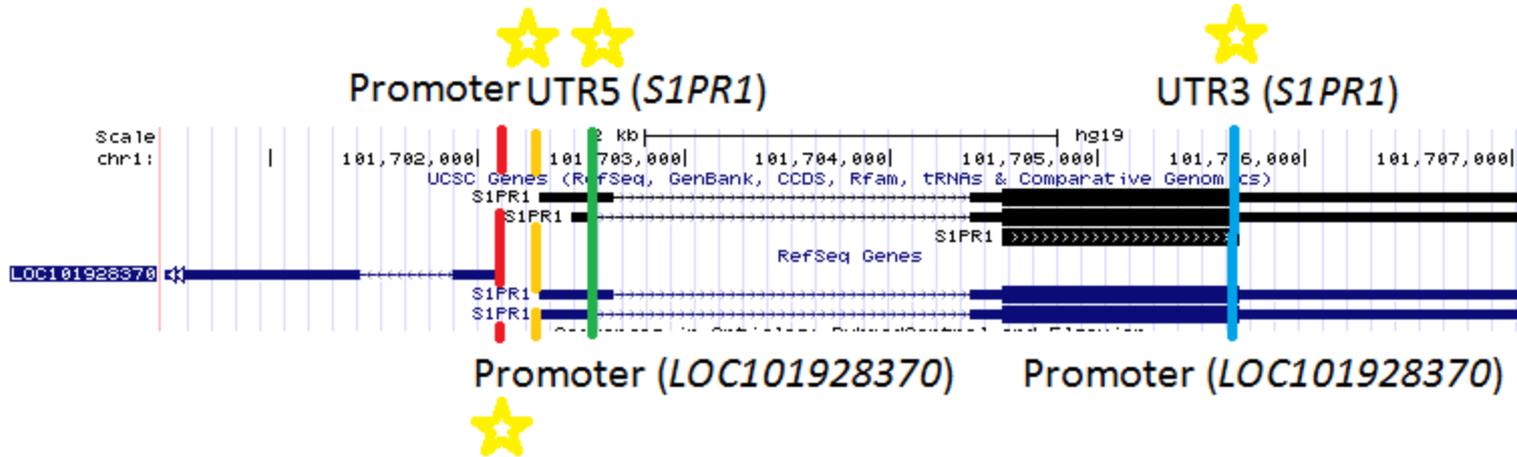
1:100598518 and 1:100614563



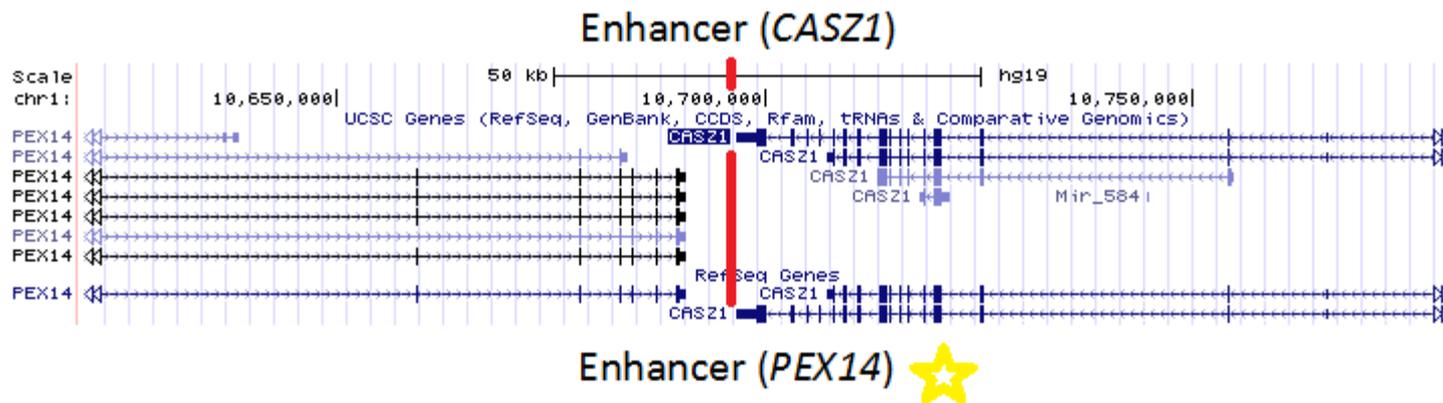
- ☆ - Signifies which variant would have been chosen, if each SNV was assigned to be “unique” to one regulatory unit.

More examples

1:101702176, 1:101702359, 1:101702566, 1:101705827

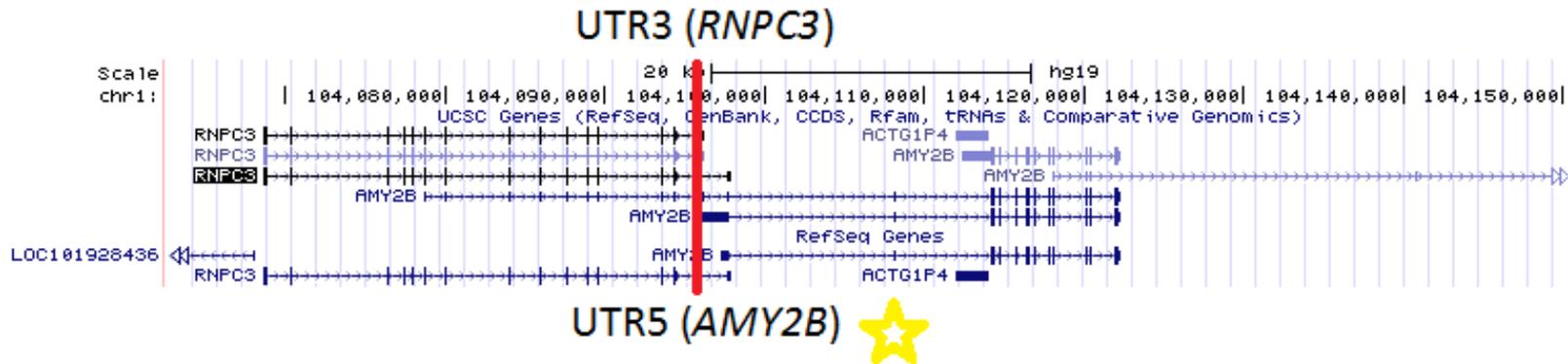


1:10695323



... and one more

1:104097688



Break

Variant Filtering and Prioritization

Prioritization

Clinical phenotypes -> Disease -> Genes

Genome -> Variants -> Genes

Candidate
variants

```
graph LR; A[Clinical phenotypes -> Disease -> Genes] --- B{ }; C[Genome -> Variants -> Genes] --- B; B --- D[Candidate variants]
```

The diagram illustrates two parallel paths for identifying candidate variants. The top path, highlighted in a red rounded rectangle, follows the sequence: Clinical phenotypes -> Disease -> Genes. The bottom path, highlighted in a purple rounded rectangle, follows the sequence: Genome -> Variants -> Genes. Both paths are grouped by a large grey curly bracket on the right side, which points to a grey rounded rectangle labeled 'Candidate variants'.

Prioritization – Genome Based

Clinical phenotypes -> Disease -> Genes

Genome -> Variants -> Genes

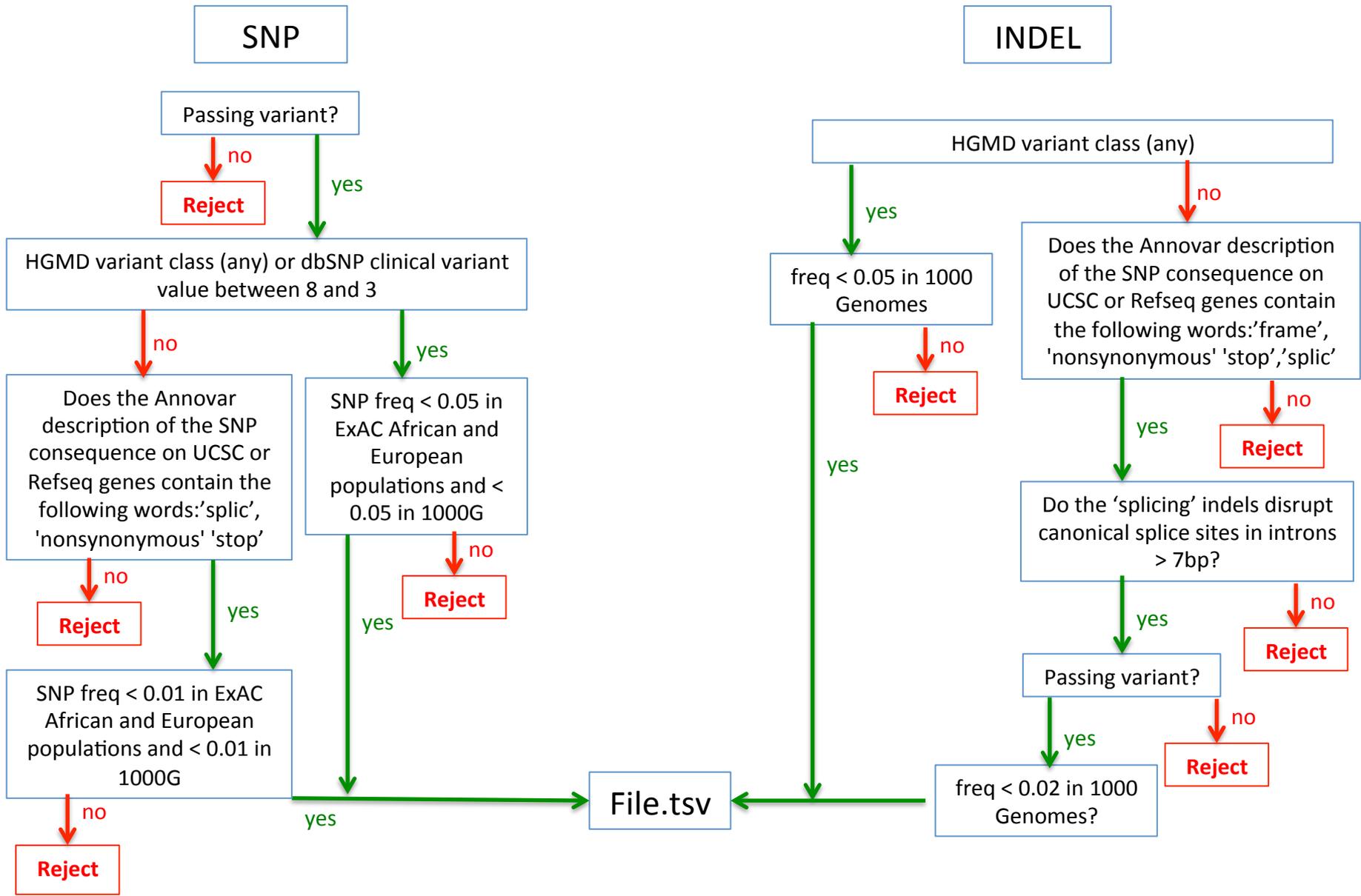
Candidate
variants

```
graph LR; A[Clinical phenotypes -> Disease -> Genes] --- B{ }; C[Genome -> Variants -> Genes] --- B; B --- D[Candidate variants]
```

The diagram illustrates two parallel paths that converge into a single outcome. The top path, shown in a light red box, represents a clinical approach: starting with clinical phenotypes, identifying the disease, and then finding the associated genes. The bottom path, shown in a purple box, represents a genome-based approach: starting with the genome, identifying variants, and then finding the associated genes. Both paths are grouped by a large grey curly bracket on the right, which points to a grey box labeled 'Candidate variants', indicating that both methods lead to the identification of potential genetic candidates.

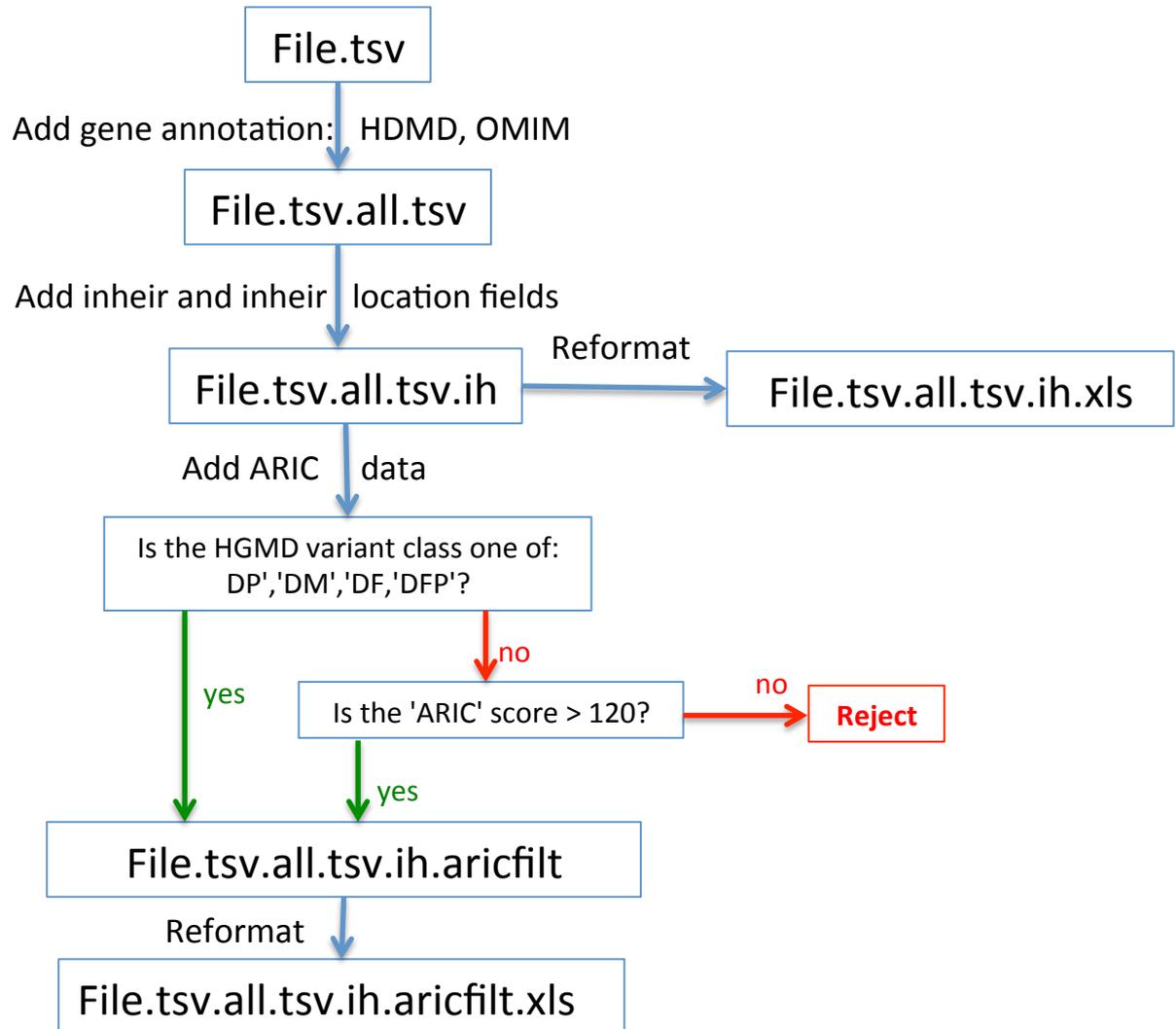
Mendelian Filter

Example filtering workflow



Mendelian Filtering

Secondary filtering



Prioritization

Clinical phenotypes -> Disease -> Genes

Genome -> Variants -> Genes

Candidate
variants

```
graph LR; A[Clinical phenotypes -> Disease -> Genes] --- B{ }; C[Genome -> Variants -> Genes] --- B; B --- D[Candidate variants]
```

The diagram illustrates two parallel paths for identifying candidate variants. The top path, highlighted in a red rounded rectangle, follows the sequence: Clinical phenotypes -> Disease -> Genes. The bottom path, highlighted in a purple rounded rectangle, follows the sequence: Genome -> Variants -> Genes. Both paths are grouped by a large grey curly bracket on the right side, which points to a grey rounded rectangle labeled 'Candidate variants'.

Phenotype Based Prioritization

Clinical phenotypes -> Disease -> Genes

Genome -> Variants -> Genes

Candidate
variants

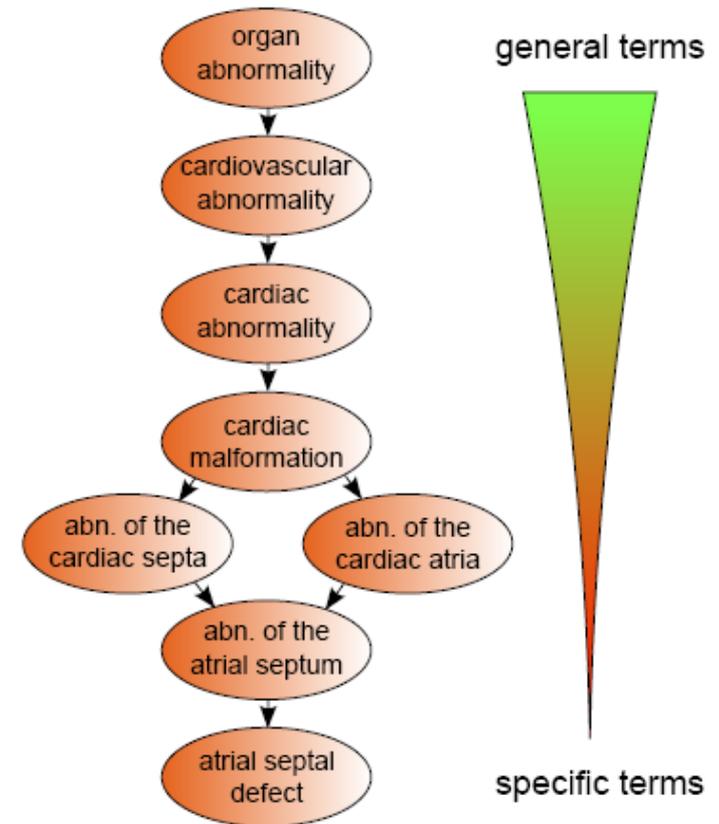
```
graph LR; A[Clinical phenotypes -> Disease -> Genes] --- B{ }; C[Genome -> Variants -> Genes] --- B; B --- D[Candidate variants]
```

The diagram illustrates two parallel workflows that converge on a single outcome. The top workflow, shown in a red box, starts with 'Clinical phenotypes', leading to 'Disease', and then to 'Genes'. The bottom workflow, shown in a purple box, starts with 'Genome', leading to 'Variants', and then to 'Genes'. Both workflows are grouped by a large grey curly bracket on the right side, which points to a grey box labeled 'Candidate variants'.

The Human Phenotype Ontology (HPO*)

www.human-phenotype-ontology.org

- The HPO provides a standardized vocabulary of phenotypic abnormalities encountered in human genetic syndromes
- Phenotypic features are formally represented as terms of a directed acyclic graph:
 - Terms are related to parent terms by „is a“ relationships, representing subclasses of more general parent terms
 - Multiple parentage allows the representation of different aspects of phenotypic abnormalities



* **Robinson P, Köhler S, Bauer S, Seelow D, Horn D, Mundlos:** The Human Phenotype Ontology: A Tool for annotating and analyzing human hereditary disease, *Am J Hum Genet.* 2008 Nov

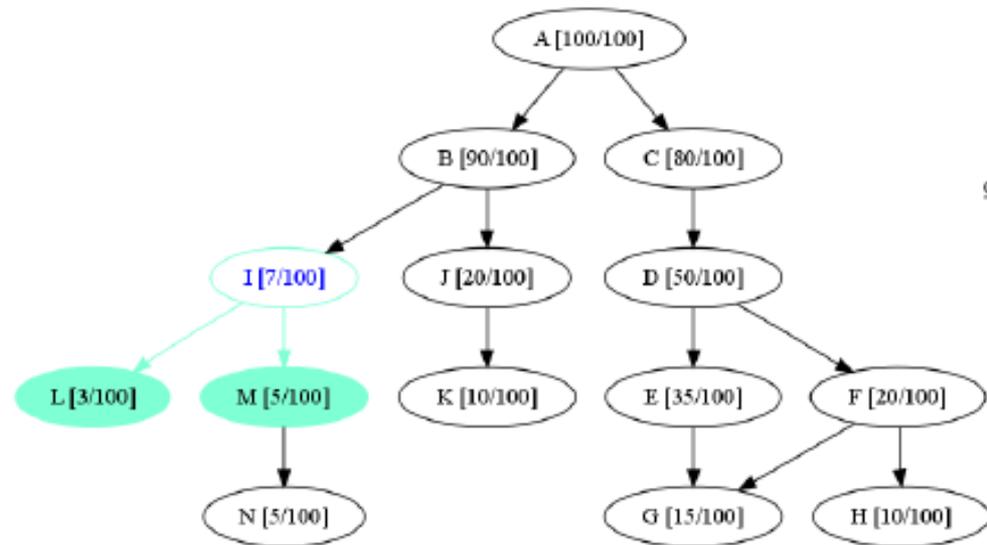
Using the HPO to calculate Phenotypic Similarities

- The importance of a clinical phenotypic finding of the differential diagnosis depends on its specificity
- The specificity of a phenotypic feature, t , is represented by its information content (IC), defined as negative natural logarithm of its frequency of occurrence:

$$IC(t) = -\ln p(t)$$

- The similarity between two terms t_1 and t_2 is defined as the IC of their most specific common ancestor:

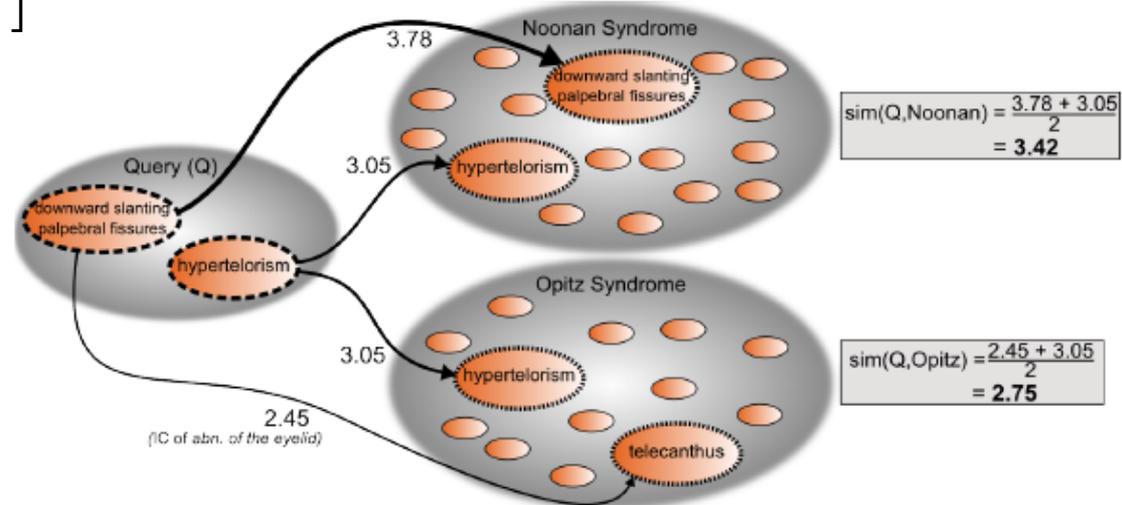
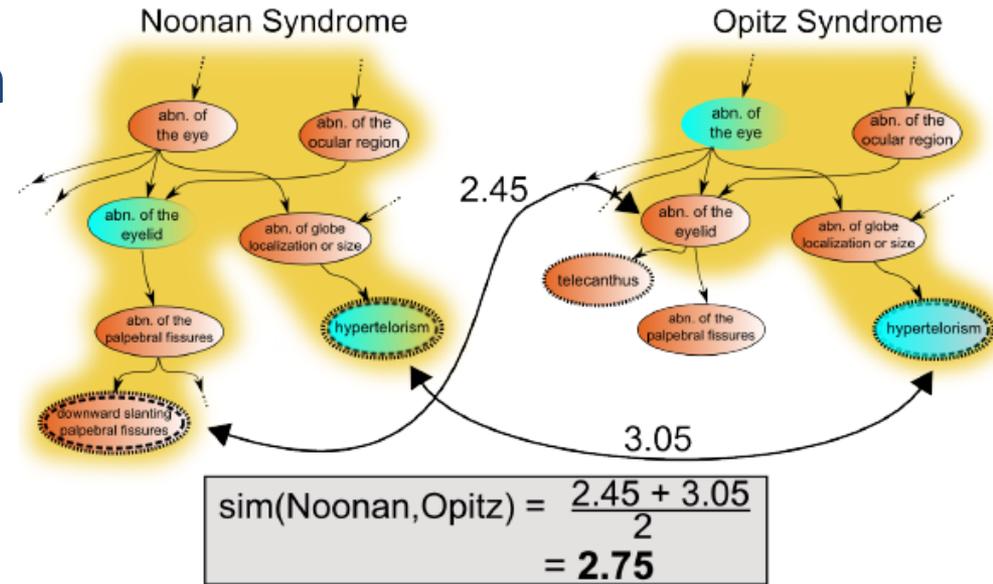
$$sim(t_1, t_2) = \max_{a \in A(t_1, t_2)} (-\ln p(a))$$



Ontological Similarity Search

- Term similarity measures may be used to compute similarity scores between genetic syndromes or phenotypic feature query sets:

$$sim(D_1 \rightarrow D_2) = avg \left[\sum_{t_i \in D_1} \max_{t_j \in D_2} (sim(t_i, t_j)) \right]$$



The Phenomizer*: A Tool for Diagnostics

<http://compbio.charite.de/Phenomizer/Phenomizer.html>

Menu ▾ Help

The Phenomizer - Next Generation Diagnostics

Features. Diseases. Ontology.

PULMONIC STENOSIS search. reset.

HPO Id.	Feature.
HP:0004957	PERIPHERAL, PULMONARY STENOSIS
HP:0001642	PULMONIC STENOSIS

Patient's Features.

HPO.	Feature. ▾	Modifier.
category: ABNORMALITY OF THE EARS (1 Item)		
HP:0000368	LOW-SET, POSTERIORLY ROTATE	observed.
category: CARDIOVASCULAR ABNORMALITY (1 Item)		
HP:0001642	PULMONIC STENOSIS	observed.

Page 1 of 1

Displaying features 1 - 2 of 2

Clear. Mode of inheritance. Get diagnosis.

Köhler S, Schulz M, Krawitz P, Bauer S, Dölken S, Ott C, Mundlos C, Horn D, Mundlos S, Robinson PN: Clinical Diagnostics in Human Genetics with Semantic Similarity Searches in Ontologies, *Am J Hum Genet.* 2009 Sep.

The Phenomizer*: A Tool for Diagnostics

<http://compbio.charite.de/Phenomizer/Phenomizer.html>

The Phenomizer - Next Generation Diagnostics

Menu ▾ Help

Features. Diseases. Ontology.

PULMONIC STENOSIS search. reset.

HPO id. Feature.

HP:0004957 PERIPHERAL PULMONARY STENOSIS

HP:0001642 PULMONIC STENOSIS

Improve differential diagnosis using 5 OMIM entries.

Scoring method: Binary search. Specific search.

HPO id.	Feature.	# Diseases.
HP:0008286	PARTIAL DEFICIENCY OF FACTOR XI	1/5
HP:0004645	TRIANGULAR FACE WITH AGE	1/5
HP:0008357	PARTIAL DEFICIENCY OF FACTOR XIII	1/5

Algorithm: resnik (UnsymmetricNo gram). 3 Features.

<input type="checkbox"/> p-value. ▲	OMIM name.	Genes.
<input checked="" type="checkbox"/> 0.0088	NOONAN SYNDROME 1	PTPN11
<input type="checkbox"/> 0.4608	FRONTOOCULAR SYNDROME	
<input type="checkbox"/> 0.4608	HYPERTELORISM AND TETRALOGY OF FALLOT	
<input type="checkbox"/> 0.5854	CARDIOFACIOCUTANEOUS SYNDROME	BRAF
<input type="checkbox"/> 0.5854	COSTELLO SYNDROME	HRAS, KRAS
<input type="checkbox"/> 0.5854	NOONAN-LIKE/MULTIPLE GIANT CELL LESION SYNDROME	PTPN11
<input type="checkbox"/> 0.5854	EMANUEL SYNDROME	
<input type="checkbox"/> 0.5854	SUBAORTIC STENOSIS-SHORT STATURE SYNDROME	
<input type="checkbox"/> 0.5854	PULMONIC STENOSIS AND DEAFNESS	

HP:0001052 NEVUS FLAMMEUS 1/5

HP:0003251 MALE INFERTILITY 1/5

Improve Differential Diagnosis. Download Results.

Köhler S, Schulz M, Krawitz P, Bauer S, Dölken S, Ott C, Mundlos C, Horn D, Mundlos S, Robinson PN: Clinical Diagnostics in Human Genetics with Semantic Similarity Searches in Ontologies, *Am J Hum Genet.* 2009 Sep.

Patient's Features.

HPO.	Feature. ▲	Modifier.	Num diseas...
[-] category.: Abnormality of metabolism/homeostasis (2 Items)			
HP:0002908	Conjugated hyperbilirubinemia	observed.	13 of 7994
HP:0001942	Metabolic acidosis	observed.	63 of 7994
[-] category.: Abnormality of the abdomen (1 Item)			
HP:0001396	Cholestasis	observed.	125 of 7994
[-] category.: Abnormality of the cardiovascular system (2 Items)			
HP:0001643	Patent ductus arteriosus	observed.	199 of 7994
HP:0004971	Pulmonary artery hypoplasia	observed.	2 of 7994
[-] category.: Abnormality of the respiratory system (1 Item)			
HP:0004971	Pulmonary artery hypoplasia	observed.	2 of 7994
[-] category.: Growth abnormality (1 Item)			
HP:0001508	Failure to thrive	observed.	370 of 7994

Clear.

Mode of inheritance.



Get diagnosis.



Deep Phenotype
Data

Patient's Features.

Diagnosis. ✕

Algorithm: resnik (Unsymmetric). | 6 Features.

<input type="checkbox"/>	p-val...	...	Disease name.	Genes.
<input type="checkbox"/>	0.0080	...	#613404 ARTHROGRYPOSIS, RENAL DYSFUNCTION, A...	VPS33B, VIP...
<input type="checkbox"/>	0.0080	...	%612726 HARDIKAR SYNDROME	
<input type="checkbox"/>	0.0107	...	#208085 ARTHROGRYPOSIS, RENAL DYSFUNCTION, A...	VPS33B, VIP...
<input type="checkbox"/>	0.0160	...	CHOLESTASIS, PROGRESSIVE FAMILIAL INTRAHEPATI...	ATP8B1
<input type="checkbox"/>	0.0160	...	#614582 COMBINED OXIDATIVE PHOSPHORYLATION D...	MRPL3
<input type="checkbox"/>	0.0160	...	#615595 COMBINED OXIDATIVE PHOSPHORYLATION D...	LYRM4, NFS1
<input type="checkbox"/>	0.0160	...	HYPERBILIRUBINEMIA, CONJUGATED, TYPE III	
<input type="checkbox"/>	0.0160	...	BILIRUBIN, SERUM LEVEL OF, QUANTITATIVE TRAIT LO...	
<input type="checkbox"/>	0.0196	...	#610205 ALAGILLE SYNDROME 2; ALGS2	NOTCH2
<input type="checkbox"/>	0.0240	...	#229600 FRUCTOSE INTOLERANCE, HEREDITARY;;FRU...	ALDOB
<input type="checkbox"/>	0.0437	...	CHOLESTASIS, PROGRESSIVE FAMILIAL INTRAHEPATI...	TJP2, ABCB11
<input type="checkbox"/>	0.0454	...	#605711 MULTIPLE MITOCHONDRIAL DYSFUNCTIONS ...	NFU1
<input type="checkbox"/>	0.0456	...	#251880 MITOCHONDRIAL DNA DEPLETION SYNDROM...	DGUOK
<input type="checkbox"/>	0.0489	...	BILIARY MALFORMATION WITH RENAL TUBULAR INSU...	
<input type="checkbox"/>	0.0489	...	#614300 HYPERMETHIONINEMIA DUE TO ADENOSINE ...	ADK
<input type="checkbox"/>	0.0489	...	#614096 COMBINED OXIDATIVE PHOSPHORYLATION D...	AARS2
<input type="checkbox"/>	0.0489	...	SUBJECT SYNDROME	UPT1A

P-value Calculation

- The raw similarity score S depends on the number and specificity of terms both of the query Q and of the diseases D represented in the database
- The distribution of similarity scores can be used to obtain the significance of search results
- The P-value for the null hypothesis that a similarity score of S or greater for a set of query terms Q and a disease D has been observed by chance is defined as:

$$P(s \geq S) = \frac{\text{number of queries such that } \text{sim}(Q, D) \geq S}{\text{total number of possible queries}}$$

Prioritization

Clinical phenotypes -> Disease -> Genes

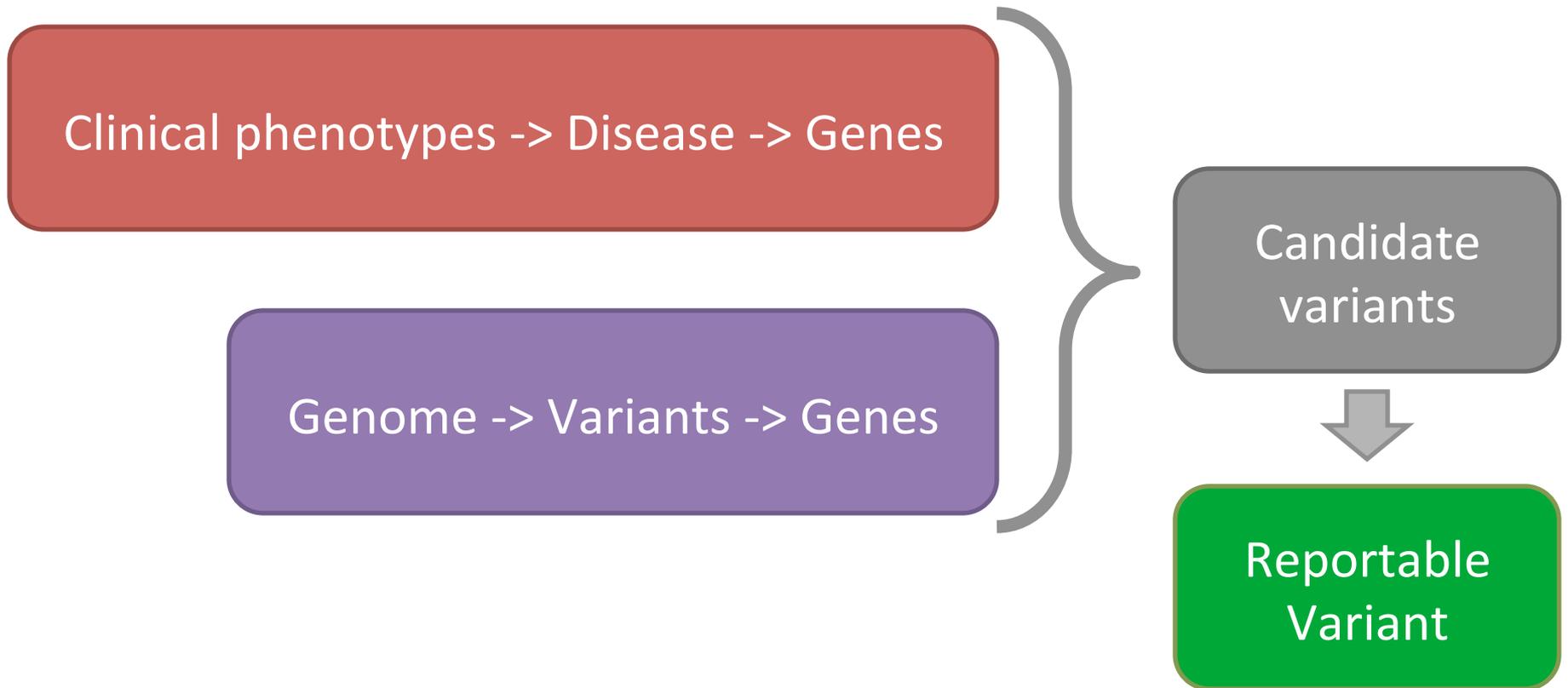
Genome -> Variants -> Genes

Candidate
variants

```
graph LR; A[Clinical phenotypes -> Disease -> Genes] --- B{ }; B --- C[Candidate variants]; D[Genome -> Variants -> Genes] --- B; style B width:0px,height:0px
```

The diagram illustrates two parallel paths for identifying candidate variants. The top path, highlighted in a red rounded rectangle, follows the sequence: Clinical phenotypes -> Disease -> Genes. The bottom path, highlighted in a purple rounded rectangle, follows the sequence: Genome -> Variants -> Genes. Both paths are grouped by a large grey curly bracket on the right side, which points to a grey rounded rectangle labeled 'Candidate variants'.

Now we have a “reportable” variant



ACMG Guidelines

Genetics
in Medicine

© American College of Medical Genetics and Genomics

ACMG STANDARDS AND GUIDELINES

Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology

Sue Richards, PhD¹, Nazneen Aziz, PhD^{2,16}, Sherri Bale, PhD³, David Bick, MD⁴, Soma Das, PhD⁵, Julie Gastier-Foster, PhD^{6,7,8}, Wayne W. Grody, MD, PhD^{9,10,11}, Madhuri Hegde, PhD¹², Elaine Lyon, PhD¹³, Elaine Spector, PhD¹⁴, Karl Voelkerding, MD¹³ and Heidi L. Rehm, PhD¹⁵;
on behalf of the ACMG Laboratory Quality Assurance Committee

Disclaimer: These ACMG Standards and Guidelines were developed primarily as an educational resource for clinical laboratory geneticists to help them provide quality clinical laboratory services. Adherence to these standards and guidelines is voluntary and does not necessarily assure a successful medical outcome. These Standards and Guidelines should not be considered inclusive of all proper procedures and tests or exclusive of other procedures and tests that are reasonably directed to obtaining the same results. In determining the propriety of any specific procedure or test, the clinical laboratory geneticist should apply his or her own professional judgment to the specific circumstances presented by the individual patient or specimen. Clinical laboratory geneticists are encouraged to document in the patient's record the rationale for the use of a particular procedure or test, whether or not it is in conformance with these Standards and Guidelines. They also are advised to take notice of the date any particular guideline was adopted and to consider other relevant medical and scientific information that becomes available after that date. It also would be prudent to consider whether intellectual property interests may restrict the performance of certain tests and other procedures.

https://www.acmg.net/docs/Standards_Guidelines_for_the_Interpretation_of_Sequence_Variants.pdf

Pathogenic

- (i) 1 Very strong (PVS1) AND
 - (a) ≥ 1 Strong (PS1–PS4) OR
 - (b) ≥ 2 Moderate (PM1–PM6) OR
 - (c) 1 Moderate (PM1–PM6) and 1 supporting (PP1–PP5) OR
 - (d) ≥ 2 Supporting (PP1–PP5)
- (ii) ≥ 2 Strong (PS1–PS4) OR
- (iii) 1 Strong (PS1–PS4) AND
 - (a) ≥ 3 Moderate (PM1–PM6) OR
 - (b) 2 Moderate (PM1–PM6) AND ≥ 2 Supporting (PP1–PP5) OR
 - (c) 1 Moderate (PM1–PM6) AND ≥ 4 supporting (PP1–PP5)

Likely pathogenic

- (i) 1 Very strong (PVS1) AND 1 moderate (PM1–PM6) OR
- (ii) 1 Strong (PS1–PS4) AND 1–2 moderate (PM1–PM6) OR
- (iii) 1 Strong (PS1–PS4) AND ≥ 2 supporting (PP1–PP5) OR
- (iv) ≥ 3 Moderate (PM1–PM6) OR
- (v) 2 Moderate (PM1–PM6) AND ≥ 2 supporting (PP1–PP5) OR
- (vi) 1 Moderate (PM1–PM6) AND ≥ 4 supporting (PP1–PP5)

Benign

- (i) 1 Stand-alone (BA1) OR
- (ii) ≥ 2 Strong (BS1–BS4)

Likely benign

- (i) 1 Strong (BS1–BS4) and 1 supporting (BP1–BP7) OR
- (ii) ≥ 2 Supporting (BP1–BP7)

Uncertain significance

- (i) Other criteria shown above are not met OR
- (ii) the criteria for benign and pathogenic are contradictory



Strong

Supporting

Supporting

Moderate

Strong

Very strong

<p>Population data</p>	<p>MAF is too high for disorder BA1/BS1 OR observation in controls inconsistent with disease penetrance BS2</p>			<p>Absent in population databases PM2</p>	<p>Prevalence in affecteds statistically increased over controls PS4</p>	
<p>Computational and predictive data</p>		<p>Multiple lines of computational evidence suggest no impact on gene /gene product BP4</p> <p>Missense in gene where only truncating cause disease BP1</p> <p>Silent variant with non predicted splice impact BP7</p> <p>In-frame indels in repeat w/out known function BP3</p>	<p>Multiple lines of computational evidence support a deleterious effect on the gene /gene product PP3</p>	<p>Novel missense change at an amino acid residue where a different pathogenic missense change has been seen before PM5</p> <p>Protein length changing variant PM4</p>	<p>Same amino acid change as an established pathogenic variant PS1</p>	<p>Predicted null variant in a gene where LOF is a known mechanism of disease PVS1</p>
<p>Functional data</p>	<p>Well-established functional studies show no deleterious effect BS3</p>		<p>Missense in gene with low rate of benign missense variants and path. missenses common PP2</p>	<p>Mutational hot spot or well-studied functional domain without benign variation PM1</p>	<p>Well-established functional studies show a deleterious effect PS3</p>	

Fortunately...

- http://www.medschool.umaryland.edu/Genetic_Variant_Interpretation_Tool1.html/

The screenshot shows the top navigation bar of the University of Maryland School of Medicine website. It includes links for SOM Home, QUICK LINKS, EMAIL, MYUMB, DIRECTORY, MAP, and CALENDAR, along with a Site Search box. Below the navigation bar is the University of Maryland School of Medicine logo. A yellow banner contains links for About Us, Departments, Research, Education, Patient Care, Faculty, News, and Alumni & Donors, along with a Give button. The main content area features the title "Program for Personalized and Genomic Medicine" and a breadcrumb trail: Programs > Program for Personalized and Genomic Medicine > Implementation Projects > Genetic Variant Interpretation Tool. A left sidebar lists navigation options: Home, Leadership, Administration, Program Members, Graduate Programs, and Shared Resources. The main heading is "Genetic Variant Interpretation Tool". The text below the heading states: "To aid our variant interpretation process, we created an openly-available online tool to efficiently classify variants based on the evidence categories outlined in the article: Richards, et al. *Standards and guidelines for the interpretation of sequence variants*. 2015. This site displays the evidence categories and descriptions from Table 3 and Table 4 with simple checkboxes for selecting appropriate criteria. The site then incorporates the algorithm in Table 5 to automatically assign the pathogenicity or benign impact based on the selected evidence categories. Since our process often requires analyzing multiple variants per patient, we have also allowed the option of aggregating each variant into an exportable table at the foot of the website for easy documentation of the variant review process for our records. Although this tool is based on the text of the variant evidence has been pulled directly from Richards, et al. *Genet Med*. 2015 May;17(5). This site does not claim authorship of any of the variant

What if you are unlucky?

Clinical phenotypes -> Disease -> Genes

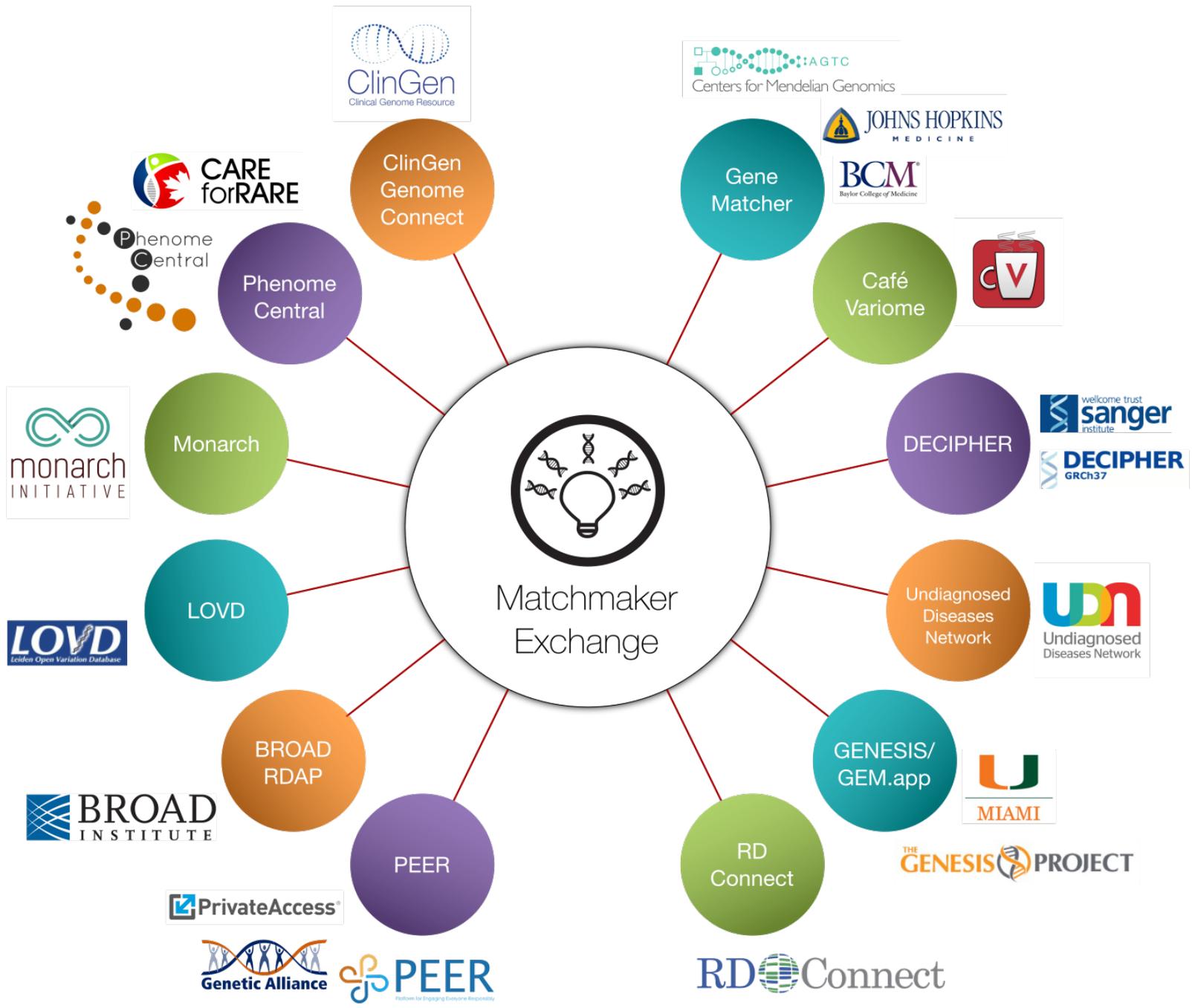
Genome -> Variants -> Genes

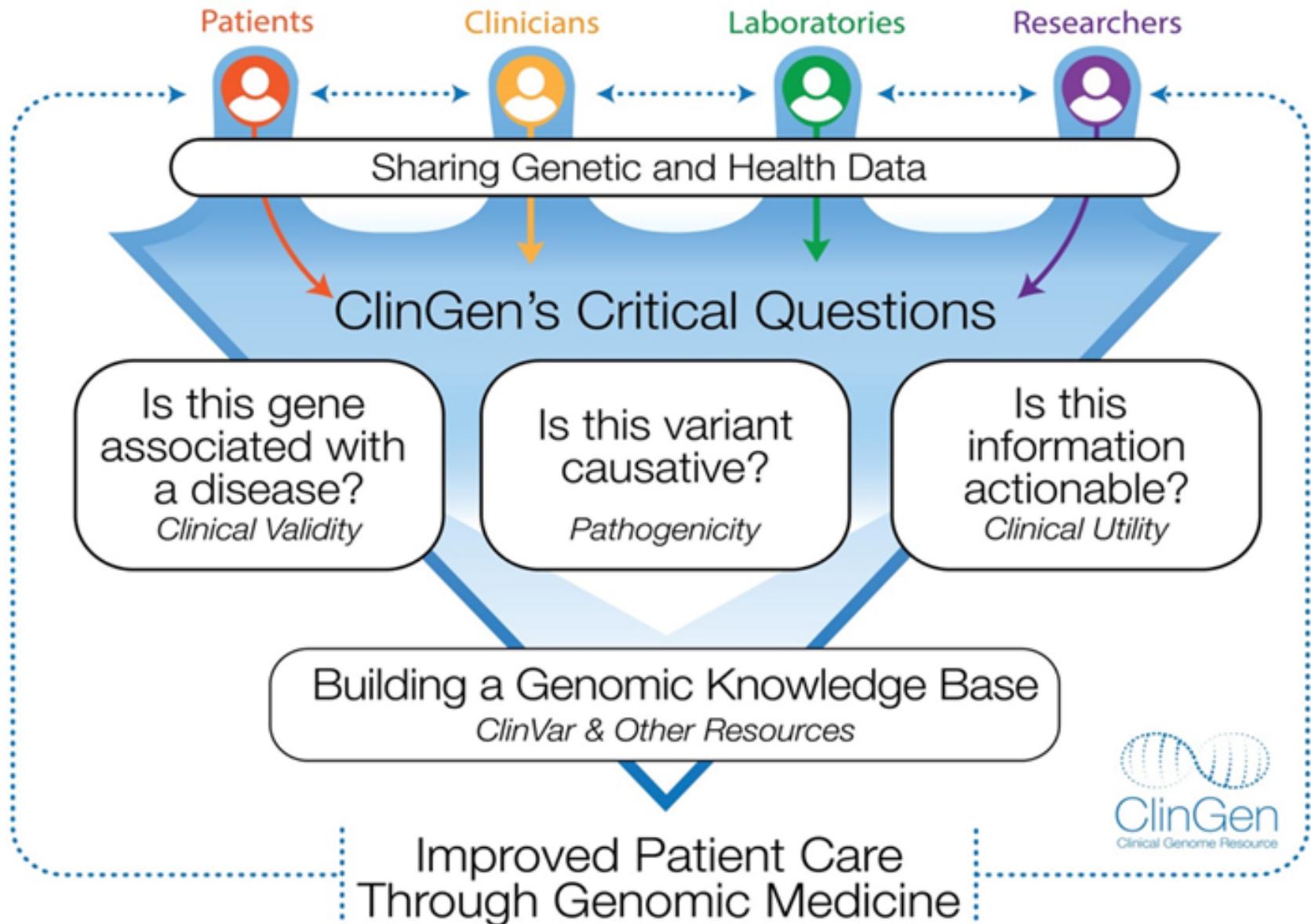
~~Candidate
variants~~

Matchmaker Exchange

 Genomic discovery through the exchange of phenotypic & genotypic profiles



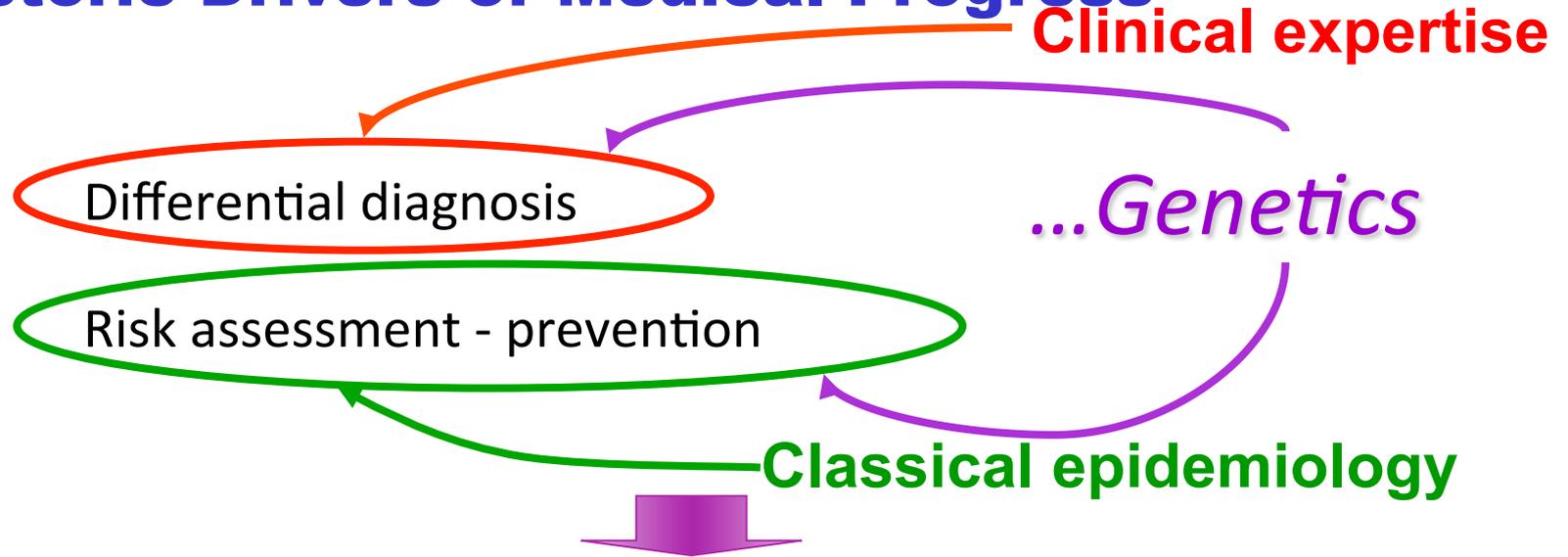




Just the start.....

Medical Progress: Evolution or Revolution?

Historic Drivers of Medical Progress



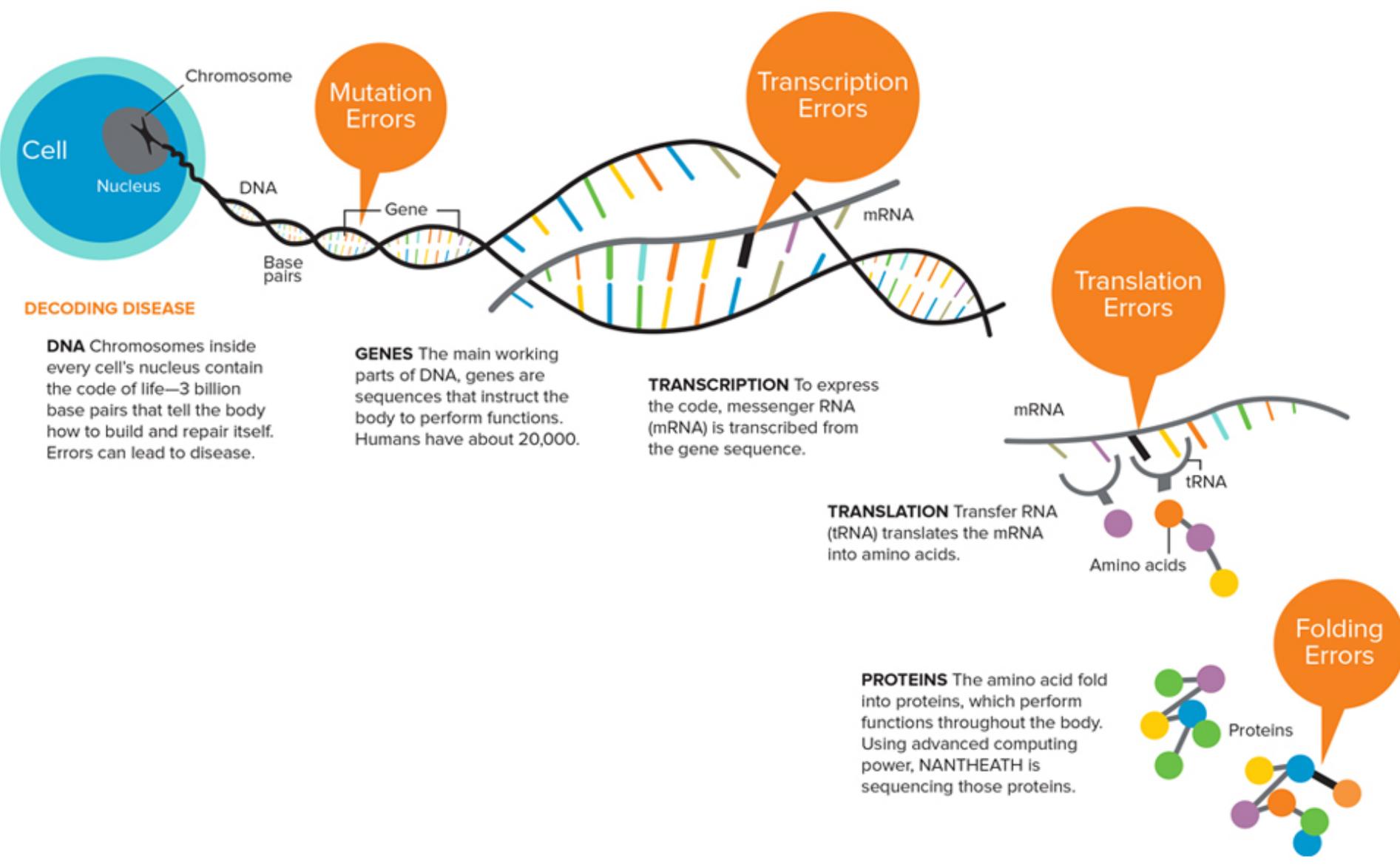
More differentiated, *molecular* understanding of pathology and drug action

Clinical Disease Definition
Clinical Diagnosis



Molecular Disease Definition
Molecular Diagnosis

in-vitro Diagnostics



DECODING DISEASE

DNA Chromosomes inside every cell's nucleus contain the code of life—3 billion base pairs that tell the body how to build and repair itself. Errors can lead to disease.

GENES The main working parts of DNA, genes are sequences that instruct the body to perform functions. Humans have about 20,000.

TRANSCRIPTION To express the code, messenger RNA (mRNA) is transcribed from the gene sequence.

TRANSLATION Transfer RNA (tRNA) translates the mRNA into amino acids.

PROTEINS The amino acid fold into proteins, which perform functions throughout the body. Using advanced computing power, NANTHEATH is sequencing those proteins.

Mutation Errors

Transcription Errors

Translation Errors

Folding Errors

Cell

Nucleus

Chromosome

DNA

Base pairs

Gene

mRNA

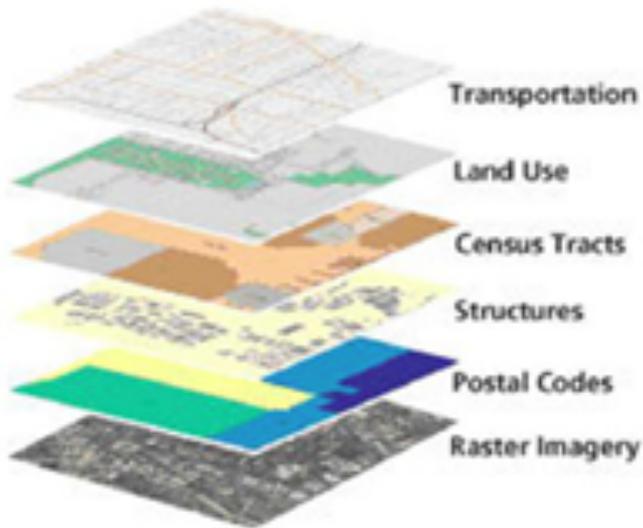
mRNA

tRNA

Amino acids

Proteins

Google Maps: GIS layers Organized by Geographical Positioning



Information Commons Organized Around Individual Patients

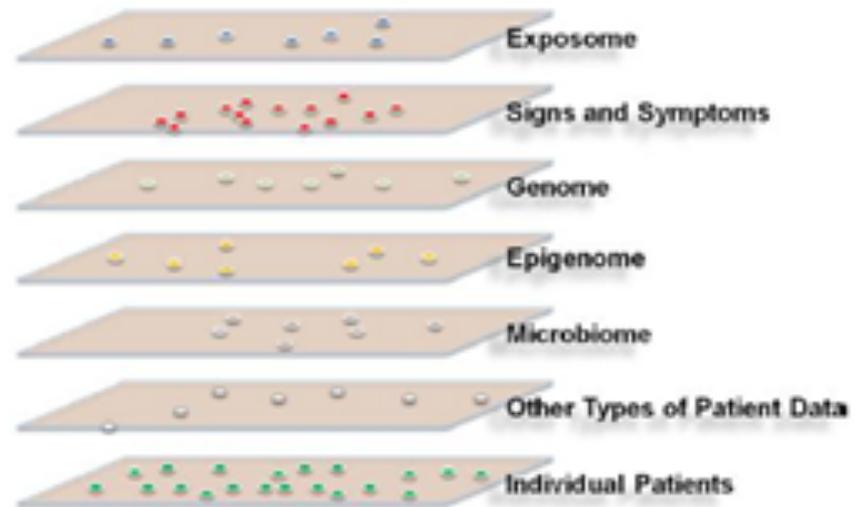


Figure 1-2: The proposed, individual-centric Information Commons (right panel) is somewhat analogous to a layered Geographical Information System (left panel). In both cases, the bottom layer defines the organization of all the overlays. However, in a GIS, any vertical line through the layers connects related snippets of information since all the layers are organized by geographical position. In contrast, data in each of the higher layers of the Information Commons will overlay on the patient layer in complex ways (e.g., patients with similar microbiomes and symptoms may have very different genome sequences). Source: FPA 2011 (left panel).