

NanoPipe – interactive tool for MinION sequencing analysis

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Victoria Shabardina

Nanopipe is a **web-based** tool for **fast and easy** processing and analysis of the **Minlon** sequencing data.

Created in the Institute of Bioinformatics, University of Münster, Germany

Wojciech Makałowski

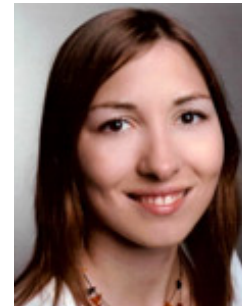


Norbert Grundmann

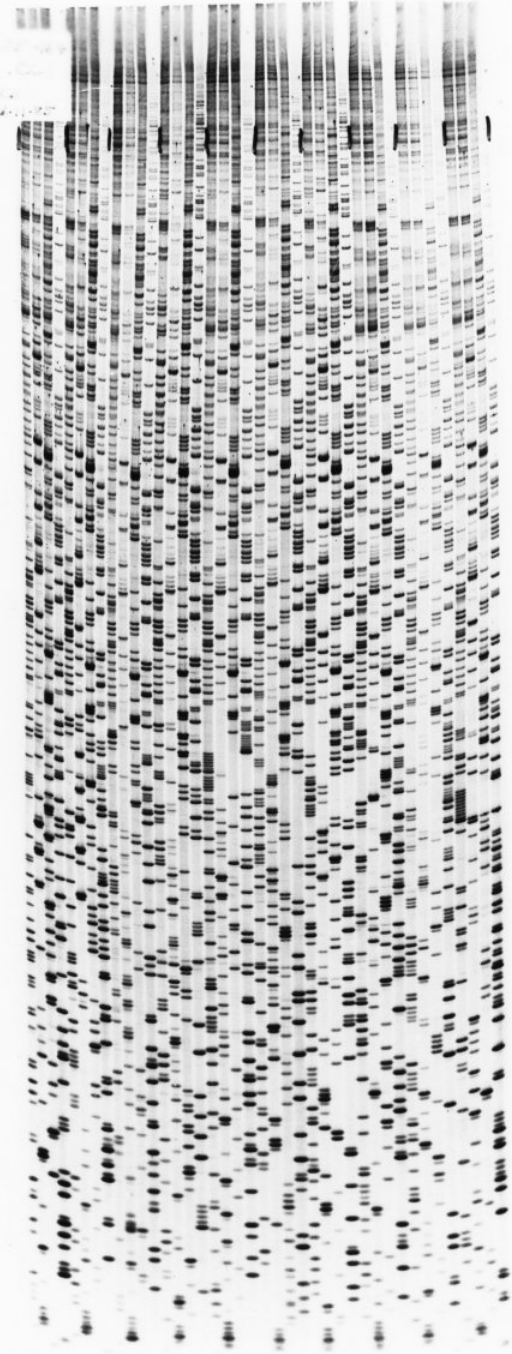


Victoria Shabardina

Tabea Kischka



1977



Elegance of the technical progress ;)

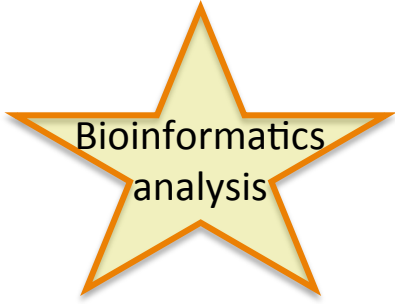
2017



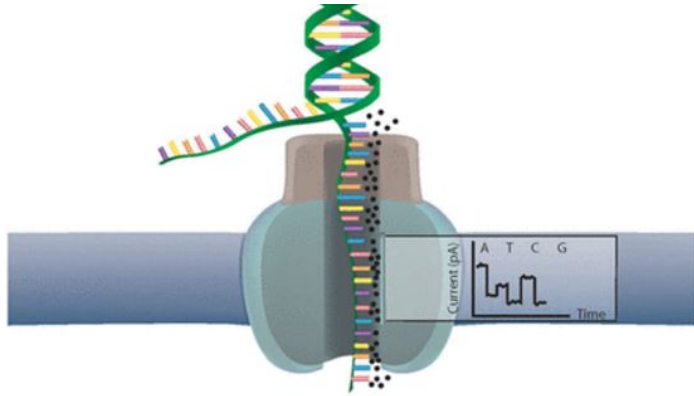


Real-time MinKnow
Albacore basecalling

HDF viewer

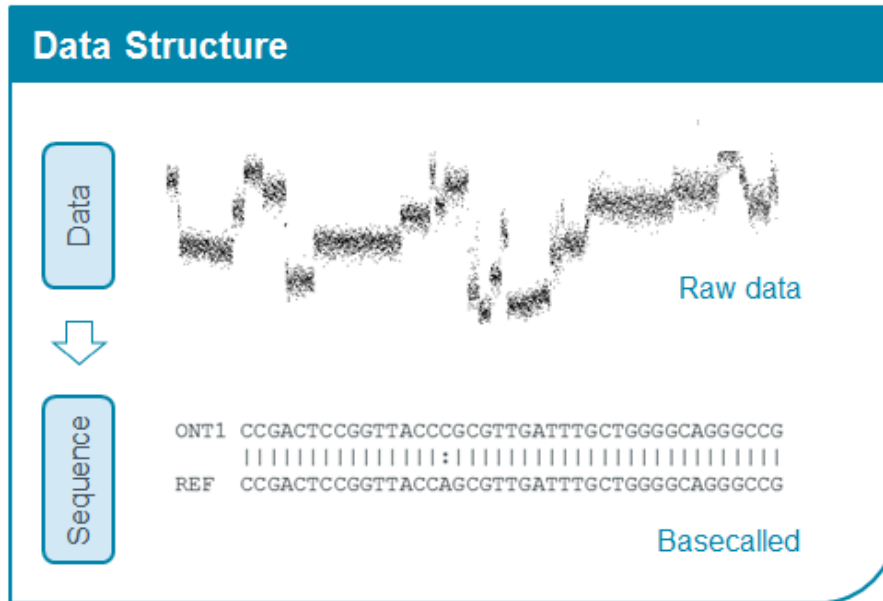


Basecalling for MinION sequencing reads



MinKNOW detects events that are measurements of 5 subsequent nucleotides

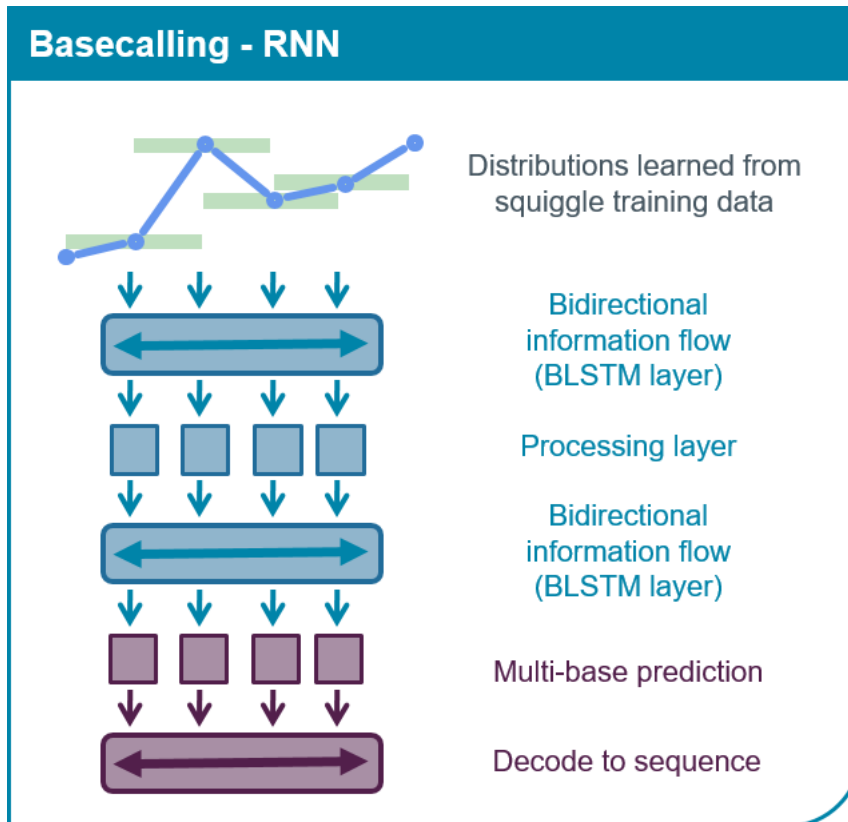
→ Albacore does basecalling:



*change in current duration of the signal noise signal

Basecalling for MinION sequencing reads

MinKNOW, Albacore use Recurrent Neural Network mechanism: it can learn and is trained on multiple data from MinIon sequencing



→ one .fast5/.fastq file per one read

.fast5 file format

.fast5 is a type of the Hierarchical Data Format (HDF) and designed for the storage of big datasets

It is binary – not readable by human

[HDFview](#) – the tool to see HGF files:

.fastq file format

Each entry consists of 4 lines:

@header

sequence

+

quality (coded by ASCII symbols: https://en.wikipedia.org/wiki/FASTQ_format#Encoding)

[Poretools](#) is the tool developed for MinIon to convert .fasta5 files to .fastq or .fasta files

Recently Albacore basecaller has allowed to create directly .fast5 or .fastq files 😊

How HDFview looks like:

<https://support.hdfgroup.org>

.fastq file
(contains base quality info)

Raw signal
(current from a Minlon channel)

HDFView 2.13

File Window Tools Help

Recent Files | 4.2017/data/reads/pass/0/iobl_goshawk_20170410_FNFAB45819_MN19777_mux_scan_SQR170407_54198_ch103_read29_strand.fast5 | Clear Text

Analyses

- Basecall_ID_000
 - BaseCalled_complex
 - Fastq
 - BaseCalled_template
 - Fastq
 - Summary
 - basecall_1d_corr
 - basecall_1d_tem
- Segmentation_000
 - Summary
 - segmentation
- Raw
 - Reads
 - Read_29
 - Signal
 - UniqueGlobalKey
 - channel_id
 - context_tags
 - tracking_id

Text

Data selection: [0] ~ [0]

```
@channel_103_062f2402-85b2-4f63-8e73-41995ce0b3d2_template
ATCAATGCTGTTTCGTTTGGTGCTACTTGCTGTCGCTCGTACAGTCTGCTGGGGCTTCCATTGGCTCTGAAGG
ATTATAAGTCCCGTCAATTTGTGCTACGGACAACCTTTGAGAAGTCTTTCTGAAGATGATGATGATTGAAGACCT
GGATGCTACGGAGAACCTTGATGGCAGCTCCTCTTCAGGTGATAGTCACTCTTATAGTCTCACTCTTTGAGTTAC
TTGACAACCTCTGTACCTTTCTTAACCTTTGTGACAGAAAAGGATGGCACCCTCGAATACTTTTGCTGTTGGAAAG
TTCACTATGACTATAAAGCAAAGCAGTCAACTAGATGCTAAATCTGGAATATATGTCACATGGTACTGGTATAGCT
GCCAGTCTGGGGAAAATCTAAGTTAAAGAGGTTTGAACAGTTCACAGGTATTCCACTGGAACCTGTTTGA
ATTTGGACTTCTGTACATCGGCCTAAACAGGAGCTTCGCTTCAAATATATGACACAATGGCAAACCCAGGCC
AAGGGATAATTTGTGCTGGGGAATTTCCATTTGGCTCTGATTATAGGGGATTCCTCGTCAATTTGGTACATGACAAC
TTTTGTCTTCTAGAAGATGATATTGAAGACCTGGATGCACGGGAGGAGTGGCAGCAATATCCTTTCAACAGAA
AACGTTCTGTTTATGTTCTTAGACACTGGTGTACAGT
+
(*)(%&((,))(*("1/2//3/-.--*+,'-+,-+&")()*+*,--+4-/5:80)-+85;<9<74114334<512./5<;8993233;
+-.*23433241),0220-/-05/255=:46555543531./13412802;33::8771'+),/.,+1-5*(+1/50+./,231
3;5<017-598/*2+,,/6=46:<9631102244<=6500::/))0215;55<84::9<71;.6.*-****("/393-1630+8
8=<<710(&+)*88-,9;<*),/-97785589842699;<<80+))'-0/'0+,*')+1.24421049317==<=<6;79
799689<9<=<<8368;9<<=<<=<<420)/4-/-,')-111141</).066=91/3-69677;82/,*1.3=<::89<
==<<83.,1/*0).1/66;=-)*.(')*(+ + -))/489<8(4-0--(/4-0)+4+068778;00888:.0+4.01348+,.01./64*6
9-8;<;5.2+.-(*)/6-403>944(2.+4972-/-/32/*-/0+845./6/52:36401/1,-./1.,4:5./-/9972))-1220
22322::22;33==<=<62,')-91,124+..-(/347=85-, '&'1))(*"/1468462222>=>=<=<=<-3-)-11*/20)/)*
(+ '&&
```

Signal at /Raw/Reads/Read_29/ |iobl_goshawk_20170410_FNFAB45819_MN19777_mux_scan_SQR170...

Table

0-based

0	677
1	638
2	589
3	562
4	554
5	570
6	586
7	581
8	573
9	562
10	576
11	578
12	554
13	564
14	559
15	577
16	571
17	579
18	583
19	571
20	568
21	578
22	567
23	568

Fastq (46173, 2)
String, length = 1502, 1
Number of attributes = 0

Log Info Metadata

.fasta file format

contains 2 lines: >header and sequence

```
>ff6c98dd-bce9-4a2b-bf13-081841413c94_Basecall_2D_minion_20170511_Ae_aegypti  
GCGCTGGTTCAGTTACATATTGCTAGGGTTAAGCAGTGGTGACCACAGATTTTTATGATTTATGGATT  
CTTTTCTTCTGGCTACATTACTGGAACAGAGCCTGCTTCTCAACAGTGTTCTTATGAACGCTTCAGCTTA  
GTATAAAGGC
```

```
>4c8a2487-0e13-41a6-ac7b-6cd2fbf5eb95_Basecall_2D_minion_20170511_Ae_aegypti  
TACGCGGTGACAAAACGTGCGTACCGGCAACCGCATGTTGAAACAGGAAAACGTACAAAGGACCC  
TCGCAAAATGCGCGACAAAATCTGCAACGTACAACATGCGATAAACGTGCGTGAGGAGATC  
>.....
```

> sing is convenient marker to brows through the .fasta files

Nanopipe from Münster: why it is a pipe?

```
pyra.uni-muenster.de - PuTTY
CCTGGTCAACGGCCAGCCGACAGCGCGCTGAACTCGGCGGAGTGGCAGGCTCCGCGG
TGAGTTCAACGGCCAGCCGCGCAATCTTCAATCAACACGGTAGGAAAGTGGAGTAGGAA
CGCGCCAGCTGGTTCGAGCCGAGTTCAATCGCCAGGAAGCAGCCGGCGCCTTCTCGCCT
GAGCAGCATGCTGAAGACACCCGCGTACAGTTCTGCTCCTCGAGCTCCACTCCAGGAAC
AAGCCGAGAATTTCTGTTGGTGTCTGATGTCTATACCTGGCCGGGGGTGACGAAGTCAGG
TTGATTCGGGAGAACCTGAGCAGCTACCACATCAAGGAAGTAGCAGACTTTAACATGAAG
ATAGAGCGACGGCGAAGTAGCAATGTCCCG
>channel_95_914f9b6b-d481-4a4e-8133-b8c3b3a94619_complement pass_april/11/iobl_goshawk_20170410_FNFAB45819_MN19777_se
TCGAGCATAAACAGCAGCGCCATATCTTCAATCAACACGGTAGGAAAGTGGAGTAGGAA
ATACTACAAGTGGAGCCTGACCCGACAAATACTACCTCAACGGGAAGATCAATAAATTTGT
TATATCATATTGATGGTATATACTCTCAATACAATAGTGCCAATTTCTGGTCTTTCTCC
CAAAACCCCTTAACCTAGTCTAATCGTAGTAAATATCAGAAAAATTTCTCAAAAATTTAAAA
TACCTGATCAGTATTTGCAAAATGTCAGTGCATCAAAACTRAACTCTGGATATTGATGCA
GCCAAGTTTGAAGATATATAGCCTAAGTAAAGTTTATTAAGTTAAAGATGGGAACCA
CAAAGCTAGAAAAATATTAGGGTTTTATTTTATAATGGGAACCTTTCTAAGTGTATGACAG
CATATAGAAAAATCAAGAAAAAGAGTTTATTTTATTTTCTAAAAATTAATAACTTTTGGAGC
AGTTTAGGTTTGAAAAAATGAGTGGCTTCGCTACTCTCTCACCCACCCTTCTCTACTA
ATATGCGCATTGTATAGGCATTGTTACAGCAGATGTGCCAATTTGATCCACTATTATTAAC
AAAATTCGGCCAGGCATGGTGTACCTATGTATCAACACTGTAGAGCCGGAAAGTGGATC
ACGAGGTGAGGAAGTTCAAAACAGACTGGCAAGTGGTGAACCCCGCTACTAAAAATACAA
AATTAGCCGCATAATGCGACCAATATCAGCATRATGCCATGCT
>channel_105_3256ac6f-4f29-440f-8bec-a480c1b7a64f_complement pass_april/11/iobl_goshawk_20170410_FNFAB45819_MN19777_se
CTATCAAGCATATGGTAAATATTGTCACACCTGGTAGGAAGATGAGGTGAGACTATGG
TCAGGGCTGCCAGGAACCAACATGAGGTTGGGTCAGGCTGAGCCTGGAGTGAAGGGT
AGTGTGGATAGCAAGCATCGGATCTCTCTCTCTGCTGAAGAAAGGGCACTTGTGAGG
TCTCATTAACCCAGCTCAGTTATGCTTTGATGTGCTGTGCTTTACGCCAGATTGGC
CTTGGTTTGAACACAGATGCGAGATGACTGAACAGATGAGGAAGTTCACCTGCAAGGTGGG
GCCAAGTTGGGGCACCAGCCAGCCTATCATAGCTGTATACAGCCCTAATGGCRATATC
CTTTACCAACAAATTTTCAGTGGCTTTGATTCCACATTAAGTGTGCAAAATGCTAAT
CGTGATAATTTAATAGCATTGTGAGATCAGCTGATAAATGATAAATGCTTCTCTGSC
CATGTGTGTGTTGAAAAGGACCTGTTAAAATGTTTGTAGTGAAGAAATTTGAATCCAGTATC
TGTTAATATAGCTACATGAGTCTTCTCACATGAAGGCAGAAATATACTTCTCATATGA
TTTATTTCTTACTTAAATAAACATTTAGTAAAAATGATGACGCCAGAGCGGTAATCCT
TGGCTGTCTTTTAAATATAGAACTAAAGAAATTAACATGATGTTATTTCTTTCTTTT
CTAAGTGAAGRAATGTTAAATAGCAACACTTGTAGTTTGGACAGTTTGGAAATTCAT
GCTAGTTACTGATGAACATTTTGTATTTTCCATATTTTCAATTTGCTATTGGAAACAGT
AGTTTGTGTTGACTGTACCCAGCTCTGGCAAGATATAATAGGAGTAGGAATTTGCCTGC
ATGGATACTTGTGTAGTACATGGGCTTGTAGTACCTTTTATTTGCAATGTTTCAACT
TCCTCTGGGAAGTGTCCAAATATCTCCCAATCTCTGCTCGGACCCCTTCTTCTCACGAG
ATTCCCTTCCCTACCGTGTGATGAGGATAAGTA
[vickas@pyra-work:KlausLabSequencing] > less readme_analysis_protocol.txt
author: vika
date: 20.04.2017
project: MinIon sequencing of human genes mutated in cancer
file content: description of the analysis procedure

*pass anf fail folder were around 4.5 GB; the sequencing protocol was run for 48 hrs; start: 480 pore were avail
flowcell.

1)poretools fastq --type fwd,rev /pass > pass_10.04.2017_fwdrev.fq
the same for the fail
2)Ayako Suzuki: "Using LAST (version 658) with parameters -a13, -A14, -b4 and -B3 which were determined by last-train
all of the 2D reads were mapped to chromosome 7 of the human genome UCSC hg38. The best alignments for each query wer

What I did:
lastdb hg38db /HG38/HG38.fasta
last-train hg38db pass_10.04.2017.fasta #the SDTOUT is saved inder "last-train 21.4.2017.txt" in the directory with t
lastal -a12 -A19 -b3 -B3 -S1 hg38db pass_10.04.2017_fwdrev.fasta | last-split > pass_10.04.2017_hg38_lasttrained.maf
maf-convert sam -d pass_10.04.2017_hg38_lasttrained.maf > pass_10.04.2017_hg38_lasttrained.sam
samtools view -bS -o pass_10.04.2017_hg38_lasttrained.bam pass_10.04.2017_hg38_lasttrained.sam
samtools sort -o pass_10.04.2017_hg38_lasttrained_sorted.bam pass_10.04.2017_hg38_lasttrained.bam
samtools index pass_10.04.2017_hg38_lasttrained_sorted.bam
-->go to IGVviewer
```

```
#
sub skipMinlen {
    return if (1-r "$INPUTPRAEFIX.minlen");
    my $minlen = readFile("$INPUTPRAEFIX.minlen");
    return if ($minlen < 1);

    my $stype = readPraefix($QUERYFILE, 1) eq ">" ? 1 : 2;

    rename($QUERYFILE, "$QUERYFILE.tmp");

    open(IN, "<", "$QUERYFILE.tmp");
    open(OUT, ">", $QUERYFILE);

    if ($stype == 1) {
        my ($header, $seq);
        while (my $line = <IN>) {
            if ($line =~ m/^\s/) {
                if ($seq) {
                    my $len = $seq =~ tr/A-Za-z//;
                    print OUT $header, $seq if ($len >= $minlen);
                }
                $header = $line;
                $seq = "";
            }
            else {
                $seq .= $line;
            }
        }
        if ($seq) {
            my $len = $seq =~ tr/A-Za-z//;
            print OUT $header, $seq if ($len >= $minlen);
        }
    }
    else {
        my $index = 0;
        my $firstline;
        my $print = 0;
        while (my $line = <IN>) {
            if ($index == 0) {
                $firstline = $line;
            }
            elsif ($index == 1) {
                my $len = $line =~ tr/A-Za-z//;
                $print = $len >= $minlen;
                print OUT $firstline, $line if ($print);
            }
            else {
                print OUT $line if ($print);
            }
            $index = ($index + 1) % 4;
        }
    }
}

close(OUT);
close(IN);
}
```

Web interface



- About
- Usage
- Run the Pipeline
- Contact

NanoPipe New Request

Previous Request

ID

New Request

Target

Query File no file selected

Minimum Sequence Length

Title

Last Parameters

Substitution Matrix

Use Matrix or Match Score / Mismatch Cost

	A	C	G	T
A	6	-19	-8	-18
C	-19	7	-20	-9
G	-8	-21	7	-19
T	-19	-10	-19	6

Gap Existence Cost (-a)

Gap Extension Cost (-b)

Insertion Existence Cost (-A)

Insertion Extension Cost (-B)

Score Matrix applies to Forward Strand (-S)

Query Letters per Random Alignment (-D)

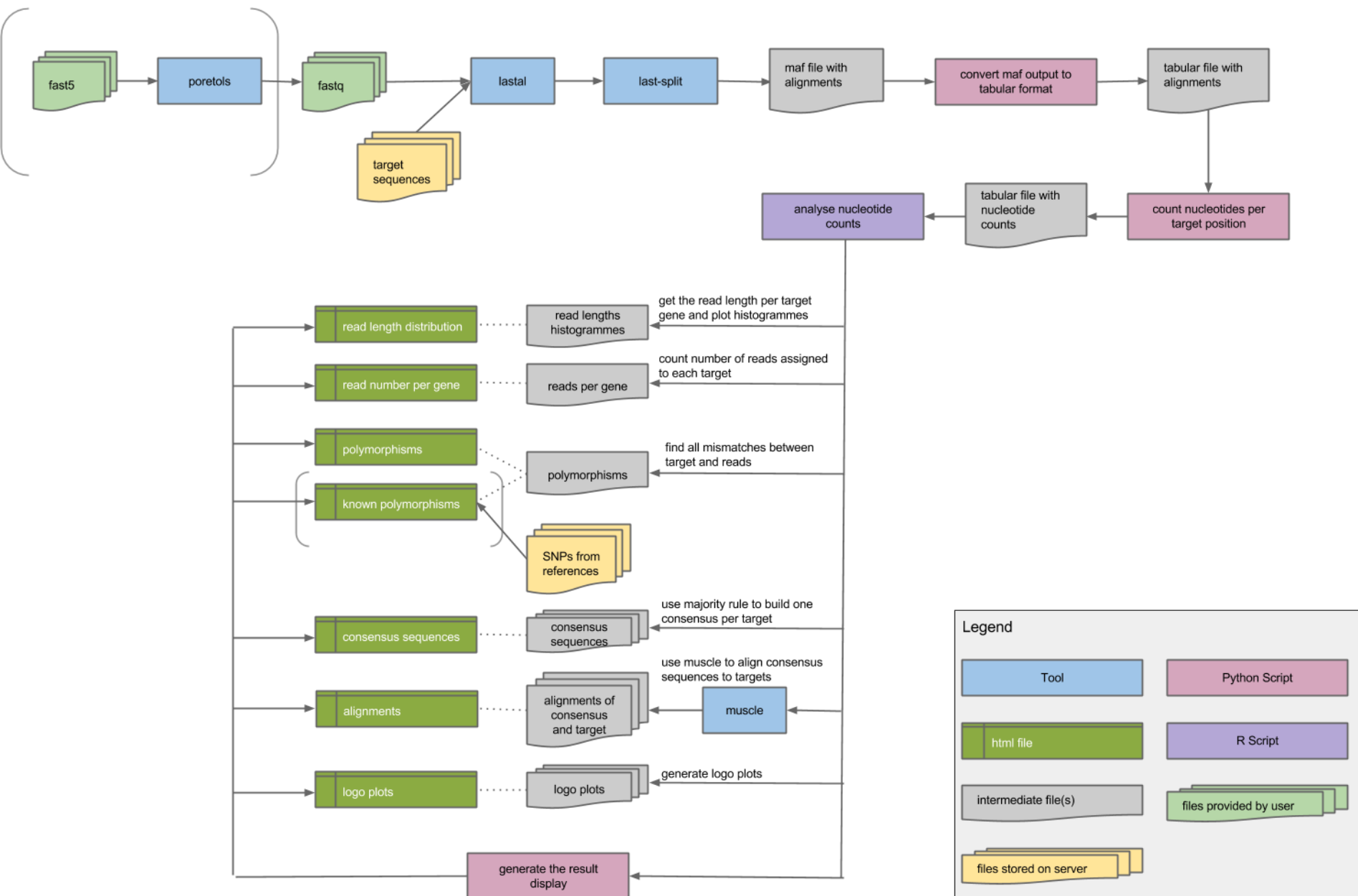
Porettools Parameters

Read type

Minimum read length

Quality filter

Nanopipe's workflow

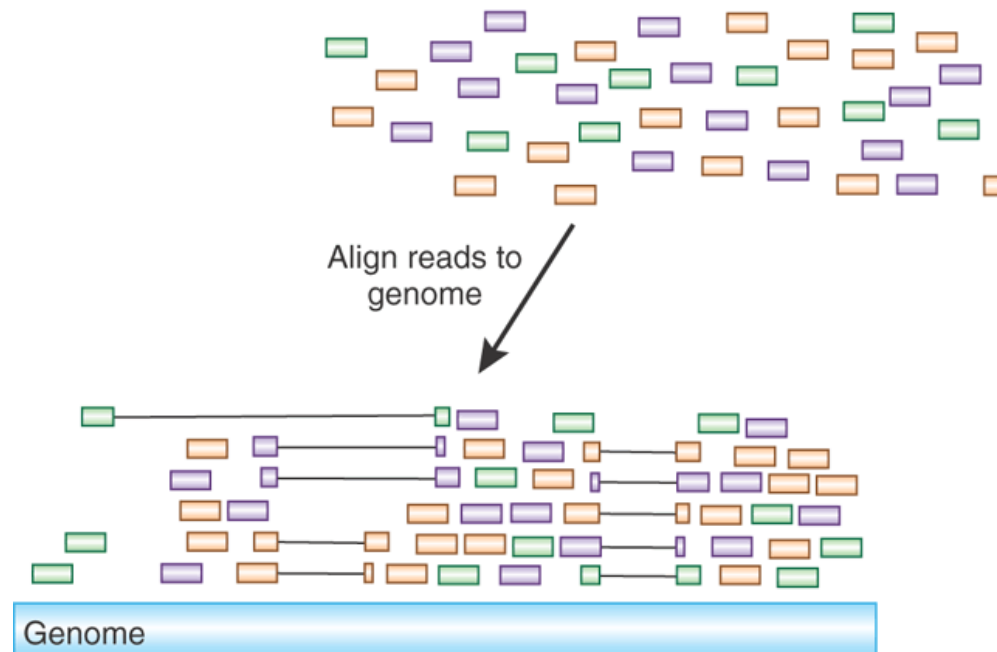


Key steps of Nanopipe

1) LAST sequence aligner maps Minlon-produced reads to a target (selected region, exon, gene, genome)

Martin Frith

University of Tokyo,
Division of Biosciences



Other aligners: BLAST (psi-BLAST, delta-BLAST), HMMER, MUSCLE, MAFFT...

File formats that are important to know when working with mapped reads:

.maf

.sam

.bam

.maf file format – shows pairs of aligned sequences with the coordinates

```
# LAST version 828
#
# a=12 b=3 A=19 B=3 e=31 d=23 x=30 y=9 z=30 D=1e+06 E=725.241
# R=10 u=0 s=2 S=1 M=0 T=0 m=10 l=1 n=10 k=1 w=1000 t=0.910239 j=3 Q=0
# readsDB
# Reference sequences=340931 normal letters=689426062
# lambda=1.10061 K=0.337715
#
#   A C G T
# A  1 -1 -1 -1
# C -1  1 -1 -1
# G -1 -1  1 -1
# T -1 -1 -1  1
#
# Coordinates are 0-based.  For - strand matches, coordinates
# in the reverse complement of the 2nd sequence are used.
#
# name start alnSize strand seqSize alignment
#                               Seq.name| Align.start| Align.length| Seq.length
# batch 0
a score=1405 EG2=0 E=0
s 6e5acca3-80ce-43d6-ac9a-6f63dfe61404 4582 2456 + 7060 TAACGCCATTACCTACAAAAGCCAGCGCGACAAAATGCCAGAGAAGCTGAAGCTGGCGAACGCGGCAATT
GTGCATCAGTGCATTGAAGCCACCACCGCCTCCGGCGTGGATAATGCAACCTCCCCCGACTTGAAGTACGCTGAACGGGATTATTTACCCTCAGAGAGAATTTGATCACTATGCAAAA-CAAC
TTT---ACGAATGTTTGGGTTTCTGTTTAAACAACATTTTCTGCGCCGCCACAAATTTTTGGCTGCATCGACAGTTTTCTTCTGCCAATTTGAAACGAAGAAATGATGAATTGATGTGGTTTCCTTT
GTAGAAAATTAACAAACCCTAAACAATGAGTTGAAATTTTATATTGTG--TATTTATTAATGTATGTCAGGTGCGATGAATCGTCATTTGTATTCCCGGATTAACATATGTCCACAGCCCTGAAG--
TGTTCTGAATGCTCTCAGTAAATAGTAATGGGT-ATCAAAGGTATAGTAATATCTTTTATGTTTATGTTGATGTTGTAACCCATCGGAAAACCTCCTGCTTTAAGATT-TTTTCCCTGTATTGCTGAAA
TACAGATAGTAAATATAATGTGAGACGTTGTGACGTTTTAGTTTCAAGATAAAACAATTCACAGTCTAAATCTTTTCGCACTTGATCGAATATTTCTTTAAAAATGGCAACCTGAGCCATTGGTAAAA
CGGTCCTGCTGGCATTCTGGAGGGAAATACAACCGACAGATGTATGTAAGGCCAACGTGCTCAAATCTTCATACAGAAAGATTTGAAGTAATATTTTAAACCGCTATAGATGAAGAGCAGAAAGCGCA
AGCATTAGAGCAATTGTGAGGCAGCGTTGGTGAAGCAGGATAATAATATGAAGGATTATT-CCTGGTGGTTGACTGATCACCATAACTTTGCTAATCATTCAAACCTATTTGGTCTGTGACAGAGCC
TGTATGCTCTCTTTTCTGACGTTGGTCTCCGACGGCAGGCT--AATGACCCAGGCTGAGAAATTCGGACCCTTTTTTGTCAAGAGCGATGTTAATTTGTTCAATCATTGGTTTAGGAAAGCGGA
ATTATTTGACGTGGTTTGTATGGCCTCCACGCACGTTGTGATATGTAGATTGATAATCATTATCACTTTACGGGTCTTTCCGGTGAAAAAAGGTACCAAAAAACATCGTC
s DNA_CS                               1072 2465 + 3560 TAACGCCATTACCTACAAA-GCCAGCGCGACAAAATGCCAGAGAAGCTGAAGCTGGCGAACGCGGCAATT
TGTCAGTGCAGTGCAGTGAAGCCACCACCGCCTCCGGCGTGGATAATGCAGCCTCCCCCGACTGGCAGACACCGCTGAACGGGATTATTTACCCTCAGAGAGAGGCTGATCACTATGCAAAAAACAC
TTACGAATGTTTGTGGGTTTCTGTTTTAAACAACATTTTCTGCGCCGCCACAAATTTT-GGCTGCATCGACAGTTTTCTTCTGCCAATTTCCAGA--AACGAAGAAATGATGGGTGATGGTTTCCTT
GTAGAAAATTAACAAACCCTAAACAATGAGTTGAAATTTTATATTGTTAATATTTATTAATGTATGTCAGGTGCGATGAATCGTCATT-GTATTCCCGGATTAACATATGTCCACAGCCCTGACGGG
TGTTCTGAATGCTCTCAGTAAATAGTAATGAATTATCAAAGGTATAGTAATATCTTTTATGTTTATGTTGATGTTGTAACCCATCGGAAAACCTCCTGCTTTAGCAAGATTTTCCCTGTATTGCTGAAA
GCTAGATAGTAAATATAATGTGAGACGTTGTGACGTTTTAGTTTCAAGATAAAACAATTCACAGTCTAAATCTTTTCGCACTTGATCGAATATTTCTTTAAAAATGGCAACCTGAGCCATTGGTAAAA
CGGTCCTGCTGGCATTCTGGAGGGAAATACAACCGACAGATGTATGTAAGGCCAACGTGCTCAAATCTTCATACAGAAAGATTTGAAGTAATATTTTAAACCGCTA-----GATGAAGAGCAAGCGCA
AGCATTAGAGCAAT--TGAGGCAGCGTTGGTGAAGCAGGATAATAATATGAAGGATTATTCCCTGGTGGTTGACTGATCACCATAAC--TGCTAATCATTCAAACCTATTTAGTCTGTGACAGAGCC
TATATGCTCTCTTTTCTGACGTTAGTCTCCGACGGCAGGCTTCAATGACCCAGGCTGAGAAATTCGGACCCTTTTTGCTCAAGAGCGATGTTAATTTGTTCAATCATTGG-TTAGGAAAGCGGA
ATTATTTGACGTGGTTTGTATGGCCTCCACGCACGTTGTGATATGTAGA-TGATAATCATTATCACTTTACGGGTCTTTCCGGTGAAAAAAGGTACCAAAAAAACATC
```


Summary: files' formats

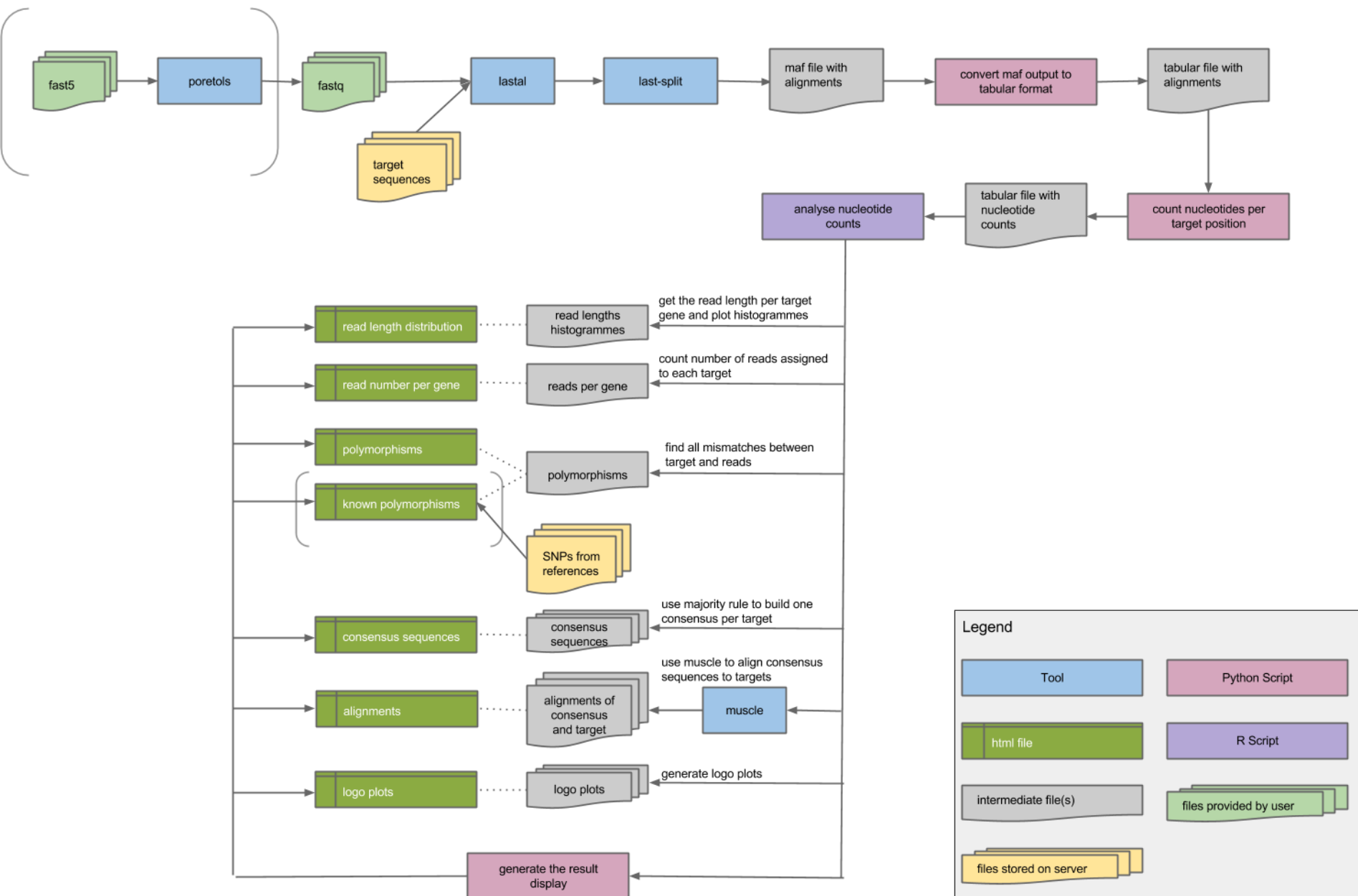
Files' formats used for storing sequences, sequencing results:

- `.fast5` – big, binary (cant read), contain a lot of metadata
- `.fastq` – readable by human, contains sequences and sequence quality
- `.fasta` – readable, contains sequences

Files' formats used for storing results of sequence alignment:

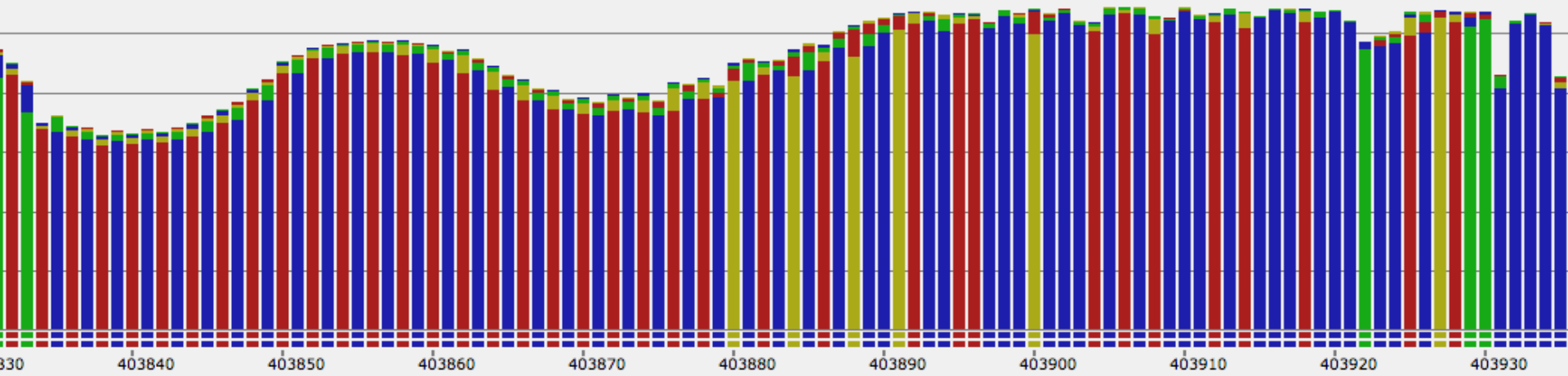
- `.maf` – contains pairs of aligned sequences with the alignment's coordinates; for example, used by LAST
 - `.bam` – binary format, includes aligned sequences, coordinates, information about bioinformatics processing, quality, ... Was created as a compact version of .bam file to save space
 - `.sam` – human readable version of .bam, much bigger in size
- `.fastq/.fasta` and `.bam` files are widely used in all DNA/RNA bioinformatics analysis

Nanopipe's workflow

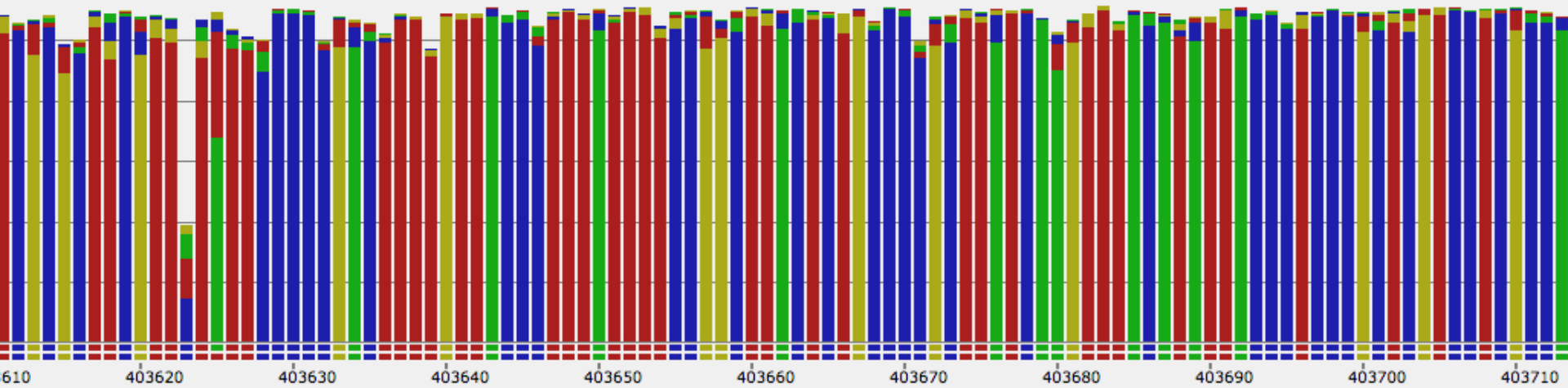


What is Consensus Sequence?

Pf3D7_07_v3:403089-404828:+:PfCRT_1



Pf3D7_07_v3:403089-404828:+:PfCRT_1



Key steps of Nanopipe

2) Tracing mutations (nucleotide frequency at a particular position) – majority rule, i.e. If alternate nucleotide is $\geq 50\%$ (?)

3) Generating consensus sequence and comparing it to the target sequence – MUSCLE aligner (using IUPAC symbols in ambiguous situation, for example, “R” means “A or G”, “Y” – for “C or T” (see <http://www.bioinformatics.org/sms/iupac.html> or on Nanopipe help-page: <http://bioinformatics.uni-muenster.de/tools/nanopipe/usage/>)

Nanopipe helps us to...

- See if our sequencing worked
- Detect insertions/deletions and nucleotide variations
- Visualization of the experiment
- We have the consensus sequence of our sample

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- Detect insertions/deletions and nucleotide variations
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We can see how our reads map to the target

IGV-viewer software uses .bam files and .fasta file of the target sequence
IGV-viewer is easy to install and use

Let's install IGVview and SEE the results of our Minlon sequencing 😊

- 1) Go to <https://software.broadinstitute.org/software/igv/download>
- 2) Click “Download Windows Package”
- 3) “unzip” the downloaded folder then open it, open next folder and click “igv” icon.
- 4) Wait 1 minute, do nothing 😊
- 5) Voila

*The link to the .fasta files with the target sequences:

http://bioinformatics.uni-muenster.de/share/Bangkok_2017

Tools from Oxford Nanopore

Metagenomics analysis: identifying different taxa in mixed samples (soil, food, human mucus,...)

EPI2ME

Bioinformatics cluster
(amazon cloud based)

→ **WIMP** (“what is in my pot”) : prokaryotes, fungi, archaea

→ **S16** : eukaryotes, prokaryotes, fungi, archaea

Find the PDF with the presentations here:

<http://www.compgen.uni-muenster.de/home/presentations/>

Practical session

Go to <http://bioinformatics.uni-muenster.de/tools/nanopipe/>

and let it begin

Reads quality

*a hint: when you finally obtain your sequencing reads it is important to know their quality. One of the tools (simple and good 😊) is FASTQC.

Download FASTQC: <http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>